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## **LDMS for the Web - User Manual**

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#### **Summary**

This manual explains how to use LDMS

## **Contents**

| Getting started     |  | 11              |
|---------------------|--|-----------------|
|                     | ence                                       |                 |
| About LDMS          |  | 11              |
|                     | mation                                     |                 |
| History             |  | 12              |
| Windows and         | d web versions                             | 12              |
| Migrating from Wir  | ndows to web versions                      | 12              |
| System requireme    | nts  | 13              |
| Signing in to LDMS  | S  | 14              |
| Acknowledgement     | of training                                | 14              |
| Changing users      |  | 16              |
|                     | nen automatically signed out               |                 |
|                     |  |                 |
|                     | erface                                     |                 |
| Applying filters to | a page                                     | 18              |
|                     | ters Code Searching                        |                 |
|                     |  |                 |
|                     | ntifiers to Select Specimens               |                 |
| Challenge codes     |  | 22              |
|                     |  |                 |
| Snecimen manage     | ment                                       | 23              |
| Terms used in LDN   | 1S   | <b>23</b><br>24 |
|                     | n Management page                          |                 |
|                     | information (Overview)                     |                 |
|                     |  |                 |
|                     | lentifiers                                 |                 |
|                     | cipants using the Specimen Management page |                 |
|                     | cipants using Quick Add                    |                 |
|                     | participant                                |                 |
|                     |  |                 |
|                     | Iments using the Specimen Management page  |                 |
|                     | rollments                                  |                 |
| Visits              |  | 32              |
| Visit identifie     | ers  | 33              |
| Adding a visi       | it to an enrollment                        | 33              |
|                     | sits                                       |                 |
| Primary specimens   | · · · · · · · · · · · · · · · · · · ·      | 35              |
| Specimen ide        | entifiers                                  | 35              |
| Specimen ide        | entifiers in LDMS for Windows              | 36              |
|                     | mary specimen to a visit                   |                 |
| Co-enrolling        | a primary specimen                         | 38              |
| Modifying a         | primary specimen                           |                 |
| Aliquot specimens   |  | 41              |

|      | Adding aliquots to a primary specimen             |    |
|------|---|----|
|      | Entering cryopreservation information             |    |
|      | Modifying aliquot specimens                       |    |
|      | Creating Subaliquots                              |    |
|      | Delete a specimen                                 |    |
|      | Removing specimens from storage                   |    |
|      | Quick add   |    |
|      | Adding specimens using Quick Add                  |    |
|      | Adding participants using Quick Add               |    |
|      | Adding enrollments using Quick Add                |    |
|      | TBD in quick add templates                        |    |
|      | Quick add templates                               |    |
|      | Differences between primaries and aliquots        |    |
|      | Specimen availability                             |    |
|      | Specimen record details                           |    |
|      | Specimen conditions                               |    |
|      | Destroying a specimen                             |    |
|      | Procedural conditions                             |    |
|      | Specimen container conditions                     |    |
|      | Temperature conditions                            |    |
|      | Shipping conditions                               |    |
|      | Qualitative conditions                            |    |
|      | Quantitative conditions                           |    |
|      | Other conditions                                  |    |
|      | Test assignment                                   |    |
|      | Assigning tests to a specimen                     |    |
|      | Indicating that a test will not be performed      |    |
|      | Deleting a test assignment                        | 03 |
|      |   |    |
| Stor | age6  | 54 |
|      | Navigation on the <b>Storage</b> page             | 64 |
|      | The storage tree                                  |    |
|      | Storage items                                     |    |
|      | Icons and their meaning                           |    |
|      | Assigning storage locations                       |    |
|      | Adding a new storage unit                         |    |
|      | Adding a new level in a storage unit              | 72 |
|      | Adding a container to a level                     |    |
|      | Assigning a storage location to specimens         | 75 |
|      | Generating a printable list of what is in storage |    |
|      | Container report                                  | 77 |
|      | Storage templates                                 |    |
|      | Creating storage item templates                   | 78 |
|      | Modifying and removing storage templates          | 80 |
|      | Moving items in storage                           |    |
|      | Moving Items in Storage - Filter Search           | 81 |
|      | Consolidating storage containers                  |    |
|      | Listing specimens that moved in storage           |    |
|      | Removing items from storage                       | 84 |
|      | Modifying the condition of stored specimens       | 85 |

|      | Storage Action Report8                                   |
|------|--|
| Sh'  | ipping8  |
| 3111 |  |
|      | LDMS shipping file compatibility8                        |
|      | Shipments are data transfer8                             |
|      | Shipment numbers8  |
|      | New shipments  |
|      | Creating new shipments 8                                 |
|      | Sending a pending shipment9                              |
|      | Generating a shipping manifest 9                         |
|      | Shipment Tracking Information 9                          |
|      | Supported shipping file formats 9                        |
|      | CSV shipping files9                                      |
|      | Cross-LIMS shipping files9                               |
|      | SeraCare shipping files 10                               |
|      | Shipping container report10                              |
|      | Shipment storage report                                  |
|      | Receiving shipments                                      |
|      | Shipping and projects                                    |
|      | Import as is   |
|      | Handling non-LDMS shipping files10                       |
|      | Shipment QA/QC   |
|      | Performing QA/QC   |
|      | Shipment history   |
|      | ·  |
|      | Removing a received shipment                             |
|      |  |
|      | Re-downloading the shipping file of a sent shipment      |
|      | Bulk Updating Condition Code and Comments of Shipments11 |
|      | Shipment Evaluations                                     |
| Re   | ports11  |
|      | Custom Report Builder11                                  |
|      | Creating a custom report11                               |
|      | Running a saved report11                                 |
|      | Modifying or deleting a saved report11                   |
|      | Example custom report11                                  |
|      | Generating a report                                      |
|      |  |
|      | Filter Reports Using a List of Specimens                 |
|      | Available report formats                                 |
|      | Administrative reports                                   |
|      | Anonymous Patients Map                                   |
|      | Transaction Log Report 12                                |
|      | User Event Report12                                      |
|      | User permissions report12                                |
|      | Barcode Reports12  |
|      | Barcoac Reports  |
|      | 1D Barcodes Report12                                     |
|      | 1D Barcodes Report12                                     |
|      |  |

| Abbott SARS-COV-2 Quant Export Report with Comments123      |
|---|
| Abbott SARS-COV-2 Quant Export Report with Sample           |
| Location  |
| Aliquot count by primary report                             |
| Aliquot Inventory Report                                    |
| CFAR export report126                                       |
| CFAR storage report   |
| Database dump report126                                     |
| Database dump report 2                                      |
| Exportable Abbott Assay Report                              |
| Laboratory 081 billing report                               |
| Laboratory 81 CNICS by date report129                       |
| Laboratory 081 CNICS general report                         |
| Laboratory 188 storage report                               |
| Lab 40 LabKey report  |
| Laboratory 48 billing report                                |
|   |
| Lab 485 aliquot report                                      |
| Lab 485 specimen count report                               |
| Lab 485 specimen count with volume report                   |
| Primary specimen database dump report                       |
| Sample counts for specified project report                  |
| Specimen export report                                      |
| Storage export report                                       |
| Westat PK Export Report                                     |
| Miscellaneous reports                                       |
| Clinic contact report                                       |
| LDMS abbreviated codes report136                            |
| LDMS assay censor codes report136                           |
| LDMS primary, additive, derivative, sub additive/derivative |
| codes report136   |
| MWCCS   |
| MWCCS Processing Report136                                  |
| Participant reports   |
| Participant identifiers report137                           |
| PK reports  |
| Pharmacology Drug Count137                                  |
| Pharmacology Drug List                                      |
| Pharmacology Proficiency Results                            |
| PK Drug Limits By Run                                       |
| PK Participant Report138                                    |
| PK Summary Report139  |
| PK Summary with Assay Name                                  |
| PK Summary with Assay Name (exportable)                     |
| Quick Add Templates   |
| Quick Add Template List Report                              |
| RPID Reports141   |
| Random PID Report141  |
| RPID Specimen Request Report141                             |
| Shipping reports  |
| Daily imported specimen log report141                       |
| Detailed imported specimen report 142                       |

|     | Imported specimen report - summary           | .142       |
|-----|--|------------|
|     | Lab 263 Summary Detail of Shipped Specimens  |            |
|     | Shipped specimen report - detail report      | 143        |
|     | Shipped specimen report - summary report     | 144        |
|     | Shipping laboratory contact report           | .144       |
|     | Specimens Marked for Shipping                | 144        |
|     | Specimen reports                             | 145        |
|     | Cell yield QA/QC summary report              | .145       |
|     | Lab 263 Processing Report                    |            |
|     | Lab 263 Summary of Specimens                 |            |
|     | Primary specimens received report            |            |
|     | Specimen count report                        |            |
|     | Specimen log report                          |            |
|     | Specimen processing report                   |            |
|     | Specimens for a given project report         |            |
|     | Specimens for a given project 2              |            |
|     | Time to freeze QA/QC summary report          |            |
|     | Time to process QA/QC summary report         |            |
|     | Storage reports                              |            |
|     | Specimens in storage per PID report          |            |
|     | Specimens not in storage report              |            |
|     | Specimens remaining in storage report        |            |
|     | Storage container location report            |            |
|     | Storage detail report                        |            |
|     | Test Result Reports                          |            |
|     | Abbott Realtime HIV1 Assay Report            |            |
|     | Abbott Realtime HIV1 Patient Report          |            |
|     | Abbott Repeat and Censored Run/Samples       |            |
|     | Abbott SARS-COV-2 Quant Assay Report         |            |
|     | Abbott SARS-COV-2 Quant Patient Report       |            |
|     | IQA Cryopreservation Patient Report          |            |
|     | TagMan HCV Repeat and Censored Run/Samples   |            |
|     | TagMan HIV-1 Repeat and Censored Run Samples |            |
|     | TaqMan Realtime HCV Assay Report             | .157       |
|     | TaqMan Realtime HCV Patient Report           | 157        |
|     | TaqMan Realtime HIV-1 Assay Report           | 158        |
|     | TaqMan Realtime HIV-1 Patient Report         |            |
|     | Taqman Realtime Qual Assay Report            |            |
|     | TaqMan Realtime Qual Patient Report          |            |
|     | WIHS   |            |
|     | Processing Log - Lab 263                     | 160        |
|     |  |            |
| Lab | els 1  | <b>L60</b> |
|     | Label formats                                | 160        |
|     | Barcodes                                     | 161        |
|     | Label sizes                                  |            |
|     | How to use a barcode reader with LDMS        |            |
|     | Printing labels anywhere                     | 162        |
|     | Support for Labelscape-generated Barcodes    | 163        |
|     |  |            |

| Generating labels                                 |     |
|---|-----|
| Defining new label formats                        | 165 |
| Customizing label formats                         |     |
| Setting the default label format for a project    | 167 |
|   |     |
| Test Results                                      | 167 |
| Test Results                                      |     |
| Test Supported in LDMS                            |     |
| Running an assay means reading data               |     |
| Test Results page                                 |     |
| Locating assay runs                               |     |
| Censor Codes                                      |     |
| Run Statuses                                      |     |
| Creating New Test Result Runs                     |     |
| Assigning Results to Blinded Controls (Pellets)   |     |
| Reviewing a Run                                   |     |
| Test History                                      |     |
| Deleting assay runs                               |     |
| Restoring deleted runs                            |     |
| Test result reports                               |     |
| Entering PK Test Results                          |     |
| Completing PK Test Results                        |     |
| Reviewing a PK Test Run                           |     |
| PK Control Lots                                   |     |
| Adding a New Lot                                  |     |
| Editing or Deleting a Lot                         |     |
| PK Control Charting                               |     |
| Generating a Report                               |     |
| PK Templates                                      |     |
| Adding a New Template                             |     |
| Updating an Existing Template                     |     |
|   |     |
| Data Submission                                   | 19/ |
| Adding Submissions                                |     |
| Editing Submissions                               |     |
| Data Submission Filters                           |     |
| Data Submission Filters                           | 100 |
|   |     |
| Printers  |     |
| Setting up the Brady IP300 printer                | 186 |
| Setting up the Brady® MVP 300 printer             |     |
| Setting up the LabXpert printer                   |     |
| Setting up the Zebra <sup>®</sup> GX 430t printer |     |
| Setting up the Brady BBP11-34L printer            | 190 |
| Setting up the Brady® BBP33 printer               |     |
| Setting up the Brady BMP53 printer                |     |
| Setting up the Brady BP-PR 300 printer            |     |
| Setting up the Zebra ZD620 Printer                |     |
| Setting up other printers                         | 193 |
|   |     |

| Preset projects | 194 |
|-----------------|-----|
| ACTIV           | 194 |
| ACTG/IMPAACT    | 195 |
| AERAS           |     |
| AIEDRP          | 196 |
| AIS             |     |
| AMC             |     |
| ATN             |     |
| BHP             |     |
| BM              |     |
| Botswana MOH    |     |
| CEMALB          |     |
| CHAVI           |     |
|                 |     |
| CIPRA-HT        |     |
| CIPRA-ZA        |     |
| CIPRA           |     |
| CONTROL         |     |
| CoVPN           |     |
| CPCRA           |     |
| CPQA            |     |
| CP-CTNET        |     |
| FACTS           |     |
| HN              |     |
| HPTN            |     |
| IDCRC           |     |
| IPREX           |     |
| IQA             |     |
| IRC             |     |
| KENPHIA2        |     |
| MACS            |     |
| MATRIX          |     |
| MAVRC           |     |
| MOSAIC          |     |
| mStudy          |     |
| MTCT            |     |
| MTN             |     |
| MWCCS           |     |
| NICHD-Westat    |     |
| PHACS           | 212 |
| PHIA            | 213 |
| REPRIEVE        | 214 |
| SHIMS           | 214 |
| SNRP            | 215 |
| TIES            | 215 |
| VQA             |     |
| VTN             |     |
| WHIN            |     |
| WIHS            |     |
| WITS            | 218 |

| ZIP  | 219  |
|--|--|
| Δ1Γ  | 220  |
| ministration   | 220  |
| Projects   | 221  |
| Local projects   | 221  |
| Government projects  | 224  |
| Adding a Reagent   | 224  |
| Adding Reagent Information   | 225  |
| Linking Reagent to Sample in Specimen Management   | 226  |
| Linking Reagent to Sample in Quick Add   | 227  |
| Linking Additive to Reagent Lot  |  |
| Reagent Lot Sample Details Report  |  |
| Specimen Anonymization   |  |
| Lab Settings   |  |
| User Settings  |  |
| User management  |  |
| Changing your password   |  |
| Password requirements  |  |
| Dropdown Customization   |  |
| RPID Requests  |  |
| Adding a New RPID Request  |  |
| Editing an RPID Request  |  |
| Clear Filters  |  |
|  |  |
| line Resources   | 233  |
|  |  |
| des, units, and abbreviations  | 234  |
| des, units, and abbreviations  Specimen type codes   | <b>234</b><br>234  |
| des, units, and abbreviations  Specimen type codes  Primary codes  | <b>234</b><br>234<br>234   |
| des, units, and abbreviations  Specimen type codes  Primary codes  | 234<br>234<br>234<br>241   |
| des, units, and abbreviations  Specimen type codes  Primary codes  Additive codes  Derivative codes  | 234<br>234<br>234<br>241   |
| des, units, and abbreviations  Specimen type codes  Primary codes  Additive codes  Derivative codes  Sub additive/derivative codes   | 234<br>234<br>234<br>241<br>251                                    |
| des, units, and abbreviations  Specimen type codes  Primary codes  Additive codes  Derivative codes  Sub additive/derivative codes.  Condition codes.  | 234<br>234<br>241<br>244<br>251                                    |
| des, units, and abbreviations  | 234<br>234<br>241<br>244<br>251<br>258<br>260                      |
| des, units, and abbreviations  | 234<br>234<br>241<br>244<br>251<br>258<br>260                      |
| des, units, and abbreviations  | 234<br>234<br>241<br>251<br>258<br>260<br>261                      |
| des, units, and abbreviations  Specimen type codes  Primary codes  Additive codes  Derivative codes  Sub additive/derivative codes  Condition codes  Unit codes  Measurement codes  Time unit codes  Visit codes   | 234<br>234<br>241<br>241<br>251<br>260<br>260<br>261               |
| des, units, and abbreviations  Specimen type codes  Primary codes  Additive codes  Derivative codes  Sub additive/derivative codes.  Condition codes  Unit codes  Measurement codes  Time unit codes  Visit codes  Assay Codes   | 234<br>234<br>241<br>244<br>251<br>260<br>260<br>261<br>263        |
| des, units, and abbreviations  Specimen type codes  Primary codes  Additive codes  Derivative codes  Sub additive/derivative codes  Condition codes  Unit codes  Measurement codes  Time unit codes  Visit codes  Assay Codes  Reasons for not running an assay censor codes                         | 234<br>234<br>241<br>244<br>251<br>260<br>260<br>261<br>263        |
| des, units, and abbreviations  Specimen type codes  Primary codes  Additive codes  Derivative codes  Sub additive/derivative codes  Condition codes  Unit codes  Measurement codes  Time unit codes  Visit codes  Assay Codes  Reasons for not running an assay censor codes  Immunology assay codes | 234<br>234<br>241<br>251<br>258<br>260<br>261<br>261<br>264<br>264 |
| des, units, and abbreviations  Specimen type codes  Primary codes  Additive codes  Derivative codes  Sub additive/derivative codes  Condition codes  Unit codes  Measurement codes  Time unit codes  Visit codes  Assay Codes  Reasons for not running an assay censor codes                         | 234<br>234<br>241<br>251<br>258<br>260<br>261<br>263<br>264<br>264 |

## **Getting started**

This manual describes how to use LDMS for Web and is intended for users at laboratories.

#### **About Frontier Science**

Frontier Science develops and maintains LDMS, and provides data hosting for laboratories.

Frontier Science and Technology Research Foundation, Inc. is a not-for profit that provides data management services to research organizations. LDMS is one of those data management services.

Many organizations that utilize LDMS also utilize Frontier Science's *data management center* (often referred to as the DMC) in Amherst, New York for long term data storage. These organizations use LDMS as a mechanism for sending specimen data for studies to Frontier Science.

Frontier Science is involved in many other statistical research efforts beyond LDMS. To learn more about the work that Frontier Science does, visit <a href="https://www.frontierscience.org">https://www.frontierscience.org</a>.

#### **About LDMS**

LDMS is a data management solution for laboratories managing biological medical specimens.

#### General information

LDMS is specifically designed for the needs of small and medium-sized research laboratories. It is available for Windows or as a web application. Laboratories generally use LDMS on one platform only. The Windows version must be installed on individual machines, but can utilize a centralized LDMS server installed at the laboratory. The web version does not require additional software to be installed and can be used in most web browsers. Users of the web version do not need to maintain a local database or create backups of data.

LDMS can perform many tasks, including the following:

- cataloging specimen information, such as its volume, additive, and current condition
- managing the transfer of specimen information between laboratories
- generating labels for specimens
- reading and entering assay result information
- keeping track of where specimens are located in storage units

Frontier Science manages data for laboratories using LDMS in a secure environment that meets the standards of 21 CFR Part 11 and NIST/FISMA guidelines.

#### **History**

LDMS was initially released for Windows in 1998 to collect data for HIV-related clinical trials.

The first projects to use LDMS were the Adult and Pediatric AIDS Clinical Trials Groups (AACTG and PACTG, now known as ACTG and IMPAACT). While LDMS continues to be used heavily by projects that focus on HIV and related cohorts, it is also used for many other specialized research purposes, including hepatitis C (HCV) and influenza studies, proficiency testing programs, and specimen repositories.

In 2014, Frontier Science released the web version LDMS.

#### Windows and web versions

Since the release of the web version of LDMS, Frontier Science has focused on the development of the web version.

The Windows version of LDMS continues to be maintained, and several updates to the Windows version have been released since the introduction of the web version. LDMS for Windows releases generally focus on improving existing features and project-specific changes. When possible, new laboratories should generally use the web version. When a new laboratory is set up, Frontier Science will guide the laboratory through the process of selecting and installing the appropriate version for their needs.

Laboratories are able to ship specimens to and receive specimens from other laboratories, regardless of which version of LDMS the laboratories are using. There may be other concerns when laboratories using the Windows and web versions need to work together. Frontier Science can work with laboratories to determine how to best handle different scenarios.

## Migrating from Windows to web versions

Laboratories currently using LDMS for Windows may be able to migrate to the web version.

Migration from the Windows to web version must be approved and coordinated by Frontier Science. Frontier Science will typically invite laboratories to migrate when it is appropriate to do so. Issues that need to be considered when migrating a laboratory include the following:

- Are all projects that the laboratory participates in supported in the web version?
- Are the features the laboratory is currently using supported in the web version?
- Does the laboratory routinely work with laboratories using the Windows version?

It is possible to migrate a laboratory's existing LDMS for Windows data to the web version. To do this, the laboratory's Windows database must be obtained by LDMS User Support. Frontier Science will then convert the data to the appropriate format and test it internally for issues. If the database can be converted successfully, LDMS User Support will help get the laboratory setup with appropriate user accounts and training.

## **System requirements**

These are the system requirements, along with recommended specifications, for using LDMS.

#### Required

**Table 1: System requirements** 

| Component         | Requirement   |
|-------------------|---|
| Web Browser       | <ul> <li>Firefox 54 or higher</li> <li>Chrome 58 or higher</li> <li>Edge 40 or higher</li> </ul>  |
|                   | JavaScript must be enabled for all browsers. Although most browsers will be able to use LDMS without issues, Frontier Science does not provide user support for web browsers that are not listed. |
| Screen Resolution | 1024 × 768  |
| Networking        | High-speed Internet connection  |
| Input             | Keyboard and 2-button mouse   |
| Video             | Monitor and video card capable of at least 1024 $\times$ 768 resolution and 16-bit color depth  |
| Label Printing    | Adobe Acrobat   |

#### Recommended

**Table 2: System recommendations** 

| Component      | Recommendation   |
|----------------|--|
| Printer        | Printers and barcode readers may have additional requirements. Consult your hardware's documentation for more information. |
| Barcode Reader | Compatible barcode scanner   |

#### Other software

LDMS can generate reports and other types of files in several different formats. Most of these files can be opened by commonly used programs that are already on our computer.

**PDF viewer**Needed to view and print reports. You can download many PDF viewers for free, such as Adobe Reader and

Evince. Many web browsers also have a built-in PDF viewer.



**Important:** An Adobe PDF viewer is required when printing labels.

## Spreadsheet viewer

Needed to view exportable reports. Microsoft Excel or LibreOffice Calc can open these files.

## Signing in to LDMS

Sign in to LDMS by using your user name and password, and selecting a laboratory database.

#### **Prerequisites**

You must have a user account before you can sign in to LDMS.

#### Steps

- **1.** In a web browser, go to *LDMS login*.
- **2.** In the upper-right corner of the page, click **LDMS Login**.
- **3.** Enter your user name and password, and then click the **Login** button. The **Select Laboratory** page will open.
- **4.** Select the laboratory from which you want to work from the **Select Lab[oratory]** box.

You will be working out of the selected laboratory's database for your session. You will only be able to select a laboratory to which you have access.

#### After you are finished

If you want to change to a different laboratory's database after signing in, click the **Change** link next to the laboratory name in the upper-right corner. To assign a default laboratory, hover over the **Administration** button and click **User Settings**. Click the drop-down box next to **Default Lab ID** and select the laboratory that you would like to make the default.

## Acknowledgement of training

If it is your first time logging into LDMS, you will be prompted to sign an acknowledgement of training after entering your username and password.

Figure 1: Acknowledgment Form



Please be advised that Frontier Science will require that all users be trained in the LDMS prior to using the system in accordance with 21 CFR Part 11. Acknowledgement of this training will be maintained electronically at Frontier Science.

Please use the acknowledgment prompt below to indicate whether you, as an LDMS user, have been trained in the use of the LDMS. Training can include but is not limited to in person training from FSTRF or other qualified laboratory staff proficient in the use of LDMS, attending a webinar overview, reading the user manual or viewing other training materials available on the LDMS website (www.ldms.org).

For those users who indicate "no," or otherwise indicate that they have NOT received LDMS training, please be advised you will not be allowed into the program. All training materials are published on the LDMS website (<a href="www.ldms.org">www.ldms.org</a>) and are available for you to complete this requirement. Please contact LDMS User Support with any questions or concerns regarding this requirement or access into the LDMS.

| <ul><li>No, I have not been trained</li></ul> |        |  |
|---|--------|--|
| <ul><li>Yes, I have been trained</li></ul>    |        |  |
| Login ID                                      |        |  |
| Password                                      |        |  |
|   |        |  |
|   | Submit |  |

If you have not been trained, you will not be allowed into the system. By selecting the option **Yes, I have been trained**, you are indicating that you have received training in the use of LDMS.

## **Changing users**

To switch users from the same web browser, you must log out and log in as a different user.

#### **Background**

Although multiple users can sign in and access data from one laboratory's database at the same time, you cannot sign in with multiple users from the same web browser on the same computer. For example, you can sign in as two different users on two different computers using Firefox, but you cannot open a new tab on the same computer and sign in as a different user. Doing so would cause problems with the tab in which you were originally signed in.

#### Steps

- 1. In the upper-left corner of any page in LDMS, click the **Logout** button.
- **2.** Sign in as a different user.

## Saving changes when automatically signed out

It is possible to save changes, even when automatically signed out of an idle session.

#### **Background**

You will automatically be signed out of LDMS if you leave a page idle for 15 minutes. When this happens, you will receive a notification that you have been signed out. If you attempt to leave the current page, you will be directed back to the sign in page. If you had unsaved changes when you were automatically signed out, those changes will be lost when you are redirected to the sign in page. This is to prevent unauthorized users from saving changes from a session that is signed out.

It is possible to save unsaved changes on a page after being logged out by signing in using another tab in the same web browser.

#### **Steps**

- **1.** Without leaving the page with unsaved changes, open a new tab in your web browser.
- **2.** In the new tab, go to the LDMS sign in page.
- **3.** Sign in as the user with the unsaved changes.
- **4.** Close the new tab.

#### Result

The original tab with the unsaved changes will now be considered signed in. You will now be permitted to continue working and to save any changes made.

## **Downloading files**

Different web browsers have different way of handling file downloads.

Many modern web browsers are configured by default to try to open certain types of files automatically or download them to a default location. For example, by default Firefox will attempt to open PDF files using a built-in PDF viewer. Likewise, Chrome is configured by default to save file downloads to a default location without asking you where to save them.

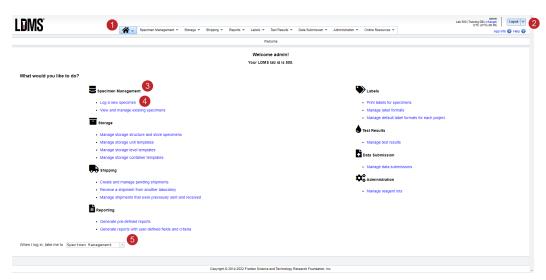
There are many times that LDMS will provide a file for you to download, such as a shipping file. Since you need to know where the file is on your computer in order to provide it to another laboratory, it is helpful to understand how your web browser handles file downloads.

How to view and modify your browser's behavior for downloading files or opening files automatically will vary by browser and version. For instructions, consult the help documentation for your web browser.

#### The LDMS user interface

When you first log in, you will be brought to the LDMS Welcome Page unless you have set another page as your default login page.

Figure 2: The LDMS Welcome page



(1) The LDMS menu bar, (2) The action menu, (3) LDMS Module Section, (4) Module task quick-link, (5) Default login page setting

**LDMS menu bar** The menu bar is used to navigate between different tasks

in LDMS. Hover the mouse pointer over a page to see its

sub-pages.

**action menu** The action menu is used in various areas of LDMS to

perform a major task. For example, on the **Specimen** 

**Management** page, the action menu is used to create a new participant.

**signing out** To sign out of LDMS as the current user, click the **Logout** 

button in the upper-right corner. This button is available  $% \left( 1\right) =\left( 1\right) \left( 1\right) \left$ 

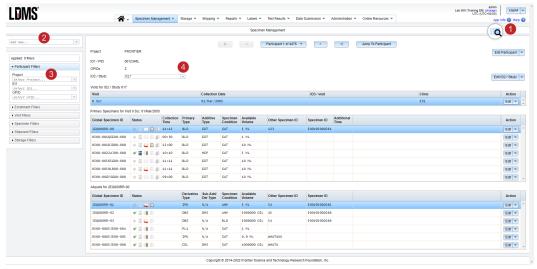
on every page.

change databases To change to a different laboratory database, click the **Change** link in the upper-right corner next to the

laboratory name.

The LDMS user interface has many common elements across different pages.

Figure 3: The main LDMS user interface



(1) Barcode scanning available (2) An applied filter, (3) Currently viewed participant, (4) Specimens

#### filters

Filters are available on most pages and will limit the information that is displayed in the working area on the page. For example, if you are on the **Specimen**Management page and apply a filter with the value FRONTIER to the **Project** field, only participants enrolled in the FRONTIER project will be displayed. The filters that are available will vary, depending on the current page. To remove a filter after it has been applied, click on it.

## Applying filters to a page

Filters are available on several pages and are used to narrow down the information that is displayed so that you can find something.

#### **Background**

On many pages in LDMS, all information for a particular task will be displayed together. For example, if you are on the Specimen Management page, all

participants in your laboratory's database will be displayed. If your laboratory has information for 200 participants, all 200 will be displayed by default and you would need to page through each one to find what you are looking for.

Filters can be used to temporarily limit the amount of participants shown so that only records of interest are displayed. This is the primary way to search for items in LDMS.

#### **Steps**

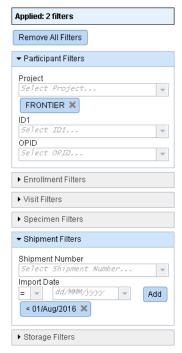
1. Open a page with filters, such as the **Storage** or **Specimen Management** pages.

Filters are displayed in the filters panel on the left side of the page, below the action menu.

**2.** Locate the desired filter.

Filters are grouped into categories, such as visit filters and specimen filters. Some categories will be specific to the current page. For example, the storage container filters will only be available on the **Storage** page.

Figure 4: The filters panel



In this example, the FRONTIER filter has already been applied to **Project**. This means that only information for that project will be displayed.

**3.** Select or enter the information to be shown from the filter box.

You can either select the information to filter from this list or enter it into the box. Filters will automatically show available options as you type.



**Note:** Filters do not affect other filters. For example, if you select the FRONTIER project, then try to apply a filter for **ID1**, all ID1 values in the laboratory's database will be available for selection, even if they don't below to the FRONTIER project.

#### Result

The filter will be applied immediately once it is selected and the page will automatically refresh to remove any records that do not meet the filter's criteria. If two filters were selected, they will both be applied using "and" logic. For example, if you selected FRONTIER from the **Project** filter and BLD from the **Primary type** filter, only blood specimens for the FRONTIER project will be displayed..

#### After you are finished

To remove a filter that has been applied, find it in the filter panel on the left side of the page and click on it. It will be removed immediately and the page will automatically be refreshed to reflect the change.

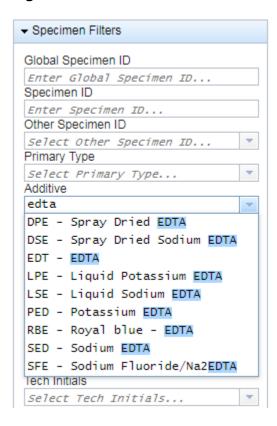
#### Specimen Filters Code Searching

If using **Specimen Filters** options, contextual suggestions will appear when typing in the following fields:

- Primary Type
- Additive
- Derivative Type
- Sub Add/Der Type
- Condition

While other filter fields will display results as you type, these specific fields provide additional aid if uncertain of the *code abbreviation*. If unsure of the code abbreviation for DSE - Spray Dried Sodium EDTA, for example, then typing **EDTA** in the **Additive** field yields a list of possible codes containing the **EDTA**.

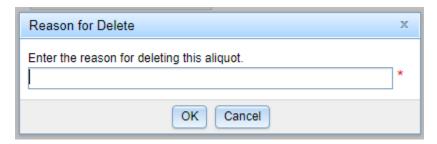
Figure 5: Contextual results



## **Deleting Items**

When a user deletes an item such as a specimen, storage item, shipment, or test run, the user will be required to specify why the item is being deleted. This reason is then displayed on the Transaction Log report to properly track the reason for deleting.

Figure 6: Reason for Delete Window



## Using a List of Identifiers to Select Specimens

You can use a list of identifiers to select a set of specimens

This feature is available in most places where specimens need to be selected including Custom Report Builder, Pending Shipments, Stored Samples, Test Results, and Print Labels using the **Upload Global Specimen Ids File** button. The file of identifiers must be a text file and must include one global specimen ID per line.

#### Figure 7: Global ID List File Example

```
0500-00ABCDE00-001
0500-00ABCDE00-002
0500-00FGHIJ00-001
0500-00FGHIJ00-002
0500-00FGHIJ00-003
```

## **Challenge codes**

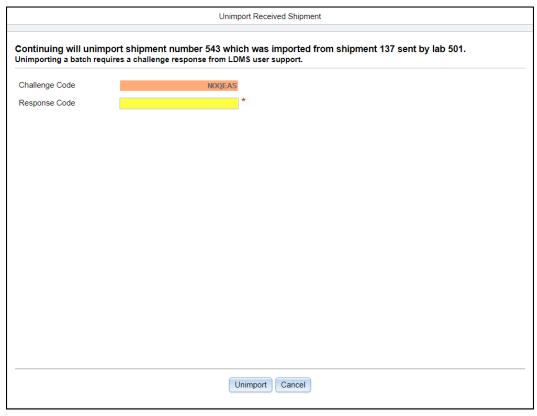
A challenge code is password that you must give to LDMS User Support to perform certain actions in LDMS.

Access to certain features in LDMS may be restricted for a number of reasons. For example, unimporting a received shipment can have very serious consequences. LDMS does not provide "undo" functionality to reverse such an action. A challenge code is a fail-safe mechanism that can prevent accidental changes to specimen data.

If you attempt to access a feature locked with a challenge code, LDMS will prompt you for the response code.

You must contact LDMS User Support and give them the **Challenge Code**. If appropriate, they will provide you with the response code which will then unlock the feature for your use. The code will change each time the feature is accessed, and will be needed even if the feature was previously unlocked.

Figure 8: Prompt for challenge password



In this example, you would provide LDMS User Support with the code NOQEAS and they will provide you with the Response Code to access this feature.



**Warning:** Do not close the challenge password window before you obtain the **Response Code**. Doing so will cause a new **Challenge Code** to be generated, which will require a different **Response Code**.

## Specimen management

The Specimen Management page is where you can manage the details of specimen records and related information for participant enrollments.

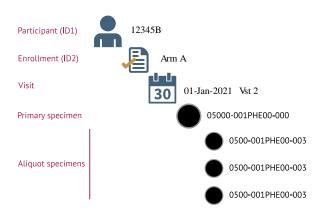
In LDMS, a participant, an enrollment, and specimens drawn from a participant (including the specimens derived from them) form a specimen record. Specimen records are added and displayed on the Specimen Management page.

#### Terms used in LDMS

Specimen records are stored as a hierarchy of related participant, enrollment, and information. All of this information for a participant-project combination comprise a record in LDMS.

A *specimen record* in LDMS is the collection of specimen, study, and visit information for a specific participant.

Figure 9: Contents of a specimen record



The top of a specimen record represents the largest part, such as a clinical trials network; the right side of this image is an example of each item

| project     | The organization that sponsors or conducts the study in which the participant is enrolled. |
|-------------|--|
| participant | The person (or specimen source if the project doesn't                                      |

work with human samples) that is participating in a given project. If a person is participating in more than one project, they will be considered two different participants in each project. Participants cannot be linked across

projects.

**enrollment** The protocol or study in which the participant is enrolled.

**visit** A specific event at which a primary specimen was

collected from a participant.

**primary** The specimen that was collected from the participant, such as an unprocessed tube of blood

**aliquot** A specimen that was created by processing a primary

specimen into a smaller specimen.

Each part of a specimen record is selected in sequence. For example, you must specify a project before enter participant information. Likewise, you must create the information for a visit event before you can add specimens that were drawn at that visit.

## **Using the Specimen Management page**

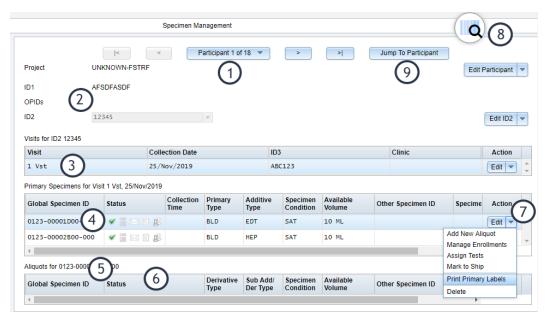
The **Specimen Management** page shows all of the visits for a specific participant and protocol combination.



**Important:** The labels for certain buttons and menus in LDMS vary based on the project. For one participant, for example, the unique participant identifier (ID) may be called "PID"; for another project it may be "PanelID".

The main work area of the Specimen Management page will display the all the visits and associated specimens for a participant. Use the navigation buttons to change between different participants.

Figure 10: The Specimen Management page



(1) Navigate between participants, (2) The current participant enrollment information, (3) Visits for the current participant, (4) Primary specimens for the currently selected visit, (5) Aliquot specimens derived from the currently selected primary specimen, (6) Specimen status indicators, (7) Actions for a primary (8) Indicator that you can scan a barcode to search for a record, (9) Jump directly to a participant

| Status Icon | Meaning                                 |
|-------------|---|
| €           | Specimen is available                   |
|             | Specimen is assigned a storage location |

| Status Icon  | Meaning   |
|--|---|
|  | Specimen has been marked to ship to another laboratory  |
|  | Specimen is on a pending shipment to another laboratory |
| <b>\(\rightarrow\)</b>   | Specimen has been shipped to another laboratory         |
| 8  | Specimen has an additional enrollment                   |
| is a second of the second of t | Specimen has an additional enrollment                   |

| Objective   | Action   |
|---|--|
| Change participants   | Click the navigation buttons at the top of the screen.   |
| View the visits for a different study that the participant is enrolled in | Select the study from the <b>Study</b> drop-down box. If the <b>Study</b> box is disabled, then the participant is only enrolled in one study. |
| See the primary specimens associated with a visit                         | Click a visit from the <b>Visits</b> section of the page.  |
| See the aliquots associated with a primary specimen                       | Click a primary in the <b>Primary Specimens</b> section.   |
| Search for a specific participant   | Use the filters on the left-side of the page.  |

## **Entering specimen information (Overview)**

Records are added to LDMS from the top-down, starting with the creation of a new participant.

#### **Background**

Because records in LDMS are hierarchical, you need to enter information starting with a participant, and then work your way down to aliquot specimens.

There are two ways to enter participant, enrollment, and specimen in LDMS:

- Using the Quick Add feature (recommended)
- Adding new participant, enrollment, and specimen information one-by-one

For most data entry in LDMS, the Quick Add feature is strongly recommended since it allows you to enter all of the information at once. Adding individual items on the **Specimen Management** page is recommended for minor changes, such as adding a single aliquot specimen to a primary specimen.

Each of the steps below can be completed independently, provided that the previous steps were completed. For example, you can create a new participant in LDMS without enrolling that participant in any protocols or creating any associated visits, however you cannot create a generic enrollment that is not associated with a specific participant visit. Likewise, it

is also not possible to create a primary specimen without associating it with a specimen participant's visit.

#### **Steps**

- **1.** Create a new participant.
  - Participants are specific to a project. If a person is participating in more than one project, then they would be considered two different participants in LDMS and would be entered separately.
- Add an enrollment to the participant.

  This means that you are associating the participant with a specific protocol. Participants can be associated with multiple protocols.
- **3.** Add a visit to the participant.
- **4.** Add a primary specimen to the visit.
- **5.** Add aliquots that were processed from the primary specimen.

## **Participants**

A participant is the source for a specimen entered into LDMS.

The participant is typically a person enrolled in a study. Similar terms that you may be familiar with include "patient" and "source". In LDMS, a participant refers to all of these concepts. This means that a participant does not necessarily need to be a person. For example, a "participant" may be a control sample from which a specimen was created.

In LDMS, a participant will be the first thing that you create when adding a new *specimen record*. Newly created participants must be associated with a project and an ID1 (or participant identifier).

If a person is taking part in more than one project, that person will be considered two different participants in LDMS.

### **Participant identifiers**

A participant identifier is a series of numbers and letters that uniquely identifies a participant within a specific project.

A participant identifier is called "ID1" in LDMS until you select a project for the participant. Each project will have a different name for ID1. For one project, it may be called "PID"; for another project, it may be called "PANEL".

## Adding participants using the Specimen Management page

A new participant can be created on the **Specimen Management** page or the **Quick Add** page. This section explains how to add them on the Specimen Management page.

#### **Background**

A new participant can be created two ways:

- If you want to create a new participant without any enrollments or visits, use the **Specimen Management** page.
- If you want to create a new participant allow with an associated visit and specimens, use the **Quick Add** feature.

The Quick Add feature requires you to enter at least one visit date with the participant. If the participant has not yet had a visit, you must use the Specimen Management page.

#### Steps

- **1.** From the LDMS menu bar, click **Specimen Management**.
- **2.** From the action menu in the top-left of the page, click **Add Participant**.

Figure 11: The action menu with Add Participant selected



**3.** From the **Project** box, select a project.

#### **Example**

Figure 12: The Create Participant page



The **ID1** box will change to appropriate name for the selected project.

- **4.** In the **ID1** box, enter the participant's identifier.
  - The label for the **ID1** box will vary by project. In the example above, ID1 is "PID".
- **5.** Optional: In the **OPIDs** box, enter an *other participant identifier*, and then click **Add OPID**.
  - More than one OPID can be added for the same participant. To remove an OPD that was added, click the **Delete** button to the right of it.
- **6.** Click **Save**.

If the project has validation checks for ID1, they will be run at this time. If the ID1 that you entered is not valid for the project, you will be prompted to correct it before you can save the new participant.

If the project and participant identifier combination already exists, you will be asked if you want to view that participant instead.

#### Result

The new participant will be created and added to the end of your laboratory's database. For example, if your laboratory previously had 250 participants, the new participant will be 251. LDMS will automatically open the record for the newly created participant so that you can add enrollment information.

#### Adding participants using Quick Add

The Quick Add feature can be used to add a new participant, along with other information, on a single page.

#### **Background**

If adding a participant in this way, at least one visit must have occurred as the date of the visit (the **Collection Date**) is required.

#### Steps

- From the LDMS menu bar, click Specimen Management > Quick Add.
- **2.** Optional: Select a template for the participant visit.
- **3.** Complete the information in the **Participant Information** section.
- **4.** In the **Visit Information** section, enter the **Collection Date**.
- **5.** At the bottom of the page, click **Add**.
- **6.** Optional: Complete any additional enrollment, visit, or specimen information for this participant.

#### Result

When the participant is successfully created, a note will appear at the top of the page. The participant will be available for select on the **Specimen Management** and **Quick Add** pages, where new enrollments, visits, and specimens can be added.

## Modifying a participant

Modify participants on the **Specimen Management** page.

#### **Background**

A participant may need to be modified to correct an error in the ID1 that was entered, or to add and remove OPIDs as needed.

#### **Steps**

- 1. Click **Specimen Management** from the LDMS menu.
- **2.** Find the participant to be modified. This can be done by following <u>one</u> of the two sets of steps below:

- Use filters from the left side of the screen to narrow down the participants that are displayed.
- Alternatively, click **Jump to Participant** and enter the appropriate project and participant ID, then click **Jump To**.
- **3.** With the participant open in the work area, click the **Edit Participant** button.

The **Edit Participant** window will open.

**4.** Modify any participant information as needed.

Note: Edits made here will change all specimens associated with the participant.

The **Enrollments** section of this window was not present when the participant was first created. It lists protocols in which the participant is enrolled, and provides a link to view and modify that enrollment, if needed.

**5.** Click the **Save** button.

If you modified the participant identifier, LDMS will check to ensure that it does not already exist for another participant.

#### **Enrollments**

An enrollment associates a participant with a protocol.

For example, a participant in the Frontier project may be participating in a study called F1526. The combination of that participant identifier on that study is called the *enrollment*.

A protocol is called ID2 in LDMS. Like ID1 for participant identifiers, the name for ID2 will vary, depending on the project. For example, a vaccine project may call the protocol box in LDMS "STUDY", whereas a pharmacology project may call it "TEST PANEL".

Also like ID1, a project may have rules for how its ID2 is formatted. For example, a vaccine project may require that protocols are in the format "STUDY 201", "STUDY 202", and so on; a pharmacology project may use "TEST PANEL A0003", "TEST PANEL A0004", etc. If the project has specified these rules, they will be enforced by LDMS.

A participant can be enrolled in more than one ID2 for a project, however because participants are specific to a project, they cannot be enrolled in protocols across multiple projects.

### Adding enrollments using the Specimen Management page

Enrollments are added to an existing participant, and associate that participant with a specific protocol.

**Prerequisites** 

The participant must already exist in LDMS before an enrollment can be added.

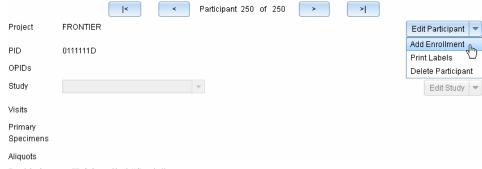
#### Steps

- **1.** From the LDMS menu bar, click **Specimen Management**.
- **2.** Find the participant.

Use filters from the left side of the page to narrow down the participants that are displayed.

**3.** From the **Edit Participant** menu, click **Add Enrollment**.

#### Figure 13: Adding a new enrollment to a participant



In this image, ID2 is called "Study"

The **Create Enrollment** window will open.

**4.** Select or enter an **ID2** in the box.

The label for this box will vary, depending on the project. It is most commonly called **Study** or **Protocol**.

5. Click Save.

#### Result

The participant is now enrolled in the protocol. You will see the protocol listed in the **ID2** box.

#### After you are finished

If the participant is enrolled in more than one protocol for a project, you can create additional enrollments in the same way.

### **Modifying enrollments**

Participant enrollments can be modified or removed after they are created on the **Specimen Management** page.

#### **Prerequisites**

- If removing an *enrollment* from a *participant*, the enrollment must not have any associated *visits*.
- If you accidentally added a visit and specimens to the enrollment in error and you want to preserve them, co-enroll the specimen(s) in another enrollment first so that the erroneous enrollment can be removed.

#### **Background**

The only property of an enrollment is the associated *ID2*. If the incorrect ID2 was selected when the enrollment was created, you can change it. Doing so will add the participant to a new enrollment, not change the existing. For example, if a participant is currently assigned an ID2 of STUDY3, and you change it to STUDY4, this change will only affect this participant, not all participants in STUDY3. If the enrollment was added in error, it can be deleted instead.



**Note:** Enrollments can also be modified using Quick Add.

#### **Steps**

- **1.** In the LDMS menu bar, click **Specimen Management**.
- 2. Find the participant with the enrollment to be modified.

  Use filters from the left side of the screen to narrow down the participants that are displayed.
- 3. Select the ID2 from the [ID2] box.
  The label for the [ID2] box will depend on the project. Common labels are "study" and "protocol".
- **4.** To modify the enrollment:
  - **4.1.** Click **Edit** [ID2], where "[Protocol]" is the appropriate label for the project.
  - **4.2.** Modify the enrollment as needed.
  - 4.3. Click Save.
- **5.** To remove the enrollment:
  - **5.1.** From the **Edit** [ID2] menu, click **Delete** [ID2].

Figure 14: The Edit Study menu



In this image, ID2 is "Study"

5.2. Click Delete.

#### **Visits**

A visit is a specimen collection event associated with an enrollment.

A visit typically involves the collection of specimens defined by the protocol. For example, a visit may specify that you collect 3 tubes of blood from the participant. In LDMS, you would add the visit to the participant, and then add the specimens to the visit.

#### **Visit identifiers**

A visit identifier defines the type of visit.

A visit is identified by two pieces of information: the *visit value* and *visit unit*. The visit value is a sequential identifier, such as a number. The visit unit is the type of *visit*.

For example, a visit of "3 WK" indicates that it is the third visit of the week type. The 3 is the visit value, and WK is the visit unit. The protocol in which the participant is enrolled likely has defined specimen collections for this visit.

The available visit units are defined by various projects that use LDMS. Users cannot create their own visit units.

#### Adding a visit to an enrollment

Visits are added to an enrollment on the **Specimen Management** page.

#### **Background**

A visit must be created before you can enter specimen information in LDMS.



**Tip:** If you want to create a new visit for a specimen that already exists, you should co-enroll the primary specimen instead of adding a new visit to the enrollment. During the co-enrollment process, you will have the opportunity to create a new visit.

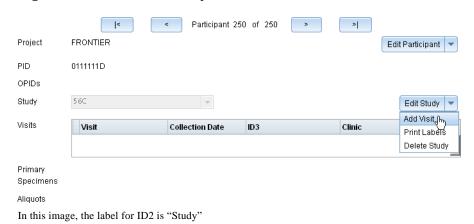
#### **Steps**

- 1. Click **Specimen Management** from the LDMS menu bar.
- 2. Find the participant to which a visit will be added.

  Use filters from the left side of the page to find the participant.
- 3. Select the [ID2] from the list.

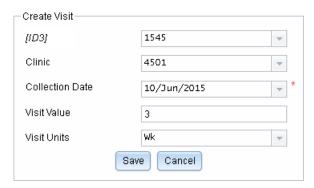
  The label for the [ID2] box will depend on the project. Common labels are "study" and "protocol".
- Click the down arrow next to the Edit [ID2] button, and then click Add Visit.

Figure 15: The Edit Study menu



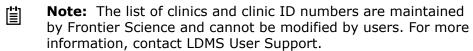
The **Add Visit** window will open.

Figure 16: The Add Visit window



**5.** Select the clinical site where the visit occurred from the **Clinic** list.

If you want to know the name of the site, hover over it with the mouse pointer.



- **6.** (If required by the project) Select the *ID3* from the **ID3** list. ID3 is typically used for a sub-study or sub-protocol.
- 7. Select the date that the visit occurred or will occur from the **Collection Date** list.

If the visit occurred or will occur over more than one day, you must add a new visit for each date.

- Note: Future dates are allowed in the Collection Date field.
- **8.** Enter the *visit value* and *visit unit* for the visit into the **Visit Value** boxes.

If you want to see what a particular visit unit means, hover over it with the mouse pointer.

**9.** Click the **Save** button.

#### Modifying visits

Visit information is modified or removed on the **Specimen Management** page.

#### Background

You may need to modify a visit after it is created to correct an entry error (for example, using the incorrect clinic). If the visit was added in error, you can also remove it.

#### **Steps**

- 1. Click Specimen Management > Available Specimens from the LDMS menu bar.
- 2. Find the participant with the visit to be modified. Use filters from the left side of the screen to narrow down the participants that are displayed.
- 3. Select the *protocol/ID2* associated with the visit from the **[Protocol]** box.

The label for the **[Protocol]** box will depend on the project. Common labels are "study" and "protocol".

- 4. To modify the visit:
  - **4.1.** Click the **Edit** button to the right of the visit. The **Edit Visit** window will open.
  - **4.2.** Modify the visit information as needed. If a primary specimen has already been entered for the visit, it will be listed along with the visit information.
  - **4.3.** Click the **Save** button.
- 5. To remove the visit, select **Delete** from the **Edit** menu to the right of the visit.

In order to remove the visit, there must be no primary specimens added to it. If you want to remove a visit without deleting the primary specimens, you can co-enroll the primary specimen. This will allow you to associate the specimen with another visit.

## **Primary specimens**

A primary specimen is a specimen that was collected from a participant during a visit.

Primary specimens are generally larger specimens that are later processed into smaller specimens called aliquots. New primary specimen are added to LDMS on the **Specimen Management** page. They are always associated with at least one participant visit.

## Specimen identifiers

All specimens in LDMS are assigned a unique specimen identifier.

There are three types of specimen identifiers that can appear in LDMS:

**global specimen** This is a unique identifier assigned to all specimens. ID specimen ID This identifier may or may not be unique. It is only

used by LDMS; LDMS will not assign a specimen ID to specimens added using LDMS.

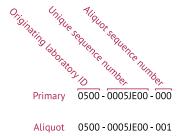
other specimen This identifier is intended for laboratories to use to assign ID their own, internal-use identifiers as needed.



**Important:** LDMS for Windows and LDMS use different formats for the global specimen ID. The format used by LDMS is longer, and contains the originating laboratory's ID. For example, the global specimen ID "0500-0005JE00-000" indicates that the specimen originated from LDMS laboratory 500.

The global specimen ID assigned by LDMS links a primary specimen to any aliquot specimens created from it. Primaries and aliquots derived from them will have the same global specimen ID up to the dash character. The last three numbers are the sequence number. A sequence number of "-000" indicates that the global specimen ID belongs to a primary specimen; any other number indicates that it belongs to an aliquot specimen.

Figure 17: The global specimen ID of primary and aliquot specimens



This image shows the relationship between a parent primary and aliquots derived from it.

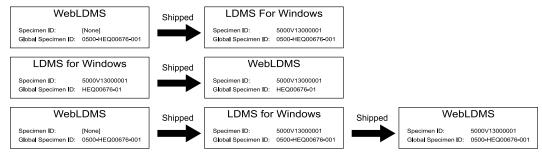
#### **Specimen identifiers in LDMS for Windows**

LDMS for Windows handles specimen identifiers in a few ways that differ from LDMS.

Understanding the differences between the way LDMS and LDMS for Windows assign specimen identifiers is important if your laboratory does work with other laboratories.

LDMS will not assign a *specimen ID* to a specimen and will only assign a *global specimen ID*. LDMS for Windows will assign both. The format of the global specimen ID differs as well, depending on which system assigned it.

Figure 18: Specimen ID and global specimen IDs interoperability



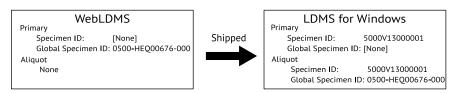
This image shows how specimen IDs and global specimen IDs are affected as specimens are shipped between LDMS and LDMS for Windows.

The global specimen ID of a primary specimen can be different. LDMS for Windows does not permit the shipping or storage of primary specimens;

these are features that are exclusive to LDMS. In LDMS for Windows, users would generally create aliquots (commonly called "ghost aliquots") that were identical to the primary specimen; the aliquot (which was really just the primary) could then be shipped and stored.

When a primary specimen is shipped to LDMS for Windows, an aliquot will automatically be created. This aliquot will have the global specimen ID of the primary specimen, and the primary specimen ID will have no global specimen ID.

Figure 19: Primary specimens shipped to LDMS for Windows



This image shows how a primary specimen without aliquots created in LDMS is converted to an aliquot if shipped to an LDMS for Windows laboratory.

In this situation, if the new aliquot is shipped to a laboratory that users LDMS, it will recognize that the global specimen ID belongs to a primary specimen (and not an aliquot), and will turn it back into a primary specimen.

### Adding a primary specimen to a visit

A new primary specimen is added to a visit on the **Specimen Management** page.

### **Background**

A primary specimen is added to an existing visit. The primary specimen can be added in LDMS even if it has not yet been processed into any aliquots.

#### **Steps**

Locate and select the visit

- 1. Click **Specimen Management** from the LDMS menu.
- 2. Find the participant to which the primary specimen will be added.

  Use filters from the left side of the screen to narrow down the participants that are displayed.
- 3. Select the ID2 associated with the visit from the [ID2] box. The label for the [ID2] box will depend on the project. Common labels are "study" and "protocol".

Add the new primary specimen

4. Click the down arrow next to the **Edit** button to the right of the visit to which the primary will be added, and then click **Add New Primary**.

Figure 20: Adding a new primary to a visit



The Create Specimen window will open.

- **5.** Fill in the information for the new primary specimen. Items marked with a red \* are required.
- **6.** Optional: Enter additional information about the primary specimens into the **Comments** box.
  - Information that is typically included here includes details for specimens that were not collected, an explanation of a condition, or an explanation of why a specimen is not available.
- 7. Optional: Enter additional information about the primary specimens into the **Internal-only comments** box. These comments will be include if the specimen's information is shipped to another laboratory. Comments entered here is for your laboratory's use only. These comments will not be included if the specimen's information is shipped to another laboratory.
- 8. At the bottom of the window, click **Save**.

  The information you entered will be checked for completeness and validity. If there is an issue with one of your entries, it will be highlighted with a brief explanation of how to correct it.

#### Result

A new primary specimen is now available. It will automatically be assigned a global specimen ID ending in "-00" by LDMS.

Note: The new primary will not have a specimen ID assigned to it. Specimen ID is only used by LDMS; LDMS uses the global specimen ID, not specimen ID, to identify specimens

### After you are finished

If the primary was collected for more than one protocol or project, you must also co-enroll the specimen so it is associated with each applicable visit.

### Co-enrolling a primary specimen

Co-enrolling is the process of associating a primary specimen with more than one visit or protocol.

### **Background**

A participant may be participating in two different protocols at the same time, and a single primary specimen might be collected to meet the needs of a visit from each protocol.

You can also co-enroll a primary specimen to move it between visits. This might happen, for example, if you selected the wrong visit for the primary specimen, and you want to move it to the correct visit rather than deleting it.



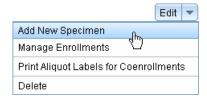
**Note:** If a specimen is co-enrolled in a local project and a government project, when shipping specimens for the government project, information about the co-enrollment will not be included in the shipment.

### **Steps**

- 1. In the menu bar, click **Specimen Management**.
- 2. Local the specimen to be co-enrolled.

  Use filters from the left side of the screen to narrow down the participants that are displayed.
- 3. Click the down arrow next to the **Edit** button to the right of the primary to be co-enrolled, and then click **Manage enrollments**.

Figure 21: The Edit button for a primary



The **Manage Enrollments** window will open.

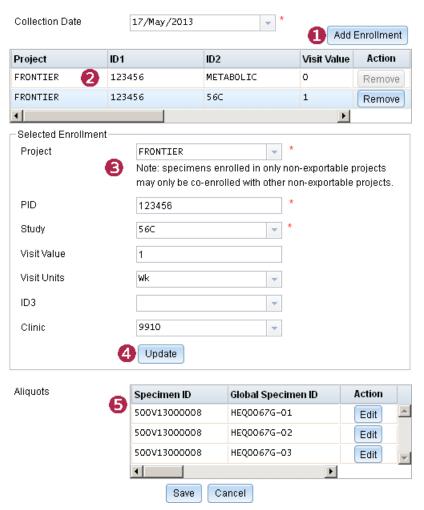


Figure 22: The Manage Enrollments window

(1) Add a blank enrollment row, (2) Enrollments for this primary specimen, (3) Details for currently selected enrollments, (4) Save changes to currently selected enrollment, (5) All aliquots derived from this primary, including all enrollments

- 4. Click the **Add Enrollment** button.
- **5.** Select the blank row created in the previous step by clicking on it.
- **6.** Fill in the information in the **Selected Enrollment** section for the new enrollment.
- **7.** Click **Update**.

The information for the enrollment will be updated in the list of enrollments.

**8.** At the bottom of the window, click **Save**.

# Modifying a primary specimen

A primary specimen can be modified on the Specimen Management page.

### **Background**

After a primary has been created, it may need to be modified for a variety of reasons, such as correcting a data entry error or updating its current condition.

### **Steps**

- 1. Click **Specimen Management** from the LDMS menu.
- 2. Find the participant with the primary specimen to be modified.

  Use filters from the left side of the screen to narrow down the participants that are displayed.
- **3.** Select the *protocol/ID2* associated with the *visit* from the **[Protocol]** box.
  - The label for the **[Protocol]** box will depend on the project. Common labels are "study" and "protocol".
- **4.** Click the **Edit** button to the right of the primary specimen to be modified.
  - The **Edit Specimen** window will open.
- 5. Modify the primary specimen as needed.
  - If any aliquot specimens were added for this primary, it will appear at the bottom of the window.
- **6.** Click the **Save** button.
  - If there are any issues with the changes that you made, such as an invalid entry, you will be prompted to correct it. An explanation will appear next to the information that needs to be corrected.

# **Aliquot specimens**

An aliquot is a specimen that is derived from a primary specimen

An *aliquot* is created by processing the specimen collected at a participant's visit into smaller specimens. For example, if 10 mL of blood was collected during a visit, that 10 mL tube is the *primary specimen*. It may be processed then into smaller, 1 mL tubes of double-spun plasma. These 1 mL specimens are the aliquots in LDMS.

# Adding aliquots to a primary specimen

An aliquot specimen is added to a primary specimen on the Specimen Management page.

### **Background**

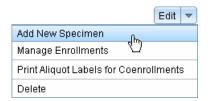
An aliquot is added to an existing primary.

#### Steps

**1.** In the menu bar, click **Specimen Management**.

- **2.** Find the participant by using filters on the left side of the screen.
- 3. Click the arrow next to the **Edit** button to the right of the primary to which the aliquots will be added, and then click **Add New Specimen**.

Figure 23: Adding a new aliquot to a primary specimen



The Create Specimen window will open.

 Enter the number of aliquots that were created into the Number of Aliquots box.

When entering more than one aliquot together, all aliquots should have the same basic properties (such as the same volume). If you want to add multiple aliquots with different information, you can still do so, but you must edit the aliquots later. For example, if you have 3 aliquots that are identical, but one has a different **specimen condition**, you can add all three together, and then change the condition of the one aliquot when you are finished.

- **5.** (If the primary is co-enrolled) Select the enrollment for the new aliquot from the **Enrollment** box.
  - An aliquot cannot be enrolled in more than one study.
- **6.** Fill in the information for the new aliquot specimens. Items marked with a red \* are required.
- **7.** Optional: Enter additional information about the aliquot into the **Comments** box.

Information that is typically included here includes details for specimens that were not collected, an explanation of a condition, or an explanation of why a specimen is not available.

**8.** Optional: Enter additional information about the aliquot into the **Internal-only comments** box.

Comments entered here are for your laboratory's use only. These comments will not be included if the specimen's information is shipped to another laboratory.

**9.** At the bottom of the window, click **Save**.

The information you entered will be checked for completeness and validity. If there is an issue, it will be highlighted with a brief explanation of how to correct it.

#### Result

The number of aliquots that you specified will be added to the participant visit. The primary specimen will automatically change to unavailable to indicate that the specimen was consumed during processing.

### After you are finished

If you added multiple aliquots and any of those aliquots differed from the others (such as one aliquot having a different condition), be sure to make those changes to the aliquot.

### **Entering cryopreservation information**

Cryopreservation information can be entered for aliquot specimens with the CEL derivative type (PBMC Cells, Viable).

### **Steps**

- 1. On the **Specimen Management** page, location the specimen for which you are entering Cryopreservation information.
- **2.** From the **Edit** button to the right of the specimen, click the down arrow, and then click **Cryopreservation**.

This option will only be available for specimens with the derivative type CEL.

- **3.** Do one of the following:
  - If results were obtained, select Results Obtained.
  - If results were not obtained, select the reason there are no results from the **Reason** list.
- 4. If results were obtained, complete the information in the HIV Status, Primary Specimen Details, Aliquot Details, and Technician Details sections
- 5. In the **Processing Tech Initials** box, enter the processing technician's initials.
- **6.** In the **Data Entered By** box, enter your initials.
- 7. Click Save.

# Modifying aliquot specimens

You can modify individual aliquots specimens or modify multiple aliquot specimens together.

### **Prerequisites**

• All specimens to be modified must be from the same primary specimen.

### **Background**

#### **Steps**

- **1.** From the LDMS menu, click **Specimen Management**.
- **2.** Locate the specimens you want to modify.
- **3.** Select the aliquot specimens to modify.

To select multiple aliquot specimens, press and hold the Ctrl key. To select multiple, contiguous specimens, press and hold the Shift key instead.

- **4.** To the right of one of the selected specimens, click **Edit**.
- **5.** (When editing multiple specimens only) In the **Edit Aliquots** window, select the check box next to any boxes you want to change.
- **6.** Modify the specimen information as needed.
- 7. Optional: Making changes to Frozen Date, Frozen Time, Processing Tech Initials, Comments or Internal Only Comments for any single aliquot will give the user the option to cascade these changes to other aliquots. Changes can be cascaded to No other aliquots, All aliquots, Only aliquots having the same derivative type. Select an option and click **OK**.

Figure 24: The Aliquot Cascade Type Window



**8.** At the bottom of the **Edit Aliquots** window, click **Save**.

### **Creating Subaliquots**

Creating subaliquots is the process of making an aliquot from a pre-existing aliquot.

**Background** 

#### **Steps**

- Scan the source aliquot barcode to find its record in Specimen Management.
- 2. In the **Visit** grid, click the **Edit** drop down menu, select **Add New Primary**.
- **3.** In the **Create Primary Specimen** menu, add the following:
  - **3.1.** Other Specimen ID enter the Global Spec ID of the source aliquot into the field
  - 3.2. Additive
  - 3.3. Volume and Volume unit
  - **3.4.** Collection Time
  - 3.5. Receive Time
  - **3.6.** Adjust thaw counter
- 4. In the **Primary** grid, find the new Primary and use the **Edit** drop-down menu. Select **Add New Aliquot** and add the following in the **Create Aliquot** menu:
  - 4.1. Enter Derivative and Sub Add/Der Enter
  - 4.2. Volume and Volume unit

- **4.3.** Set Harvest Date to current date
- **4.4.** Adjust thaw counter
- 5. Click Save
- **6.** Find the Source aliquot and click the **Edit** button.
  - **6.1.** Adjust Available volume (subtract the volume removed)
  - **6.2.** Click Save
- **7.** Find the Subaliquot, click the **Edit** drop down menu:
  - **7.1.** Print Label

# **Delete a specimen**

If a specimen was added in error, it can be deleted completely.

### **Prerequisites**

Before deleting a specimen, ensure that it is appropriate to do so. Some projects require laboratories to assign specific condition codes or adjust specimen volume rather than deleting.

### Background

In most cases a specimen should not be deleted unless it was created in error. It is typically more appropriate to assign a specific condition code, such as *DSR* to the specimen.

Specimens that have been shipped to another laboratory cannot be deleted.

#### Steps

- **1.** On the navigation bar, click **Specimen Management**.
- **2.** Locate the specimen to be deleted.
- 3. On the specimen's **Edit** button menu, click **Delete**.
- **4. Warning:** This operation cannot be undone.

At the bottom of the Delete window, click **Delete**.

# Removing specimens from storage

Specimens can be removed from their storage locations directly on the **Specimen Management** page, without the need to switch to the **Storage** page.

#### **Steps**

- **1.** On the navigation bar, click **Specimen Management**.
- **2.** Locate the specimen to be removed from storage.
- **3.** On the specimen's **Edit** button menu, click **Remove From Storage**.

This option is only available if the specimen is assigned a storage location.

- 4. Optional: To change the specimen to *unavailable*, select **Set as Unavailable to be Stored Again**.
- 5. Click Remove.

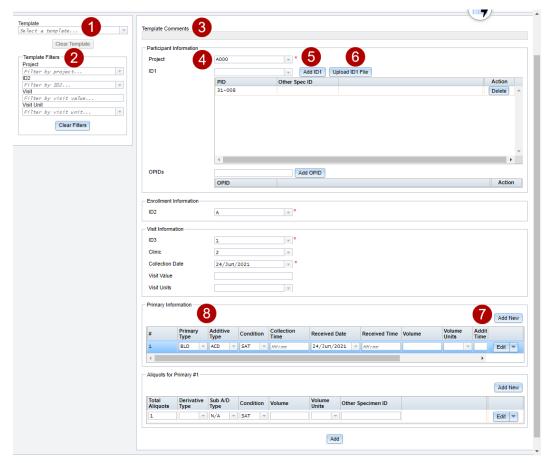
# Quick add

The quick add feature allow you to create all of the specimens from a participant visit at once, and is the preferred method for entering new visits in LDMS.

If you add specimens to a participant on the **Specimen Management** page, you would need to add each primary and set of aliquots individually. The quick add feature allows you to select or create a participant and enrollment, create a new visit, and enter all of the specimens collected at that visit from one screen. For new visits, this significantly reduces the amount of time it takes to enter a new visit in LDMS. You can even create templates for common visits to save more time.

Once a template has been used, any visits or specimens that were created are not associated with the template. This means that if the template is later changed or removed, visits that were entered using the template won't be changed.

Figure 25: Quick Add



(1) Select and apply template for Quick Add entry, (2) Filter list of templates, (3) Instructions applicable to selected template appear here, (4) Select project and ID1, (5) Add ID1 button used to add multiple ID1s simultaneously, (6) Upload list of ID1s, (7) Add new primary specimen row, (8) Primary specimens, numbers for convenience

# Adding specimens using Quick Add

Quick Add is used to enter information for a participant, enrollments, and multiple specimens all on one page.

### **Background**

This is useful when you want to enter a lot of information for a participant at once. Templates (if available) can be used to fill in predefined information for you.

#### Steps

- From the LDMS menu bar, select Specimen Management > Quick Add.
- **2.** Optional: From the **Template** list, select a template to apply.

Use the **Template Filters** to narrow down the list of templates. When you apply a template, a message with further instructions will appear.

A

**Warning:** Applying a template will clear any information you already entered on the **Quick Add** screen. In addition, once the template is applied, you cannot change projects unless you apply a different template or click **Clear Template**.

- **3.** Optional: Complete the **Participant Information**.
  - **3.1.** Select the project from the **Project** box.
  - **3.2.** Select the **ID1** for the participant *or* enter a new ID1 if you are adding a new participant.
  - **3.3.** To add multiple PIDs, click **Add ID1** or **Upload ID1 File** for each selected ID1
- **4.** Complete the **Enrollment Information** by selecting an **ID2** *or* by entering a new ID2.
- **5.** Complete the **Visit Information** section.
- **6.** Optional: Add a primary specimen.
  - **6.1.** In the **Primary Information** section, click **Add New**.
  - **6.2.** In the row that appeared, enter the information for the primary specimen to be created.
  - **6.3.** Optional: To view and modify all of the information for the primary, click **Edit** in the right column.
- **7.** Optional: Add an aliquot specimen.
  - **7.1.** Highlight the primary specimen for the aliquot.
  - 7.2. In the Aliquot Information section, click Add New.
  - **7.3.** In the row that appeared, enter the information for the aliquot specimen to be created.
- **8.** At the bottom of the page, click **Add**.

### Adding participants using Quick Add

The Quick Add feature can be used to add a new participant, along with other information, on a single page.

### **Background**

If adding a participant in this way, at least one visit must have occurred as the date of the visit (the **Collection Date**) is required.

#### Steps

- From the LDMS menu bar, click Specimen Management > Quick Add.
- 2. Optional: Select a template for the participant visit.
- **3.** Complete the information in the **Participant Information** section.
- 4. In the **Visit Information** section, enter the **Collection Date**.
- **5.** At the bottom of the page, click **Add**.

**6.** Optional: Complete any additional enrollment, visit, or specimen information for this participant.

#### Result

When the participant is successfully created, a note will appear at the top of the page. The participant will be available for select on the **Specimen Management** and **Quick Add** pages, where new enrollments, visits, and specimens can be added.

### Adding enrollments using Quick Add

The Quick Add feature can be used to add an enrollment to an existing participant, along with other information.

#### **Prerequisites**

The participant must already be added to LDMS before an enrollment can be added; enrollments cannot exist without being associated with at least one participant.

### **Background**

Using Quick Add to add an enrollment to a participant is especially useful if you want to enter other information at the same time, such as a visit or specimens.

### **Steps**

- 1. In the LDMS menu bar, click Specimen Management > Quick Add.
- 2. In the **Participation Information** section, select the **Project** and **ID1** for the participant.
- **3.** In the **Enrollment Information** section, select or enter the enrollment.
  - Note: If you enter an ID2 that does not yet exist, it will be created.
- **4.** Optional: Complete any additional visit or specimen information for this participant.
- **5.** At the bottom of the page, click **Save**.

### TBD in quick add templates

The TBD code can be used in quick add templates for certain sections that might need to be completed when the template is used.

For example, you may need create a template for a visit where the additive time used for a primary specimen varies based on certain factors. In these situations, using the **TBD** code will require the user to select the correct code when the template is used.

#### Places where TBD can be used in quick add templates

Primary type

- Additive type
- Derivative type
- Sub A/D type

### **Quick add templates**

Quick add templates are predefined specimen entry scenarios (typically for specific study visits) that can be used to automatically create some default specimens.

Templates are best used for specific study visits to create the expected specimen collections for the visit. They can also be used to complete other information, such as the ID2 and project that would be the same whenever the template is applied.

There are two types of templates: templates created by Frontier Science and templates that are created by your laboratory. Templates created by Frontier Science are designed in collaboration with leadership for projects and cannot be modified by users. Templates created by users at your laboratory can be modified. User-created templates are only available at the laboratory where they were created; they are not shipped or sharable with other laboratories.

Regardless of the type, once a template is applied, you will be prevented from changing the participant's project (meaning you can't apply a template intended for one project, and then change the project, effectively using the template for a project for which it wasn't intended). In addition, when you apply a template, any information that you already entered on the Quick Add screen will be cleared. This means that the template must be applied first, before you enter any other information.

### Creating quick add templates

Create quick add templates by defining the information for the visit.

#### Steps

- 1. From the LDMS menu bar, click **Specimen Management** > **Quick** Add Templates.
- **2.** Complete the **Template Information** section.

The **Name** is the name of your template as it will appear when selecting it when using the quick add feature. The **Comments** is a brief description of the template that will appear when the template is selected, and can be used to provide the person entering data if additional information or instructions, such as a reminder to remove an optional specimen if it was not collected.

- **3.** In the **Participant Information** section, select a **Project**.
- **4.** Optional: In the **Enrollment Information** section, select an **ID2** to be applied.
- **5.** Optional: In the **Visit Information** section, select an **ID3** to be applied.
  - **Important:** Either an ID2 or ID3 **must** be applied in order for the template to be created. Alternatively, both an ID2 and ID3

can be applied. These cannot be edited once the template has been triggered.

- **6.** Optional: Add a primary specimen.
  - **6.1.** In the **Primary Information** section, click **Add New**.
  - **6.2.** In the row that appeared, enter the information for the primary specimen to be created.
  - **6.3.** Optional: To view and modify all of the information for the primary, click **Edit** in the right column.
- **7.** Optional: Add an aliquot specimen.
  - **7.1.** Highlight the primary specimen for the aliquot.
  - 7.2. In the Aliquot Information section, click Add New.
  - **7.3.** In the row that appeared, enter the information for the aliquot specimen to be created.
- **8.** At the bottom of the page, click **Save Template**.

### Modifying or deleting a quick add template

Quick add templates can be changed or deleted without affecting specimens added using the template.

### **Background**

When a template is modified, the changes will be reflected when new visits are created using the template; existing entries will not be changed. If a template is deleted, it will be permanently removed, however any entries made using the template will remain.

### **Steps**

- 1. From the LDMS menu bar, click **Specimen Management** > **Quick** Add Templates.
- 2. In the **Saved Templates** box, select the template to be modified or deleted.
- **3.** Do one of the following:
  - Make any changes to the template, and then click **Save Template** at the bottom of the page.
  - At the bottom of the page, click **Delete Template**.
  - Click "Make Copy" to make edits to a copy of the selected template while preserving the original template.

# Differences between primaries and aliquots

This section describes the differences between how LDMS treats primaries and aliquots.

Table 3: Primary and aliquot comparison

|                    | Primary  | Aliquot                                       |
|--------------------|--|---|
| Global Specimen ID | Ends in -00  | Ends in -01, -02, etc.                        |
| Processing date    | Entered by user  | Uses value from primary                       |
| Enrollment         | Can be associated with more than one visit or protocol | Must select exactly one associated enrollment |

Certain information can be cascaded from primaries to derived aliquots specimens, so that if the primary is updated users will be asked if they want to update the aliquots as well.

- Frozen date
- Frozen time
- Comments
- · Internal comments
- · Tech initials

If a comment is cascaded from a primary to an aliquot, it will be added to any existing comment the aliquot already has.

# Specimen availability

Availability refers to whether or not a specimen exists at a laboratory.

A specimen's physical availability may change for a variety of reasons. For example, if a primary specimen was processed into aliquots, the primary specimen no longer exists and is thus not available. Likewise, if an aliquot's tube was damaged and its contents could not be recovered, it is also not available.

If a specimen is available, it can be added to a shipment and it can be assigned a storage location. If it is not available, LDMS will prevent you from shipping or storing the specimen.



**Note:** In LDMS for Windows, availability is called "never store".

Availability is a property of both primary specimens and aliquot specimens. By default, new specimens are available until you change them to unavailable or a specific condition automatically changes them.

There are several times where LDMS will automatically modify a specimen's availability. There are other places where you can manually change availability.

- You can manually change the availability of both primary and aliquot specimens, unless the specimen has been shipped.
- Specimens will automatically become unavailable if they are shipped.
- If a specimen's volume changes to zero, it is automatically made unavailable.

- A primary specimen will automatically become unavailable after aliquots have been derived from it, and you cannot change it back to available unless the aliquots are deleted.
- Certain condition codes, such as DSR (destroyed) will automatically change a specimen to unavailable.

Table 4: Specimen conditions that modify availability

| Code | Description             |  |
|------|-------------------------|--|
| ANM  | Sample Anonymized       |  |
| ANP  | Aliquot Not Prepared    |  |
| CDT  | Consumed During Testing |  |
| DSR  | Destroyed               |  |
| LSH  | Lost Shipment           |  |
| QNS  | Quantity Not Sufficient |  |
| SNC  | Sample Not Collected    |  |
| SNP  | Sample Not Processed    |  |
| SNR  | Sample Not Received     |  |

# **Specimen record details**

This section describes each specimen record entry box that appears on the Specimen Management page.

| Field            | Usage   | Example | Notes   |
|------------------|---|---------|---|
| Additional Time  | A value and a unit  | 4 Hrs   | Used to indicate information such as the amount of time a participant was fasting. Typically used for Pharmacology specimens to indicate the amount of time after the last dosage was taken.                  |
| Additive Type    | The additive in the primary collection tube                     | EDT     | Selected from a pre-defined list of codified values. If there was no additive in the tube, use NON. Hover the mouse pointer over an additive for a more detailed description.                                 |
| Additive Reagent |   |         |   |
| Aliquots         | List of any<br>aliquots for the<br>selected Primary<br>Specimen | -       | Displays the same specimen overview grid as seen on the <b>Specimen Management</b> page, specific to the primary's aliquots only. Aliquots can be edited from this window by clicking the <b>Edit</b> button. |
| Assigned Tests   |   |         |   |

| Field                 | Usage  | Example             | Notes   |
|-----------------------|--|---------------------|---|
| Available Volume      | A value and a unit   | 1.00 ML             | Used to indicate the volume of the sample available for use. Hover the mouse pointer over the units for a more detailed description.  |
| Clinic                | The clinical site where a visit occurred                         | 102                 | Select from a pre-defined list. For a description of a clinic, such as its name, hover the mouse pointer over it.   |
| Collection Date       | The date that a visit occurred                                   | 02/Apr/2014         | This will typically be one day. If a visit occurred over multiple days, each event will considered a separate visit and listed as separately in LDMS. They will still, however, have the same visit value and visit unit.   |
| Collection Time       | The time that the specimen was collected                         | 13:30               | This time should be the local time for the clinical site, and be in 24-hour format. In LDMS for Windows, collection time is known as specimen time  |
| Comments              | Additional<br>details about the<br>specimen                      |                     | Used to provide additional information, such as an explanation of a specimen's current condition. This comment will be included if the specimen data is shipped to another laboratory.  |
| Derivative Type       | The type of aliquot specimen created                             | PL2                 | Selected from a pre-defined list of codified values. Hover the mouse pointer over a type for a more detailed description.   |
| Enrollment            | The project-<br>ID2 combination<br>associated with<br>an aliquot | project/ID2         | The enrollment is in the format [project]/[ID2].  |
| Frozen Date           | The date the specimen was frozen                                 | 04/Jun/2014         | This is generally used to indicate the date the freezing process began.   |
| Frozen Time           | The time the specimen was frozen                                 | 15:00               | This is generally used to indicate the time the freezing process began.   |
| Global Specimen<br>ID | A unique identifier for the specimen                             | 500-<br>AEQ0052R-01 | This will be unique for the specimen. LDMS will never generate the same global specimen ID, even at two different laboratories. Global specimen IDs for primaries always end in "-00"; global specimen IDs for aliquots will have the same ID as their parent primary, but end in "-01", "-02", and so forth. Specimens added in LDMS for the web will have the originating laboratory's ID number as the first 3 digits. Global specimen IDs generated by LDMS for Windows will lack this feature. |

| Field                     | Usage   | Example                                | Notes   |
|---------------------------|---|--|---|
| Harvest Date              | Indicates the date that a culture derivative sample was harvested.                    | 04/Jun/2014                            | -   |
| ID3                       | A sub-protocol or<br>study  | A50250562I                             | Used by some projects to further identify the protocol. This is typically selected from a pre-defined list. For some protocols, a temporary value such as "NOSID" may be used until the exact value is assigned to the participant. For other protocols, this field may not be used or may be optional. |
| Imported                  | Indicates if the sample was received and if so, on what date.                         | Shipment 12 on<br>11/Aug/2005 or<br>No | Link to <b>Received Shipment Review</b> page, if applicable.  |
| Import Date               | Date the shipment with the specimen was received                                      | 11/May/2016                            | If the specimen was not received in a shipment, this will be blank.   |
| Internal-Only<br>Comments | Additional details about the specimen   |  | These comments will <i>not</i> be included if the specimen data is shipped to another laboratory.   |
| Is Available              | Indicates if the specimen is available  | €                                      | Availability means that the specimen exists and can be shipped, stored, and so forth. Reasons that a specimen might not exist include being destroyed or being shipped to another laboratory.   |
| Owner<br>Enrollment       | The enrollment<br>that "owns" a<br>primary specimen                                   | project/ID2                            | For primary specimens that have not been processed into aliquots, this is the enrollment that "owns" the specimen   |
| Original Volume           | Indicates the volume of the sample before any aliquots were created or tests were run | 1.0 ML                                 | -   |
| Other Specimen<br>ID      | A laboratory-<br>defined identifier   |  | There are no rules or validation for this field; it can be used to assign a special identifier if needed by your laboratory.  |
| Primary Type              | The type of primary specimen  | BLD                                    | Selected from a pre-defined list of codified values. Hover the mouse pointer over a type for a more detailed description.   |
| Processing Date           | Date a primary<br>specimen was<br>processed into<br>aliquot specimens                 | 15/Oct/2012                            | Use local time. If present, the value entered for the parent primary specimen will be used for aliquots derived from the primary.   |

| Field                            | Usage   | Example                          | Notes  |
|----------------------------------|---|----------------------------------|--|
| Processing Tech<br>Initials      | The initials of<br>the person who<br>processed a<br>primary specimen<br>into aliquot<br>specimens             | JD                               | If the processing tech initials are set for a primary, aliquots derived from the primary will receive those initials as well. You can change the initials for individual aliquots, if needed.  |
| Processing Time                  | Time a primary specimen was processed into aliquot specimens  | 16:15                            | Use local 24-hour time. If present, the value entered for the parent primary specimen will be used for aliquots derived from the primary.  |
| Reason Specimen<br>Not Collected | Explanation of<br>why an expected<br>specimen was not<br>collected during a<br>visit                          | Specimen potentially compromised | Typically used in conjunction with a specimen condition code that indicates an issue with the specimen, such as "QNS" (quantity not sufficient). If necessary, use the <b>Comments</b> box for the specimen provide more details.                        |
| Received Date                    | The date the specimen was received by the processing laboratory where the primary was processed into aliquots | 12/Mar/2014                      | -  |
| Received Time                    | The time the specimen was received by the processing laboratory   | 14:45                            | Times should be entered in 24-hour local time.   |
| Shipped                          | Whether the specimen has been shipped to another laboratory   | Yes, no, or pending              | Provides link to shipment, if applicable   |
| Specimen<br>Condition            | The current condition of the specimen   | SAT                              | Selected from a pre-defined list of codified values. Hover the mouse pointer over a condition for a more detailed description.   |
| Specimen ID                      | A legacy<br>specimen ID used<br>only by LDMS  | 500V10000110                     | The specimen ID is an identifier for the specimen that was used by older laboratory management software. It will only be present for specimens that were entered or migrated from LDMS. Specimens created using LDMS will not be assigned a specimen ID. |

| Field                       | Usage  | Example     | Notes   |
|-----------------------------|--|-------------|---|
| Status                      | Icons indicating<br>various<br>properties about<br>a specimen  |             | Specimen is available Specimen is assigned a storage location Specimen has been shipped to another laboratory Specimen has an additional enrollment   |
| Stored                      | Whether the specimen currently has a storage location assigned | Yes or no   | Provides link to storage location   |
| Sub-Additive/<br>Derivative |  |             | If the specimen does not have a subadditive/derivative, select NON.   |
| Thaw Count                  | The number of times the specimen has been frozen and thawed.   | 3           | -   |
| Total Cell Count            | Total number of cells (in millions)                            | 10          | When entered, it will be multiplied by 1 million. For example, to indicate 10 million cells, enter 10; to indicate 500,000 cells, enter 0.5.  |
| Tube Count                  | The number of primary tubes collected for the pooled sample    | 5           | This field is used to record the number of primary tubes collected for the sample in scenarios where the tubes are pooled into a single sample when recording in LDMS.  |
| Unavailable Date            | The date the specimen is no longer available                   | 12/Mar/2014 | -   |
| Visit Value                 | The protocoldefined identifier for a visit                     | 1 Wk        | This comprises of two boxes, a visit number and visit type. A visit value is typically a number or letter. The visit unit is selected from a pre-defined list. Hover the mouse pointer over a visit unit for a brief description. |

# **Specimen conditions**

A specimen condition is a 3-letter code that indicates the status of a specimen.

Every *primary specimen* and *aliquot specimen* in LDMS has a condition assigned to it. The default condition for new specimens is SAT (satisfactory), but there are many others.

The majority of conditions indicate a deviation from the expected handling for a specimen. For example, if a specimen was supposed to be collected in one

type of tube but another was used, the INT (incorrect tube type) condition code could be used to indicate this.

There are several places where the condition of specimens can be changed:

- on the Specimen Management page, by editing an individual specimen
- when receiving specimen data from a shipping file
- on the storage page all specimens in a storage location can be modified

A common use for specimen conditions is indicating when an expected primary specimen was not collected. By setting the specimen's condition to an applicable code (such as QNS if not enough sample could be collected) and by entering descriptive comments, you can explain the omission of the expected specimens.

### **Destroying a specimen**

Specimen destruction is noted in LDMS by applying the  ${\tt DSR}$  specimen condition.

### **Background**

When a specimen is destroyed, it is important to document that destruction in LDMS.

### **Steps**

- **1.** From the menu bar, click **Specimen Management**.
- **2.** Using filters on the left, locate the specimen to be destroyed.
- **3.** To the right of the specimen, click **Edit**.
- 4. Change Specimen Condition to DSR.
- **5.** (If the specimen has been assigned a storage location) Write down the storage location of the specimen, if it has not already been physically removed.



**Warning:** The specimen will automatically be un-assigned from its storage location in LDMS. This is your last chance to get the specimen's storage location.

**6.** In the **Comments** box, enter an explanation of why the specimen was destroyed.

### After you are finished

If you have not already done so, use the storage location that was noted by LDMS to find the specimen and physically remove it from storage. The specimen was automatically removed from storage in LDMS.

### **Procedural conditions**

These specimen condition codes describe issues that can occur in the collection and processing environment.

**Table 5: Procedural conditions** 

| Code | Description                       | Usage   |
|------|-----------------------------------|---|
| ANP  | Aliquot Not Prepared              | Indicate that an expected aliquot was not processed, but no other more specific condition code applies. The <b>reason specimen not collected</b> box should be used in conjunction with ANP.              |
| EQF  | Equipment Failure                 | There was an issue with the processing equipment, such as a power failure during processing.  |
| INV  | Invalid                           | The specimen is not valid for testing purposes. Generally another condition code, such as LBE or PST, would be more appropriate.  |
| LBE  | Laboratory Error                  | The laboratory made a general error during processing, such as setting equipment up incorrectly or mislabeling specimens.   |
| OPR  | Outside Protocol<br>Requirements  | The specimen was drawn during the protocol-defined window, but not in a manner consistent with the protocol. For example, if a participant was supposed to be fasting but was not, this code would apply. |
| osw  | Outside Visit Window              | The specimen was collected correctly, but outside the window for the visit as specified by the protocol.  |
| PST  | Processed After Specified<br>Time | The specimen was collected and processed into aliquots, but the processing was done after the time frame specified by the protocol. This is commonly applicable for PBMC and pharmacology specimens.      |

# **Specimen container conditions**

These condition codes describe physical problems with the specimen's container. If there is a qualitative issue with the specimen as a result of the container issue, a qualitative code may be more appropriate.

**Table 6: Specimen container conditions** 

| Code | Description    | Usage  |
|------|----------------|--|
| BKV  | Broken Vial    | The container was broken beyond recovery, such as being dropped and shattered on the ground.   |
| DMG  | Damaged        | The container is not leaking, but is damaged in another way, such as a tear in a label or a dent in a plastic container.                               |
| EXP  | Expired        | The additive in a container, or some other component used during specimen collection, was expired.   |
| INT  | Incorrect Tube | A tube type other than the one specified by the protocol was used, and the tube used was determined to be an acceptable alternative by the study team. |
| LKD  | Leaked         | The specimen leaked from the container, and it was placed inside another container and recovered.  |

### **Temperature conditions**

Temperature condition codes are typically applicable if there was an issue during shipping (such as sublimed dry ice) or if a piece of storage equipment failed. Other codes are applied as part of routine and expected handling of specimens. For example the FRO condition code can be used to indicate that a specimen intended to be stored ambient or refrigerated was frozen.

**Table 7: Temperature conditions** 

| Code | Description             | Usage  |
|------|-------------------------|--|
| DIM  | Dry Ice Melted          | The specimen was kept on dry ice, but the dry ice sublimed. This does not necessary imply that the specimen was damaged or TNO.  |
| FRO  | Frozen                  | The temperature of the specimen has been lowered below its freezing point. This applies only to specimens that were not intended to be frozen, such as specimens that were shipped at ambient temperature during the winter.   |
| REF  | Refrigerated            | The specimen is cooler than room temperature but not below its freezing point. This applies only to specimens that were not intended to be refrigerated.   |
| TNO  | Temperature Not Optimal | The specimen was stored or shipped at any temperature other than the temperature specified by the protocol.  |
| TWD  | Thawed                  | The specimen was frozen, and has been warmed so that it is no longer frozen. LDMS has a field to track a thaw count, which can be used to keep track of how many times the specimen has been thawed. This information can be found on the Details Window for the aliquot in Specimen Management. |

### Shipping conditions

These condition codes apply to issues with a specimen shipment where the specimens are unaccounted for or were received late. If the shipping issue damaged the specimens, a temperature-related code such as DIM (dry ice melted) may be more appropriate.

**Table 8: Shipping conditions** 

| Code | Description         | Usage  |
|------|---------------------|--|
| DSH  | Delayed Shipment    | A shipment did not occur on schedule, but there is no obvious damage to the specimens. If there was obvious damage, a more descriptive, temperature-related code (such as DIM or TNO) may be more appropriate. |
| LSH  | Lost Shipment       | A shipment was created and sent, but did not arrive at the receiving laboratory. Neither the shipping nor the receiving laboratory can locate it.  |
| SNR  | Sample Not Received | A data collection form, such as a CRF, was received, but a specimen listed on the form was not received. This differs from LSH in that only some specimens were not received, compared to an entire shipment.  |

### **Qualitative conditions**

These codes refer to the quality of a specimen. For example, if a blood specimen clotted, this is a qualitative issue that could prevent processing. If there was a qualitative or participant abnormality that would prevent processing but no other code is applicable, the SNP condition code can be used.

**Table 9: Qualitative conditions** 

| Code | Description   | Usage  |
|------|---|--|
| BLD  | Bloody  | The specimen, such as a throat swab, contained blood.  |
| CLT  | Clotted   | The specimen has clotted, often because the additive did not mix correctly with the specimen.  |
| СТМ  | Contaminated  | The specimen is visibly contaminated.  |
| DCG  | Discharge present   | A specimen, such as a vagina swab, that contains discharge material  |
| HEM  | Hemolyzed   | A blood specimen that has hemolyzed.   |
| HUM  | Humidity  | The specimen has been exposed to high humidity.  |
| ICT  | Icteric   | There are excessive amounts of bilirubin in the specimen.  |
| LIP  | Lipemic   | There is excessive fat content in the specimen.  |
| LYS  | Lysed   | There has been a breakdown of cells in the specimen other than hemolyzation.   |
| NQA  | Real-time QA for viability<br>and viable recovery not<br>performed due to low<br>volume of sample collected | Used by a Leukopak processing laboratory to indicate that the full specimen volume needed for PBMC viability and viable recovery QA could not be obtained.   |
| SNP  | Sample Not Processed  | A generic code for when there is a qualitative issue that prevents an otherwise correctly collected primary from being processed into aliquots, but no other code applies. The Reason sample not collected field should be used in conjunction with SNP. |
| VPL  | Viability percentage may<br>be less than the expected<br>parameter  | Used by a processing laboratory to indicate that the PBMC viable percentage of the PBMC specimen may be lower than what's expected.  |
| VRU  | Viable recovery may<br>be outside expected<br>parameters (higher or<br>lower)                               | Used by a processing laboratory to indicate that the PBMC viable recovery of the PBMC specimen may be outside what is expected.  |

# **Quantitative conditions**

These condition codes refer to the volume of specimen collected.

**Table 10: Quantitative conditions** 

| Code | Description             | Usage  |
|------|-------------------------|--|
| DFB  | Difficult bleed         | There was problems trying to obtain a blood specimen from the participant, typically resulting in a lower volume than was expected. This code is more specific than SHV, which does not indicate why the volume was low.   |
| QNS  | Quantity Not Sufficient | There was not enough specimen available to create the aliquot. For example, if specimen collected from the participant was supposed to be 10 mL but only 5 mL was collected, the aliquots that could not be created due to the low volume would be considered QNS. QNS means there is no volume at all for the aliquot, even though there was some volume for the primary. |
| SHV  | Short Volume            | The primary or aliquot specimen has at least some volume, but not the full expected volume.  |
| SNC  | Sample Not Collected    | The primary specimen was not collected from the participant at all. This might happen if the participant declined to provide a specific sample.  |

### Other conditions

These condition codes are either automatically assigned to specimens or indicate some combination of issues. The default condition code that is assigned to all new specimens is SAT (satisfactory).

**Table 11: Other conditions** 

| Code | Description                | Usage   |
|------|----------------------------|---|
| ANM  | Anonymized                 | The specimen was created using the anonymization tool in LDMS. This code is automatically assigned by LDMS and cannot be assigned manually.   |
| coc  | Combination of Codes       | More than one condition code applies; the comments field in LDMS can be used to list the applicable codes.  |
| DSR  | Destroyed                  | The specimen has been destroyed.  |
| ОТН  | Other                      | There is something noteworthy or unusual about the specimen, but no other available condition code applies.   |
| SAT  | Satisfactory               | The default condition code for new specimens, indicating that the specimen was collected, processed, and handled as expected.   |
| UNK  | Unknown                    | Indicates that there is a significant gap in knowledge in the specimen's history. For example, if the specimen was collected by another laboratory that was not using LDMS and is several years old, and it may have been stored or treated improperly, this code may be appropriate. |
| YST  | Did consent to storage     | The participant has provided consent to storage of the specimen.  |
| NST  | Did not consent to storage | The participant did not provide consent to storage of the specimen.   |

# **Test assignment**

Specimens can be assigned tests to indicate a specific test that is expected.

While specimens cannot be tested in LDMS for the web, specimens may be shipped to a laboratory using a LDMS for Windows. The laboratory using LDMS for Windows may then enter results for the expected test.

### Assigning tests to a specimen

Expected tests can be assigned to a specimen, which will make it easier to locate the specimen when setting up assay test runs.

### **Steps**

- 1. On the navigation menu, click **Specimen Management**.
- 2. Locate the specimen to have a test assigned. From that specimen's **Edit** menu, click **Assign Tests**.
- In the Test list, select the test to assign.
  If Other was selected, enter the name of the test in the Other Test
  Name box.
- 4. Click Assign Test.

### Indicating that a test will not be performed

If an assay is expected but will not be performed, the test can be assigned and then indicated that it will not be tested.

#### Steps

- **1.** On the navigation menu, click **Specimen Management**.
- 2. Locate the specimen to have a test assigned. From that specimen's **Edit** menu, click **Assign Tests**.
- **3.** For the test, do the following:
  - **3.1.** In the **Test Not Performed** column, select the check box.
  - **3.2.** In the **Reason For No Result** column, select the appropriate code.
    - Hover over a code to see a brief description.
  - **3.3.** Optional: In the **Comment** column, enter more information about why the test was not performed.
- 4. Click Save.

### **Deleting a test assignment**

#### **Background**

A test assignment will typically only be deleted if it was added in error. If the test was expected but will not be run, the appropriate code could be assigned to the test instead.

### Steps

- **1.** On the navigation menu, click **Specimen Management**.
- **2.** Locate the specimen to have a test assigned. From that specimen's **Edit** menu, click **Assign Tests**.
- **3.** In the **Action** column, click **Delete**.
- 4. Click Save.

# **Storage**

The Storage page is used to define how your real-world storage is set up and where individual specimens are stored.

The Storage page is organized into four sections:

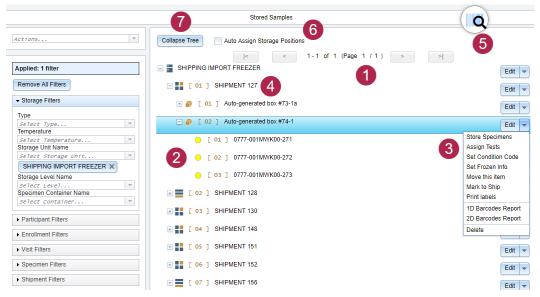
| Stored specimens       | This is where you will actually perform storage work, such as creating a new storage unit or assigning a storage location to specimens. |
|------------------------|---|
| Storage unit templates | This is where you can define templates for commonly used storage units.   |
| Level templates        | This is where you can define templates for commonly used <i>storage levels</i> .  |
| Container templates    | This is where you can define templates for commonly used storage containers.  |

# Navigation on the Storage page

The **Storage** page shows a representation of your real-life storage as a collapsible tree.

This tree is called the storage tree view, and shows all of the items in storage as you have defined them in LDMS.

Figure 26: The Storage page



(1) Navigate between pages (if your storage page does not fit on one). (2) The location of the specimen in "Autogenerated box #74-1". (3) Edit menu for storage item named "Auto-generated box #74-1". (4) The storage tree view. (5) Indicator that you can scan a barcode to add samples into a storage location. (6) The ability to automatically assign storage locations. (7) Close all units, levels, and containers.

The storage units in the storage tree are sorted alphabetically by name. The contents within storage units are sorted by position. Before expanding any storage items however, you can scan a barcode to search for a stored item automatically when the following icon is shown at the top of the page:



Click on the + button to the left of a storage item to open it. The contents in the square brackets next to the item's name (such as [ 1, 2 ]) indicates that items position in its parent storage item. To the right of each storage menu is the **Edit** menu for that item. The options available from this menu will vary, depending on the type of storage item.

### The storage tree

The storage system in LDMS is a hierarchy of storage items.

There are several types of storage items in LDMS. From biggest to smallest, these are:

**storage unit** This is the main cooling unit used to store specimens, such as a freezer or a refrigerator.

**Ievel** This is an intermediate part of a storage system, such as a shelf within a freezer. You can add an additional level to

another level. For example, you might have a shelf that contains racks of boxes, where the rack is a level on the shelf, which is a level in a freezer.

#### container

This is the storage item that holds specimens, such as a box.

Figure 27: The LDMS storage hierarchy



The storage hierarchy is visualized on the **Storage** page in LDMS as a tree.

Figure 28: The storage tree on the Storage page



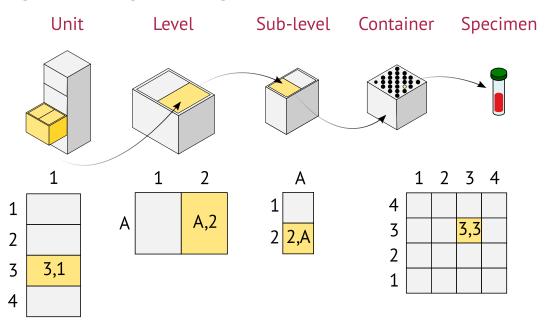
(1) A storage unit. (2) A level. (3) A container. (4) Specimens in the container. (5) The name for this specific storage unit. (6) The position of the item in its parent. (7) The global specimen ID for each specimen.

# **Storage items**

Storage items in LDMS are represented as nested grids.

Each storage item contains rows and columns that represent individual locations within that storage item. For example, a shelf that can hold 10 boxes might have the dimensions of  $2 \times 5$  (2 rows with 5 columns to represent each box). This is true of all storage items, from big storage units to small containers.

Figure 29: Storage items as grids

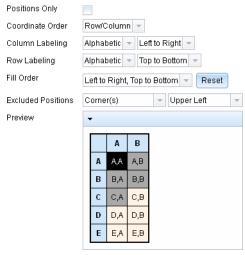


This visual illustrates the nested grid of storage items. Below each item is a grid showing where the highlighted item is positioned.

Rows and columns can be represented as either numbers or letters. Individual sections of a storage item can also be represented as a position number instead of coordinates. For example, if you have a  $1 \times 3$  shelf, each position could be represented as 1, 2, and 3, rather than (1,1), (1,2), and (1,3).

Storage items can optionally have an *excluded position*. An excluded position is useful for determining which side of the storage item is the front and which is the back. For example, if you know that the upper-right corner of a container will be empty, you know where to start inserting specimens.

Figure 30: Configuring a storage item layout



The preview shows how these settings will be interpreted by LDMS. A,A (black) is an excluded position. A,B to C,A (gray) contain another storage item or specimen. C,B to E,B (tan) are empty positions.

All of these properties of storage items can be configured when adding new items to the storage tree. They can also be defined in advance by using a storage template.

### Icons and their meaning

The icons next to the items in the storage tree indicate the type of storage item, and special properties such as its additive or whether the item is part of a pending shipment.

Table 12: Storage item icons

| Icon     | Meaning  |
|----------|--|
|          | Storage unit   |
| II.      | Storage unit<br>that has been<br>flagged 'mark<br>to ship' |
|          | Storage unit<br>that is part<br>of a pending<br>shipment   |
|          | Level  |
| <b>=</b> | Level that has<br>been flagged<br>'mark to ship'           |
|          | Level that<br>is part of<br>a pending<br>shipment          |

| Icon     | Meaning   |
|----------|---|
| ==       | Sub-level   |
|          | Sub-level<br>that has been<br>flagged 'mark<br>to ship' |
|          | Sub-level<br>that is part<br>of a pending<br>shipment   |
|          | Container   |
|          | Container<br>that has been<br>flagged 'mark<br>to ship' |
| <b>₫</b> | Container<br>batched                                    |
| •        | Specimen  |
|          | Specimen<br>that has been<br>flagged 'mark<br>to ship'  |
|          | Specimen that is part of a pending shipment             |

Table 13: Additives with colored tubes on the storage page

Any additive not listed in this table uses a black icon.

| Additive | Icon |
|----------|------|
| ACD      | •    |
| AHP      | •    |
| DPE      |      |
| DSE      | •    |
| EDT      |      |
| HEP      | •    |
| LHG      |      |
| LHP      | •    |
| LPE      | •    |
| LSE      | •    |

| Additive | Icon |
|----------|------|
| NON      | •    |
| PED      |      |
| SCI      | •    |
| SED      | •    |
| SFL      |      |
| SST      | •    |
| THM      | •    |

# **Assigning storage locations**

### Adding a new storage unit

A storage unit is the largest type of storage item, and is used to hold levels.

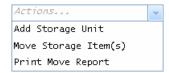
### **Background**

A new storage unit is created by defining various properties for it, such as its size and temperature. You can manually define these properties for the storage unit as you create it, or you can use a template that applies default settings for you.

#### Steps

- Click Storage > Stored Specimens from the LDMS menu bar.
- **2.** From the action menu, select **Add storage unit**.

Figure 31: The LDMS action menu



The **Create storage unit** window will open.

**3.** Optional: Select a template to use for the new storage unit from the **From template** box.

All settings from the template will be set for you. You can override any settings from the template if needed. Any changes made will only affect the new storage unit; the template will not be modified.

**4.** Optional: Click the **Empty Structure** button to remove any levels that were automatically added by a template.

This is only necessary if you do not want to use the default levels (if present) from a template.

All levels added by a template will be removed.

- **5.** Enter a descriptive name for the new storage unit in to **Name** box. This is the name for the new storage unit as it will appear in the storage tree in LDMS.
- **6.** Select the type of storage unit from the **Type** box.
- **7.** Select the temperature of the new storage unit from the **Temperature** box.
- **8.** Enter the number of rows and columns for the new storage unit into the **Number of rows** and **Number of columns** boxes.

This represents the size and capacity of the new storage unit in terms of the number of *levels* that it can hold. For example, if you are creating a freezer that can hold 5 shelves vertically, you might have 1 column and 5 rows.

**9.** Select or deselect the **Positions only** option as desired.

If the positions only option is selected, each item in the level in the new storage unit will be identified by a number. If it is not selected, each level will be identified by its coordinates.

- **10.** (If **Positions only** is not selected) Define how levels in the new unit should be identified.
  - **10.1.**Select either Row/Column or Column/Row from the Coordinate order box.

If Row/Column is selected, each level's position will be identified in the format (row,column)

**10.2.**Select how columns and rows are to be labeled from the **Column labeling** and **Row labeling** boxes.

To use letters to identify rows or columns, select Alphabetic; to use numbers, select numeric.

- **10.3.**Select the fill order for the new unit from the **Fill order** box. Fill order can be used when adding a new level to the unit. While you must specify a fill order, you can override it when adding levels to the storage unit.
- **10.4.**Optional: Select a position(s) in the unit to be excluded from the **Excluded positions** box.

Excluded positions are used to help identify the orientation of the container (meaning which side is the front and which is the back).

**11.** Review the **Preview** of the new unit.

If any empty levels will automatically be added the new unit, they will be listed in the **Default Levels** section.

- **12.** Optional: To save the current configuration as a new template, do the following.
  - **12.1.**Select **Save As New Template**.
  - **12.2.**In the **New Template Name** box, enter a name for the template.
- **13.** Click the **Save** button.

#### Result

The new unit will have been added to the storage tree. The storage tree is in alphabetical order, so you may need to change pages in the storage tree to find your new storage unit.

### Adding a new level in a storage unit

A new level is added to an existing storage unit or storage level.

### **Prerequisites**

You must have created the storage unit before you can add levels to storage in LDMS. In other words, you cannot create a level that is not associated with a storage unit.

#### Steps

- 1. Click **Storage** > **Stored Specimens** from the LDMS menu bar.
- 2. Find the *storage unit* to which you are adding a *level*.

  If you want to add a sub-level to an existing level, find that level instead. You can used the filters on the left side of the page to help find the storage item.
- Click the down arrow next to the Edit button to the right of the intended parent storage unit or level, and then click Add new level.

Figure 32: The Edit Storage Unit button



The **Select position for new level** window will open.

- **4.** Optional: Select a template to use for the new level from the **From template** box.
  - All settings from the template will be set for you. You can override any settings from the template if needed. Any changes made will only affect the new level; the template will not be modified.
- Enter a descriptive name for the new level in to Name box.
  This is the name for the new level as it will appear in the storage tree in LDMS.
- **6.** Enter the number of rows and columns for the new level into the **Number of rows** and **Number of columns** boxes.
  - This represents the size and capacity of the new level in terms of the number of levels or *containers* that it can hold. For example, if you are creating a shelf that can hold 5 boxes side-by-side, you might have 5 columns and 1 rows.
- **7.** Select or deselect the **Positions only** option as desired.

If the positions only option is selected, each item in the new level will be identified by a number. If it is not selected, each level will be identified by its coordinates.

- **8.** (If **Positions only** is not selected) Define how levels or containers in the new level should be identified.
  - **8.1.** Select either Row/Column or Column/Row from the Coordinate order box.

If Row/Column is selected, each level or container position will be identified in the format (row,column)

**8.2.** Select how columns and rows are to be labeled from the **Column labeling** and **Row labeling** boxes.

To use letters to identify rows or columns, select Alphabetic; to use numbers, select numeric.

- 9. Select the fill order for the new level from the **Fill order** box. Fill order can be used when adding a new level or container to the level. While you must specify a fill order, you can override it when
- **10.** Optional: Select a position(s) in the level to be excluded from the **Excluded positions** box.

Excluded positions are used to help identify the orientation of the container (meaning which side is the front and which is the back).

- **11.** Review the **Preview** of the new storage item.
- 12. Click the **Continue** button.
  The **Select position** window will open.

adding items to the level.

- **13.** Specify the position within the storage unit for the new level by doing one of the following:
  - Select a position for the new level from the **Position** box.
  - Click on a position in the **Preview** section.

Positions that cannot be selected are either already occupied by another level or empty by exclusion rules for the storage unit. Gray positions are occupied; black positions are excluded.

- **14.** Optional: To save the current configuration as a new template, do the following.
  - 14.1. Select Save As New Template.
  - **14.2.**In the **New Template Name** box, enter a name for the template.
- **15.** Click the **Save** button.

## Adding a container to a level

A new storage container is added to an existing level or sub-level.

### **Prerequisites**

You must have created the storage units and levels for the container before adding the container. In other words, you cannot add a container to a freezer without a level, nor can you create a container that is not within a freezer.

### **Steps**

- 1. Click **Storage** > **Stored Specimens** from the LDMS menu bar.
- Find the storage unit to which you are adding a level.
  If you want to add a sub-level to an existing level, find that level instead.
- **3.** Click the down arrow next to the **Edit** button to the right of the level that will hold the container, and then click **Add New Container**.

Figure 33: The Edit level button



If the level has had sub-levels added, the **Add new container** option will not be available. A container cannot be added side-by-side with a sub-level.

The **Create container** window will open.

- **4.** Enter the number of new *containers* to add in the **Number to add** box.
- 5. Optional: Select a template to use from the **From template** box. The information from the template will be populated into the remaining boxes. You can still modify this information, if needed. Any changes made will only affect the *containers* that you are creating.
- **6.** Enter the number of rows and columns for the new container into the **Number of rows** and **Number of columns** boxes.

This represents the size and capacity of the new container in terms of the number of specimens that the container can hold. For example, if you are creating a box that can hold 25 specimens, you might have 5 columns and 5 rows.

- Note: The system will warn the user when a box is outside the max 26x26 dimensions.
- **7.** Select or deselect the **Positions only** option as desired.

If the positions only option is selected, each specimen in the container will be identified by a number. If it is not selected, each specimen will be identified by its coordinates.

- **8.** (If **Positions only** is not selected) Define how specimens in the new container should be identified.
  - **8.1.** Select either Row/Column or Column/Row from the Coordinate order box.

If Row/Column is selected, each specimen's position will be identified in the format (row,column)

**8.2.** Select how columns and rows are to be labeled from the **Column labeling** and **Row labeling** boxes.

To use letters to identify rows or columns, select Alphabetic; to use numbers, select numeric.

- **8.3.** Select the fill order for the new level from the **Fill order** box. Fill order will be used when adding new specimens to the container, and you chose to use automatic positioning. While you must specify a fill order, using the automatic positioning feature is optional.
- **8.4.** Optional: Select a position(s) in the container to be excluded from the **Excluded positions** box.

  Excluded positions are used to help identify the orientation of

the container (meaning which side is the front and which is the back).

- **9.** Optional: To save the current configuration as a new template, do the following.
  - **9.1.** Select **Save As New Template**.
  - **9.2.** In the **New Template Name** box, enter a name for the template.
- Click the Continue button.
   The Select position window will open.
- **11.** Enter a descriptive name for the first container into the **Name** box. This is the label for the new container as it will appear in the storage tree.
- **12.** Specify the position for the new container on its parent level by doing one of the following:
  - Select a position for the new container from the **Position** box.
  - Click on a position in the **Preview** section.

Positions that cannot be selected are either already occupied by another container or empty by exclusion rules for the parent level. Gray positions are occupied; black positions are excluded.

**13.** Click the **Continue** button.

You will need to specify the **Name** and position for each new container that you are creating.

### Result

The new *containers* will be created in the positions specified.

## Assigning a storage location to specimens

Individual specimens are added to containers on the **Storage** page.

### **Prerequisites**

You must create the specimens on the Specimen Management page prior to adding them to a container. You must also have created the *container* to which the specimens will be added.

### **Background**

Individual specimens are stored in containers. You cannot add specimens to a level or *storage unit* without a container.

### **Steps**

- Click Storage > Stored Samples from the LDMS menu bar.
- **2.** Find the storage container to which you want to add specimens in the storage tree.

Use the filters on the left side of the screen to assist in finding the container.

**3.** From the **Edit** menu to the right of the container, select **Store specimens**.

Figure 34: The Edit container menu



The **Select Specimens** window will open.

- **4.** Do one of the following:
  - Use filters to find specimens to limit the specimens displayed at the bottom of the page
  - Click Upload File: Unique specimen IDs (file must be a text file with one specimen ID per line)
    - Note: Supported ID formats include Specimen IDs, Other Spec IDs, Global Spec IDs, and Database IDs.
  - Scan a specimen barcode
- **5.** For each specimen to be stored, select the check box in the **Selected** column.

You can also click **Select All** to store all currently displayed specimens.

If you change filters at this point, any specimens selected will remain selected.

- **6.** At the bottom of the Select Specimens window, click **Continue**. The **Select position for specimen** window will open.
- **7.** Select the position where the first specimen will be stored from either the **Position** box or the **Preview** image.

The *global specimen ID* for the specimen will be displayed in the window. Positions that are gray are not available because a specimen already occupies that location. A position that is black is an *excluded position* in the container's configuration.

- **8.** Optional: To place the remaining specimens to be stored automatically based on the container's fill order setting, select the **Auto-fill from selected position** option.
- 9. Click Continue.

If there are more specimens to be stored and you did not select the **Auto-fill from selected position** option, you will be prompted to select a position for those specimens, otherwise, the window will close.

### Result

The specimens are now stored. If you look at the storage tree, you will see that they are now listed under the specified container.

### After you are finished

Generate the Storage Details and Container reports. These will help you know where the specimens need to physically be stored.

## Generating a printable list of what is in storage

The Storage Details and Container reports can be used to print a list of all the storage items or specimens in a specific location.

### **Background**

In addition to assigned specimens to a storage location in LDMS, you will need to actually put the real-life specimens in storage. The Storage Details and Container reports can assist you in putting storage items in the correct location. It is good practice to print them after making changes to storage in LDMS.

### **Steps**

- 1. Click **Storage** > **Stored Samples** from the LDMS menu bar.
- 2. In the storage tree, locate the storage item for which you need to generate a report.
- **3.** To the right of the storage item, click **Edit**.
- **4.** Click the down arrow next to the **Reports** button, and then click either **Storage detail** or **Container**.
- 5. In the **File Type** box, select PDF (\*.pdf) and then click **Generate Report**.

## Container report

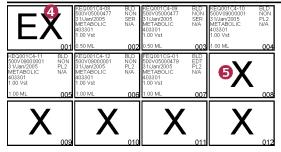
The Container report is a printable, graphic representation of the specimens within a storage container.

This report is useful for reconciling the storage locations assigned to specimens in LDMS and the actual real-life contents of a storage item. For example, if you initially assign specimens to a storage location in LDMS, you can print the container report and use it as a guide when inserting the actual specimens in the container.

Figure 35: Container report

Container Report

4 X 3 Box Legend: Global Spec ID Prim Specimen ID Add Spec/Harv Date Der Protocol / ID2 Sub-AD PID PID VID Vid Unit Other Spec ID Prim Specimen ID Add Spec/Harv Date Der Protocol / ID2 Sub-AD PID VID Vid Unit Other Spec ID Prim Spec I



(1) Storage location of container, (2) Dimensions of container, (3) Legend showing how to interpret the information for a specimen, (4) An excluded position, (5) An empty location

The top-right corner of the container report contains a key that shows how to interpret the details of a specimen listed on the report.

# Storage templates

Templates allow you to add commonly used storage items quickly by defining them in advance.

Template allow you to setup commonly used storage items, such as a box that your laboratory uses, and re-use the template as often as needed.

If you need to add a storage item that is a slight variation of an existing template, you can modify the template while creating the new storage item. For example, if you need to create a storage container that has an excluded corner that is not in the template, you can add it manually without needing to modify the template.

## **Creating storage item templates**

Storage item templates can be created for storage units, levels, and containers.

### **Background**

These templates can be selected when adding storage items to LDMS to save time.

### Steps

- Hover the mouse pointer over Storage from the LDMS menu bar, and then click either Storage unit templates, Level templates, or Container templates, depending on the type of template you want to create.
- **2.** Enter a name for the template into the **Name** box.

This is the name for the template as it will appear when creating new storage items.

- **3.** (For storage units only) Select the type of storage unit from the **Type** box and the temperature from the **Temperature** box.
- **4.** Enter the number of rows and columns for storage items based on this template into the **Number of rows** and **Number of columns** boxes.

This represents the size and capacity of new storage items based on this template in terms of the number of levels or containers that it can hold. For example, if you are creating a shelf that can hold 5 boxes side-by-side, you might have 5 columns and 1 rows.

**5.** Select or deselect the **Positions only** option as desired.

If the positions only option is selected, each item in storage items based on this template will be identified by a number. If it is not selected, each storage item will be identified by its coordinates.

- **6.** (If **Positions only** is not selected) Define how *levels* or *containers* in storage items based on this template should be identified.
  - **6.1.** Select either Row/Column or Column/Row from the Coordinate order box.

If Row/Column is selected, each *level* or container position will be identified in the format (row,column)

**6.2.** Select how columns and rows are to be labeled from the **Column labeling** and **Row labeling** boxes.

To use letters to identify rows or columns, select Alphabetic; to use numbers, select numeric.

**7.** Select the fill order for storage items based on this template from the **Fill order** box.

Fill order can be used when adding a new level or container to storage items based on this template. While you must specify a fill order, you can override it when add storage items.

**8.** Optional: Select a position(s) in storage items based on this template to be excluded from the **Excluded positions** box.

Excluded positions are used to help identify the orientation of the container (meaning which side is the front and which is the back).

- **9.** Review the **Preview** for storage items based on this template.
- **10.** Optional: (For storage units only) Add default levels for new storage units based on this template.
  - 10.1.Click the Add level button.

The Add Level window will open.

**10.2.**Select a template from the **Saved Templates** box.

The information for the template will be displayed, however you will not be able to modify it.

- **10.3.** Specify how many levels to add in the **Number to add** box.
- **10.4.**Click the **Continue** button.

The **Select Position for Level** page will open.

**10.5.**Select a position for the first storage item you are adding from either the **Position** box or on the **Preview**.

If you are adding more than one item, you can select the **Auto-fill all from selected position**. This option will place the remaining storage items based on the fill order of the storage item containing it. If you do not select this option, you will be prompted to manually position each storage item.

### **10.6.**Click the **Continue** button.

The storage item(s) will now appear in the **Default Levels** list.

You can add sub-levels and containers to the default levels that you've added as needed.

**11.** Click the **Save template** button.

## Modifying and removing storage templates

Storage templates can be modified or removed after they have been created.

### **Background**

Changes to templates will only affect new storage items based on the template. Existing storage items that were based on the template will not be updated.

### **Steps**

- Hover the mouse pointer over Storage from the LDMS menu bar, and then click either Storage unit templates, Level templates, or Container templates, depending on the type of template you want to modify.
- Select the template to modify from the Saved Templates box.
  - Note: If you want to clear the screen so that you can create a new template instead of modifying the selected template, click the **Add new** button.

The settings for that template will be displayed.

**3.** Modify the template as needed.

If you want to remove the template, click the **Delete template** button. Deleting a template makes it unavailable to be used for new storage items. Existing storage items that were based on the template will not be affected.

**4.** Click the **Save template** button.

#### Result

The template will modified and any new storage items based on it will reflect the changes. Existing storage items that were based on the template will not be modified.

# Moving items in storage

Storage items can be moved to a different location or rearranged in their current location.

### **Background**

Any storage item can be moved to another location in LDMS. Moving a storage item will require you to specify its position in its new location. If you move a storage item from its current location to its current location, you will be given the opportunity to rearrange its contents.

### **Steps**

- Click Storage > Stored Samples from the LDMS menu bar.
   Alternatively, you may use a barcode scanner to select the samples to be moved.
- Click Move this item from the action menu.
   The Move Selected to Storage Container window will open.
- **3.** From the list of storage containers, click the **Select** button to the right of the location where the selected item(s) should be moved to. The amount of free space is listed next to each container.

The storage items selected must be moved to the same destination. You cannot move two storage items at once that cannot occupy the same location. For example, you cannot select to move a specimen and a level at the same time.

The **Select Positions** window will open.

- 4. For each item being moved, select a position in the destination.

  If you want to automatically place the items being moved into the destination, select a position and then select the **Auto-fill all from selected position** option. This will cause all remaining storage items to be placed into the destination based on the destination's fill order.
  - Note: If samples are moved to a container that will not hold all selected samples, the remaining samples will remain selected to be moved to a different container.
- **5.** Click the **Continue** button.

## **Moving Items in Storage - Filter Search**

### **Background**

Stored items can also be selected to be moved using various filters.

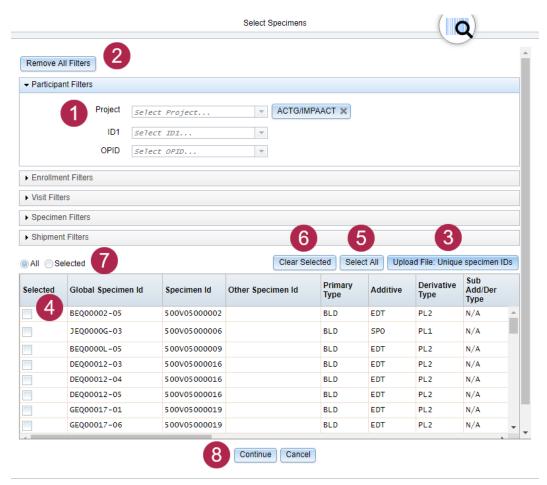


Figure 36: Specimen Filter Screen

(1) Various filters that can be applied to the list of specimens, (2) remove all specimens filters, (3) upload a unique specimen IDs file, (4) manually select specimens from filtered list to be moved, (5) select all specimens in filtered list to be moved, (6) clear selected specimens in filtered list, (7) filter list of specimens by selected only, (8) continue to Move Specimens screen

### Steps

- 1. Navigate to the **Stored Samples** page under the **Storage** module.
- In the Actions dropdown, select Move Stored Specimens Filter Search. The Select Specimens screen will open.
- **3.** Use the various filters to locate the specimens that need to be moved. The list of filtered specimens will appear below.
- 4. If not all specimens in the filtered list are intended to be moved, select the specimens that are to be moved and click the **Selected** radio button.
- **5.** A global specimen IDs file can also be added to the list of filtered specimens by clicking **Upload File: Unique specimen IDs**.
- **6.** Click **Continue**.

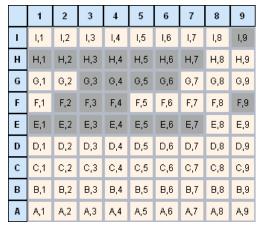
- **7.** Use the dropdown menus on the following screen to find the location the specimens will be moved to and click **Select**.
- **8.** Click **Continue** to confirm the move.

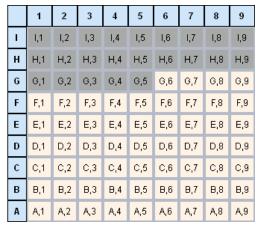
## Consolidating storage containers

Consolidating a storage container automatically moves specimens within a container based on its fill order to eliminate empty spaces.

### **Background**

Figure 37: Container consolidation





The effect of container consolidation before (left) and after (right).

### Steps

- **1.** On the navigation menu, click **Storage**.
- Locate the storage container to consolidate, and then click Edit.
- **3.** Below the Preview section, click **Consolidate Container**.
- **4.** At the bottom of the Edit Storage Container window, click **Save**.

## Listing specimens that moved in storage

After moving specimens in LDMS's storage, you can print a list of these changes so that you can move the actual specimens.

### **Background**

When specimens are moved in LDMS they also need to be moved in your actual specimen storage area so that the storage location assignments in LDMS remain accurate. To help you keep LDMS and your storage in sync, you can print a list of specimens that have been assigned new storage locations in LDMS.

### Steps

1. In the menu bar, click **Storage** > **Stored Samples** 

- 2. From the action menu in the upper-left corner, click **Print Move Report**.
- **3.** Next to **Generate for**, select one of the following:
  - Select **This Login Session** if you want to see all specimens that you have moved while signed in.
  - Select **Date Rage** if you want to see all specimens that have been moved during a given date range.
- **4.** If using a date range, specify the **Start Date** and **End Date**
- 5. Leave the File Type as PDF (\*.pdf).
  This report is not intended to be generated in a format other than PDF.
- **6.** Click **Generate Move Report**.

### After you are finished

Each row in the *Storage Move Report* will represent one specimen that has changed locations, with its original location and its now location. Use the report to find the specimen in its original location so that it can be related to the new position.

# Removing items from storage

Storage items and specimens can be removed from storage

### **Background**

Removing a storage item removes all of the items that it contains. If the location being removed contains specimens, you will be given the opportunity to change their condition and apply comments to them, which can be accessed when viewing the specimens.

Removing a storage item that contains specimens does *not* delete the specimens. They will still be visible on the **Specimen Management** page, and (unless you chose to make them *unavailable*) they can be added to a different storage container.



**Warning:** Removing a storage item cannot be undone.

### **Steps**

- 1. Click **Storage** from the LDMS menu bar.
- 2. Find the storage item or specimen that you want to remove.

  Use the filters on the left side of the **Storage** page to help locate the items to be removed.
- **3.** From the **Edit** combo button to the right of the item, select **Delete**.
  - Note: For specimens, this option will be called **Remove** instead of Delete.

The **Delete [Item]** window will open.

**4.** Optional: If you want to change the specimens that are stored at the location being removed to *unavailable*, select the **Unavailable** check box and add a date to the **Unavailable Date** field.

This means that the specimens will not be available to add to storage again. They will also not appear as available to ship when shipping specimen data to another laboratory. You will still be able to find the specimens, however, on the **Specimen Management** page.

If you do select this option, you will also be prompted to select a condition for the specimens and enter a comment. This comment will be applied to each specimen.

5. Click the **Delete** or **Remove** button.

# Modifying the condition of stored specimens

The condition of all specimens in a specific storage location can be modified at once.

### **Prerequisites**

The storage location must not contain any specimens that are part of a pending shipment.

### **Background**

There will be occasions when you need to update the condition of all the specimens in a storage location at once. This might happen, for example, if there was an equipment failure and all of the specimens in a storage unit were thawed.

### **Steps**

- 1. Click **Storage** > **Stored Specimens** from the LDMS menu bar.
- **2.** Locate the storage item that contains the specimens you want to modify.

If you select a higher-level storage item, such as a *storage unit* or *storage level*, the specimens in all sub-levels and containers will be modified as well.

**3.** From the edit menu to the right of the storage item, click **Set** condition code.

Figure 38: The Edit container menu



The **Set Condition Code** window will open.

- **4.** Select the condition code to apply from the **Specimen condition** box.
- **5.** Enter additional information about the specimens into the **Comments** box.

Information that is typically included here includes details for specimens that were not collected, an explanation of a condition, or an explanation of why a specimen is not available.

**6.** Enter additional information about the specimens into the **Internal-only comments** box.

Comments entered here are for your laboratory's use only. These comments will not be included if the specimen's information is shipped to another laboratory.

**7.** Click the **Save** button.

#### Result

The condition code, comments, and internal-only comments for all of the specimens in the storage location will be updated as you specified.

# **Storage Action Report**

The Storage Action Report shows you changes that have been made to your storage structure, such as adding specimens or containers.

### **Steps**

- 1. Click **Storage** > **Stored Samples** from the LDMS menu bar.
- 2. From the Action Menu, click **Print Storage Action Report**
- **3.** Select the appropriate radio button for the **Generate for** field:
  - **This Login Session**: the report will display all changes made in Storage during the current login session.
  - Events Since Your Last Printing of Report: the report will display all changes made in Storage since the last time the Storage Action Report was run.
  - **Date Range**: the report will display all changes made in Storage during a selected date range. Select the date range using the **Start Date** and **End Date** fields.
- **4.** Select the criteria for actions that will appear on the report.
- 5. Click Generate Storage Action Report.

# Shipping

The **Shipping** page in LDMS is where you create shipping data files and receive shipping data files from other laboratories.

The shipping page is organized into three sections:

**Pending** This page is where you create new data shipments, view shipments

and modify shipments that have been created but not yet

sent, and generate shipping files.

Receive shipments This page is where you will import shipping data files.

Shipment history

This page shows a record of all shipping files, created and received, for your laboratory. Shipments that are still

pending will not be shown.

# LDMS shipping file compatibility

LDMS shipping files are compatible with certain version of LDMS for Windows.

The format of the shipping files used by LDMS is an encrypted XML data file. LDMS for Windows support will vary based on the version of LDMS for Windows.

LDMS for Can ship and receive shipping files with laboratories

Windows using LDMS without restriction

version 10.0 or

higher

LDMS for Can ship specimens to but not receive specimens from

Windows LDMS

version 9.x

LDMS for **Windows** 

version 8.x or

earlier

# No shipping compatibility with LDMS

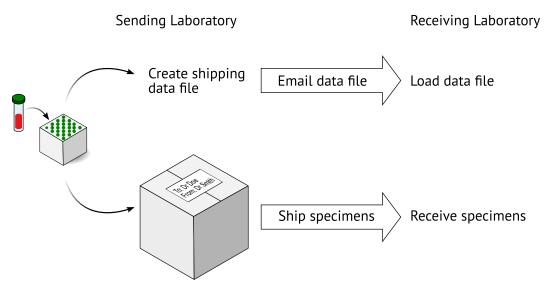
# Shipments are data transfer

Shipping in LDMS is the process of transferring specimen information from one laboratory to another.

This is a parallel process to physically packaging and shipping specimens to another laboratory. As specimens are packaged, you will specify in LDMS that they are to be shipped. You will then generate a data file with the information for those specimens called a shipping file. Once created, you provide that file to the laboratory that will receive the specimens. In this sense, it is more accurate to think of "shipping" in LDMS as "data transfer" between two laboratories.

LDMS does not assist with the physical shipment of specimens. LDMS does not create postal labels for shipping boxes, track the progress of shipments of couriers, and other logistical tasks. Laboratories need to implement their own solutions for the actual shipping process.

Figure 39: Shipping vs data transfer



This figure illustrates how creating a shipping data file in LDMS and transferring it to a laboratory is a separate process from physically boxing and shipping the specimens to the recipient.

There are two ways to ship specimens using LDMS:

- **1.** You can select individual specimens to be shipped, and then tell LDMS how you are going to put them in a shipping container.
- **2.** You can assign the specimens to a container on the **Storage** page, then ship the container.

Both of these methods can be used together within the same shipment. Sending a container from LDMS storage is generally preferred over creating ad hoc shipping containers. By using a container that has already been organized and checked, you will save time and reduce the possibility of sending the wrong specimens.

# Shipment numbers

Shipment numbers are assigned to sent and received shipments as a way to identify it at your laboratory.

When a new shipment is created or received, it is assigned the next available shipment number by LDMS. For example, if the last shipment that you received had the shipment number 152, the next shipment (whether you receive it from another laboratory or create it to send) will be assigned the shipment number of 153.

Shipment numbers are *not* unique between laboratories, nor will they be the same at the laboratory that sent and shipment and the laboratory that received it. For example, if you create a shipping file, it may be assigned the number 153 at your laboratory. When the receiving laboratory receives the shipping data file, it will be assigned the next available shipment number at

their laboratory, which will probably be different from the shipment number at your laboratory.

For this reason, shipment numbers cannot be directly matched to shipments between laboratories. Instead, you would need to use the shipment setup date and other information to match a shipment that you sent to the same shipment received at another laboratory.

# **New shipments**

New shipments are created on the **Pending Shipments** page

The **Pending Shipments** page shows all data shipments that have been created but not yet sent. This page serves the following purposes:

- 1. Create new shipments
- 2. View or modify shipments that have been created but not yet sent.
- **3.** "Ship" a shipment.

Figure 40: The Pending Shipments page

| Shipment<br>Number | Shippment<br>Format | Destination<br>Lab | Shipment<br>Temperature | Setup Date  | Ship Date   | QA/QC                   |             |
|--------------------|---------------------|--------------------|-------------------------|-------------|-------------|-------------------------|-------------|
| 119                | LDMS                | 300                | Dry Ice                 | 08/Sep/2015 | 08/Sep/2015 | Not Performed           | Edit/Ship 🔻 |
| 118                | LDMS                | 1                  | Dry Ice                 | 02/Sep/2015 | 02/Sep/2015 | Not Performed           | Edit/Ship ▼ |
| 117                | LDMS                | 350                | Ambient                 | 02/Sep/2015 | 02/Sep/2015 | Not Performed           | Edit/Ship 🔻 |
| 116                | LDMS                | 350                | Ambient                 | 02/Sep/2015 | 02/Sep/2015 | Not Performed           | Edit/Ship ▼ |
| 115                | LDMS                | 350                | Cold Packs              | 02/Sep/2015 | 02/Sep/2015 | Not Performed           | Edit/Ship ▼ |
| 113                |                     | 499                |                         | 13/Aug/2012 |             | Not Performed           | Edit/Ship 🔻 |
| 108                |                     | 999001             |                         | 01/Sep/2011 |             | Not Performed           | Edit/Ship ▼ |
| 102                |                     | 17                 | Dry Ice                 | 11/Jul/2011 |             | Complete With<br>Errors | Edit/Ship 🔻 |
| 66                 |                     | 999001             | Cold Packs              | 05/Apr/2010 |             | Not Performed           | Edit/Ship 🔻 |
| 55                 |                     | 300                | Cold Packs              | 25/Mar/2010 |             | Not Performed           | Edit/Ship 🔻 |

The term "ship" in LDMS means that all of the specimens you want added to a shipment have been added and it is ready to be sent. When you ship a shipment, you will be prompted to save a shipping data file—this file is the shipment. In addition to sending a physical package of specimens to the receiving laboratory, you will need to provide the laboratory with the shipping data file. This is most commonly done using email, but you could also put the file on a disc or re-writable USB drive and include it with the physical shipment.

## Creating new shipments

Creating a shipment means defining which specimens will be sent and where they are going to be sent.

### **Prerequisites**

The specimens must have already been entered into LDMS

• If you want to ship a storage container with specimens, you must have already assigned the specimens to the container in LDMS.

## **Background**



**Note:** You can save an incomplete shipment (such as leaving the destination blank), however you will need to complete the shipment before it can be sent.

### Steps

- Click **Shipping** > **Pending Shipments** from the LDMS menu bar. 1. The Pending Shipments page will open.
- 2. From the LDMS action menu, click **Create Shipment**. The **Pending Shipment Preview** window will open.
- 3. Complete the information on the **General** tab.

All information on this tab is required.

shipment date This is the date that the physical shipment was (if it was already shipped) or will be shipped. If you do not know when you will be shipping the specimens, select an approximate date—it can be changed before the shipping file is generated.

format

This is the type of shipping file you are creating. Unless the destination does not use LDMS, select LDMS.

temperature

This will be the method of temperature control for the specimens during transport.

4. Complete the information on the **Shipping Destination** tab.

> Existing laboratories that are using LDMS and LDMS are already available for selection. The contact information for these laboratories is maintained by Frontier Science.

- **4.1.** Select either **Contact**, **Lab Number**, or **Lab Name**.
  - This is how you will select the laboratory. **Contact** is the shipping contact person on file for the laboratory. Lab Number is the laboratory's LDMS ID. Lab Name will show each laboratory in alphabetical order.
- **4.2.** Select a laboratory from the drop-down box below. If you selected **Contact** or **Lab Name**, the number in parenthesis next to each laboratory's name is the laboratory's LDMS ID number.
- **4.3.** Select or enter the person to receive the shipment at the receiving laboratory from the **Contact Person** box.

Laboratories can be added to LDMS. These will only appear as available for selection by your laboratory. Since all laboratories that use LDMS and LDMS are already listed, the new laboratory is assumed to be using some other laboratory management system.

- 4.1. Select New lab.
- **4.2.** Complete all the boxes in the **Shipping Destination** and **Contact Information** sections.
- **4.3.** Optional: If you want to save this laboratory for future use, select the **Save address** box.
- 5. Complete the information on the Contact at sending lab tab. This is where you enter the information for the person at your laboratory who should be contacted if there is an issue with the shipment.
- **6.** Optional: Complete the information on the **Shipment notes** tab.

The **Comment** and **Disclaimer** will appear on the shipping manifest and will be shown to the receiving laboratory when they load the shipping file. These comments are in addition to comments that were entered for individual specimens. Use the **Comment** box for general information about the shipment. The **Disclaimer** box can be used to provide information about the restrictions placed on the specimens' usage.

The **Shipment Carrier** and **Tracking Number** will *not* appear on the shipping manifest and are not included in the shipping file.

- **7.** Add specimens to the shipment on the **Shipment Contents** tab.
  - Add a container already setup on the **Storage** page to the shipment from the **Storage Containers** list.
  - Add specimens to a single use shipping container in the Shipping Containers section.
- **8.** Add storage containers to the shipment.
  - **8.1.** Click the **Add New** button to the right of the **Storage Containers** list.

The **Storage Containers** window will open.

- **8.2.** For each storage item to be shipped, select the check box next to it.
- 8.3. Click Add Selected to Shipment.
- **9.** Add individual specimens to the shipment.
  - **9.1.** Click the **Add New** button to the right of **Shipping Containers**. The **Create Shipping Container** window will open.
  - **9.2.** Enter the number of rows and columns for the container into the **Number of rows** and **Number of columns** boxes.

This represents the size and capacity of the container. For example, a container with 8 columns and 5 rows could hold up to 40 specimens. The container as it is currently defined will appear in the **Preview** section.

**9.3.** Select or deselect the **Positions only** option as desired.

If the positions only option is selected, each specimen in

If the positions only option is selected, each specimen in the shipping container will be identified by a number. If it is not selected, each specimen will be identified by its coordinates. **9.4.** (If **Positions only** is not selected) Define how specimens in the container should be organized.

**coordinate** This is how the specimens position will be **order** identified, such as (8,5) or (5,8).

column labeling and row labeling This is how rows and columns will be named in the container. Alphabetic uses letters while numeric uses numbers. The second option specifies where labeling should start.

**9.5.** Select the fill order for the shipping container from the **Fill order** box.

The fill order is the logic that is used to determine where specimens will be placed in the container.

**9.6.** Select the sort order for the container from the **Primary Sort Order** box.

This is how specimens will be sorted before they are put into the container.

- **9.7.** (If **Prot/ID2** was selected for primary sort order) Select a **Secondary Sort Order**.
- 9.8. Click the Add Specimens button.
- **9.9.** Do one of the following:
  - In the list of specimens, select the check box next to each specimen to be shipped. Use the available filters to find the specimens you need.

Or:

- Click the **Upload File: Unique specimen IDs** button to upload a file of samples to be included in the shipment.
- **9.10.**In the list of specimens, select the check box next to each specimen to be shipped.

Use the available filters to find the specimens you need.

**9.11.**At the bottom of the Select Specimens to Ship window, click **Continue**.

The **Preview** section for the shipping contain will indicate the positions that now hold specimens. If you added more specimens than could fit in the container, multiple containers will have been created. You can switch between these containers using the arrow buttons in the **Preview** section.

9.12.Click the Add button.

The shipping container will appear in the **Shipping boxes** list.

- **10.** Do one of the following:
  - To save the shipment so that it can be modified again before the shipping file is generated, click the **Save** button.
  - To generate the shipping file now and change the shipment status to shipped, click the **Ship** button.

### Result

If the shipment was saved, a new shipment will be added to the **Pending Shipments** page. It will appear at the top of the list and will have been assigned the next available shipment number.

### After you are finished

You should perform QA/QC on the shipment after it has been created before sending it.

## Sending a pending shipment

Sending a shipment is the process of generating a shipping data file on the **Pending Shipments** page.

### **Background**

The shipping file can either be in LDMS format or CSV format. The file that is created will need to be transferred to the laboratory to add the specimens to their database.

### **Steps**

- 1. Click **Shipping** > **Pending Shipments** from the LDMS menu bar.
- **2.** Locate the pending shipment to be sent.
- 3. Click the **Edit/Ship** button to the right of the shipment. The **Pending Shipment Preview** window will open.
- **4.** Click the **Ship** button.

LDMS will warn you that this is your last chance to generate the Shipment Storage Report. If you want to view this report, click the **Cancel** button. On the **Shipment Contents** tab, click the **Shipment Storage Report** button to generate the report.

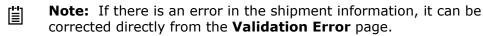
LDMS will check that the information for the shipment has been fully completed. If there are any issues, it will be highlighted in red and you will be prompted to correct it before continuing.

**5.** When asked to confirm if you want to send the shipment, click the **OK** button.

The shipping file will be generated. When it is ready, you will be prompted to save it to your computer.



**Warning:** Do not close the window until the shipping file has been saved.



- **6.** Click the **Close** button to go back to the **Pending Shipments** page.
- **7.** Optional: Generate a shipping manifest.
  - **7.1.** Click **Shipping** > **Shipment History** from the LDMS menu bar.
  - **7.2.** Locate the shipment that you sent.

**7.3.** Click the arrow next to the **View** button, and then click **Generate Manifest**.

Figure 41: Generate Manifest



### Result

The shipping file is now saved at the location you specified on your computer.

### After you are finished

This is the file you will need to transfer to the laboratory that will be receiving the specimens. You can either email this file to the receiving laboratory or provide it on removable media (such as a USB drive) with the physical shipment of specimens.

## Generating a shipping manifest

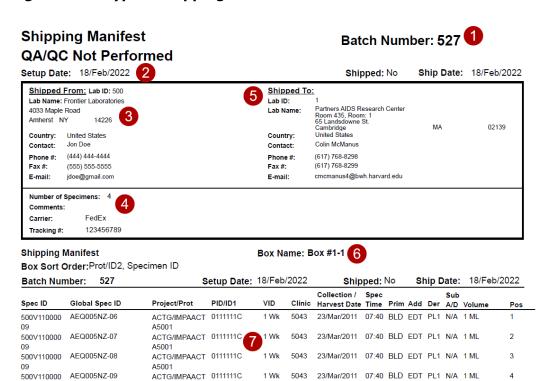
Shipping manifests contain an overview of the contents of a shipment, and are intended to be printed and included with shipments.

### **Background**

A shipping manifest should be printed and included with all shipments. This will help the laboratory receiving the shipment to identify its contents and match it to the shipping file that you provided to them. You can also save the manifest as a PDF file, and then send it to the receiving laboratory long with the shipping file.

The shipping manifest can be generated before or after a shipment is sent.

Figure 42: A typical shipping manifest



(1) The shipment number, (2) The date that the shipment was initially created, (3) Sending laboratory's contact information, (4) Shipment carrier and tracking information (5) Receiving laboratory's contact information, (6) Name of shipping container, (7) Specimens in the shipping container

### Steps

- **1.** Do one of the following:
  - For pending shipments, click Shipping > Pending Shipments from the menu bar.
  - For sent shipments, click Shipping > Shipment History from the menu bar.
- Locate the shipment for which you want to generate a manifest.
- Click the arrow next to the View button to the right of the shipment, and then click Generate Manifest.
- **4.** Select PDF from the **File type** box.

A5001

Because manifests are intended to be printed, they have been designed with PDF output in mind only. If you select a different file format for the manifest, you may experience formatting issues.

### Result

You will be prompted to save the manifest or it may open automatically, depending on the default behavior of your web browser.

# **Shipment Tracking Information**

Information about a shipping carrier and the tracking number can be entered into LDMS at any point by both a sending a receiving laboratory.

The **Shipping Carrier** and **Tracking Number** are on the **Shipment Notes** tab when viewing a shipment. This information can be updated at any time for any shipment, including shipments that have already been sent or received by your laboratory.

Shipment tracking information will not appear on printed shipping manifests and will not be included in shipping files.

# Supported shipping file formats

These are the file formats that are supported for sending and receiving shipping files.

Table 14: Supported shipping file formats

| Format              | Encrypted? | Intended destination  |
|---------------------|------------|---|
| LDMS                | Yes        | Laboratories using LDMS (both web and windows version)            |
| CSV                 | No         | Laboratories not using LDMS                                       |
| SeraCare            | No         | Specimens being sent to<br>the Precision Bioscience<br>Repository |
| Cross-LIMS Manifest | No         | Laboratories not using LDMS                                       |

## **CSV** shipping files

LDMS can generate and accept CSV (comma separated value) shipping files that meet these requirements.

- The file must be a comma delimited text file with the .csv file extension.
- The first line in the file must be a header row.
- The file must contain, at minimum, all required columns (in any order) to be processed by LDMS.
- When a CSV shipping file is generated by LDMS, all columns listed in the following table will be included.



**Note:** Field headers in the file are not case-sensitive.

| Column<br>Label    | Example<br>Value | Column<br>Required<br>in File | Must be<br>Defined in<br>File | Notes   |
|--------------------|------------------|-------------------------------|-------------------------------|---|
| Shipment<br>Number | 145              | No                            | No                            | ID number for the shipment.                             |
| Sending Lab        | 500              | Yes                           | Yes                           | ID number for the laboratory sending the shipping file. |

| Column<br>Label     | Example<br>Value  | Column<br>Required<br>in File | Must be<br>Defined in<br>File | Notes   |
|---------------------|-------------------|-------------------------------|-------------------------------|---|
| Receiving<br>Lab    | 143               | No                            | No                            | ID number for the laboratory receiving the shipping file.   |
| Setup Date          | 05/<br>Jan/2016   | No                            | No                            | Date shipping file was created. Must be in the format dd/Mmm/YYYY.  |
| Ship Date           | 05/<br>Jan/2016   | No                            | No                            | Date specimens were shipped. Must be in the format dd/Mmm/YYYY.   |
| Temperature         | Dry Ice           | No                            | No                            | Temperature of the shipment.  |
| Shipment<br>Comment | This is a comment | No                            | No                            | Notes about the shipment.   |
| Container           | Box #1-1          | No                            | No                            | Name of container holding specimen.   |
| Row                 | 1                 | No                            | No                            | Position of specimen in container. Alpha and numeric coordinates are allowed.   |
| Column              | 1                 | No                            | No                            | Position of specimen in container. Alpha and numeric coordinates are allowed.   |
| QA<br>Performed     |                   | No                            | No                            | Whether shipping file was compared to the actual shipment to ensure specimens were correct and included in the correct container locations. |
| project             | FRONTIER          | Yes                           | Yes                           | Name of project for which the specimen was collected.   |
| ID1                 | 0111111C          | Yes                           | Yes                           | Participant or specimen source ID.  |
| ID2                 | F5309             | Yes                           | Yes                           | Protocol ID.  |
| ID3                 |                   | Yes                           | No                            | Sub-protocol ID.  |
| Visit               | 1                 | Yes                           | No                            | Visit value.  |
| Visit Unit          | WK                | Yes                           | No                            | Unit for visit. Must be a valid LDMS visit unit code.   |
| Clinic              | 701               | Yes                           | No                            | Clinical site where specimen was collected.   |
| Specimen<br>Date    | 05/<br>Jan/2016   | Yes                           | Yes                           | Collection date. Must be in the format dd/Mmm/YYYY.   |
| Specimen<br>Time    | 15:20             | Yes                           | No                            | Collection time. Must be in 24-hour format.   |
| Received<br>Date    | 05/<br>Jan/2016   | Yes                           | Yes                           | Date specimen was received by laboratory. Must be in the format dd/ Mmm/YYYYY.  |
| Received<br>Time    | 15:20             | No                            | No                            | 24-hour time specimen was received by laboratory. Must be in 24-hour format.  |
| Specimen<br>ID      | 500V050000        | )1 <b>½</b> lo                | No                            | Specimen ID (used for specimens entered in LDMS for Windows only).  |
| Global Spec<br>ID   | CEQ0000P-0        | )3No                          | No                            | If not specified, a global specimen ID will be generated automatically when the shipping file is received in LDMS.                          |

| Column<br>Label             | Example<br>Value  | Column<br>Required<br>in File | Must be<br>Defined in<br>File | Notes   |
|-----------------------------|-------------------|-------------------------------|-------------------------------|---|
| Other Spec<br>ID            |                   | No                            | No                            | Other specimen ID.  |
| Primary                     | BLD               | Yes                           | Yes                           | Specimen primary type. Must be a valid LDMS primary code.   |
| Additive                    | EDT               | Yes                           | Yes                           | Specimen additive. Must be a valid LDMS additive code.  |
| Derivative                  | PL2               | Yes                           | No                            | Specimen derivative. Must be a valid LDMS derivative.   |
| Sub A/D                     | N/A               | No                            | No                            | Specimen sub-additive/derivative. Must be a valid LDMS sub-additive/derivative code. If not specified, will default to N/A. |
| Volume                      | 1.5               | Yes                           | Yes                           | Volume of specimen  |
| Volume<br>Units             | ML                | Yes                           | Yes                           | Units used for specimen volume.   |
| Condition                   | SAT               | No                            | No                            | Three letter code representing specimen's current condition. If not specified, will default to SAT.                         |
| Comments                    | This is a comment | No                            | No                            | Notes about the specimen  |
| Tests                       |                   | No                            | No                            | Assays assigned to the specimen   |
| Processing<br>Date          | 05/<br>Jan/2016   | No                            | No                            |   |
| Processing<br>Time          | 15:20             | No                            | No                            |   |
| Frozen Date                 | 05/<br>Jan/2016   | No                            | No                            |   |
| Frozen Time                 | 15:20             | No                            | No                            |   |
| Total Cell<br>Count         |                   | No                            | No                            |   |
| Processing<br>Tech          |                   | No                            | No                            |   |
| Second<br>condition<br>code |                   | No                            | No                            |   |
| Freezer                     |                   | No                            | No                            |   |
| Level 1                     |                   | No                            | No                            |   |
| Level 2                     |                   | No                            | No                            |   |
| Harvest<br>Date             | 05/<br>Jan/2016   | No                            | No                            |   |
| Additional<br>Time          | 15:20             | No                            | No                            |   |

| Column<br>Label         | Example<br>Value  | Column<br>Required<br>in File | Must be<br>Defined in<br>File | Notes  |
|-------------------------|-------------------|-------------------------------|-------------------------------|--|
| Additional<br>Time Unit |                   | No                            | No                            |  |
| Thaw Count              |                   | No                            | No                            |  |
| Internal comments       | This is a comment | No                            | No                            | Import this field if it's in the file, but do NOT ship it out from WebLDMS   |
| Reason not collected    |                   | No                            | No                            |  |
| Primary<br>Database ID  | 12345             | No                            | No                            | Primary Specimen's Database ID value. Must be sourced from a primary specimen that already exists in the database. Setting this value will ensure new aliquots are organized under the specified primary specimen provided other dependent values are properly matched. Value can be determined from Custom Report Builder or from the primary's Edit Primary Specimen popup URL in Specimen Management. |

## **Cross-LIMS shipping files**

LDMS can generate and accept cross-LIMS shipping files that meet these requirements.

- The file must be a tab delimited text file with the .txt file extension.
- The first line in the file must be a header row.
- The file must contain, at minimum, all required columns (in any order) to be processed by LDMS.
- When a cross-LIMS shipping file is generated by LDMS, all columns listed in the following table will be included.
- If additional columns are present in a cross-LIMS file, those columns will be ignored.
- Cross-LIMS shipping import sets the clinic value for VTN records based on the PID. This allows LDMS labs to receive cross-LIMS shipments of VTN samples if needed.

| Column Label | Example Value      | Required | Notes   |                    |
|--------------|--------------------|----------|---|--------------------|
| SHIP_ID      | 0500-0999-00000014 | 7Yes     | In the format [Sending_Lab_ID]-Receiving_leading zeros must be added to make the ID the correct length (for example 500-19-147 is not valid, but 0500-0019-0000000147 is valid) | _Lab_ID]-[Shipment |
| SHIP_DATE    | 06-Jan-16          | No       | Date shipped. Date<br>must be in the format<br>dd-Mmm-yy  |                    |

| Column Label | Example Value   | Required | Notes  |
|--------------|-----------------|----------|--|
| RECIPIENT    | 999             | Yes      | Receiving laboratory's ID  |
| SHIPPED_FROM | 500             | Yes      | Sending laboratory's ID  |
| GLOBAL_ID    | GEQ00017-03     | No       | A valid LDMS global specimen ID  |
| project      | FRONTIER        | Yes      | The project for which the specimen was collected.  |
| PROTOCOL     | F5309           | Yes      | The protocol (ID2) for which the specimen was collected  |
| PID          | 0777777F        | Yes      | The participant identifier (ID1)   |
| VID          | 7               | Yes      | The visit value  |
| VID_UNIT     | Day             | Yes      | The visit unit. Must<br>be a valid LDMS visit<br>code  |
| COLL_DT_TM   | 17-Jan-05 09:12 | Yes      | Collection date and time, in the format dd-Mmm-yy HH:mm, using 24-hour clock                   |
| PRIM         | BLD             | Yes      | Primary type. Must be a valid LDMS primary code.   |
| DER          | PL2             | Yes      | Derivative type. Must<br>be a valid LDMS<br>derivative code.                                   |
| SUBDER       | N/A             | Yes      | Sub-additive/<br>derivative type. Must<br>be a valid LDMS sub-<br>additive/derivative<br>code. |
| ADD          | EDT             | Yes      | Additive type. Must be a valid LDMS additive code  |
| QTY          | 1               | Yes      | Volume of the specimen collected.  |
| QTY_UNIT     | ML              | Yes      | Volume unit. Must be a valid LDMS volume unit code.  |
| CONDITION    | SAT             | No       | Condition of the specimen. Must be a valid LDMS condition code.                                |

| Column Label | Example Value     | Required | Notes  |
|--------------|-------------------|----------|--|
| OTHERSPECID  | VTN               | No       | Other Specimen ID. Must be alpha/ numeric with a max field length of 17 characters.              |
| TIME         | 0.00              | No       | Must include the Additional Time. Format must be numeric/decimal with precision to two decimals. |
| TIMEUNIT     | HRS, TR, or RAN   | No       | Must include Additional Time Unit Abbreviation. The field must be three characters long.         |
| COMMENT      | This is a comment | No       | A comment about the specimen.  |
| BOX          | #1-1              | No       | Name of shipping container   |
| ROW          | 1                 | No       | Row in shipping container (from top-to-bottom)   |
| COL          | 1                 | No       | Column in shipping container (from left-to-right)  |

# SeraCare shipping files.

LDMS can generate (but not receive) "SeraCare" shipping files that meet these requirements.

| Column Label    | Example Value  | Notes   |
|-----------------|----------------|---|
| Global Spec ID  | 0777-001MWK00- | 286   |
| project         | FRONTIER       | The project for which the specimen was collected. |
| PID             | 0111111C       | Participant ID (ID1)                              |
| VID             | 8              | Visit value.                                      |
| VID Unit        | Vst            | Visit unit.                                       |
| Collection Date | 1-Mar-13       | Collection date                                   |
| Collection Time | 1:00           | Collection time.                                  |
| Prim            | BLD            | Primary type.                                     |
| Der             | BLD            | Derivative type.                                  |
| Add             | ACD            | Additive type.                                    |
| Other Spec ID   | PLORQ          |   |
| Volume          | 1              |   |
| Volume Unit     | ML             |   |
| Volume Unit     | ML             |   |

| Column Label    | <b>Example Value</b> | Notes  |
|-----------------|----------------------|--|
| Custom Local ID |                      |  |
| Box             | BOX 2                | Name of shipping container.                              |
| Row,Col         | 1,1                  | Position in shipping container.                          |
| Condition       | SAT                  |  |
| Comment         | This is a comment    |  |
| Protocol        | F5308                | The protocol (ID2) for which the specimen was collected. |
| Sub/Der         | N/A                  | Sub-additive/derivative type.                            |

# **Shipping container report**

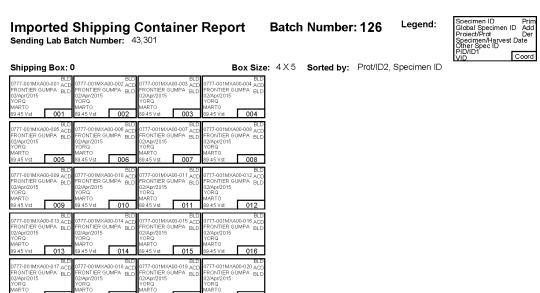
This report shows a visual map of each container's contents, along with the details for each specimen being shipped.

The shipping container report can be generated while you are creating a shipment or after a shipment has been sent.

The shipping container report can be found on either the **Pending Shipments** or **Shipment History** page. To the right of the shipment, click the down arrow next to the button and click **Shipping Container Report**.

There is a key in the top-right corner of the report that shows what information is within each box. Each box on the report corresponds to one location in the container. Locations that are marked with a large X indicate locations in the container that are empty.

Figure 43: The Shipping Container Report



# **Shipment storage report**

This report lists the specimens in a shipment and their assigned storage location.

This report is intended to be printed and be used to help locate and pull specimens from storage. This would only be applicable if you assigned a storage location to the specimens being shipped on the **Storage** page.

You can generate the Shipment Storage Report when generating a shipping file. After a shipping file has been created, you can access the report from the **Shipment History** page by clicking the drop-down next to the **View** button for the shipment and selecting **Shipment Storage Report**.

## Information displayed on report

- · Shipment number
- Shipment setup date
- Specimen ID
- Global specimen ID
- Other specimen ID
- project/ID2
- ID1
- Visit
- · Collection date
- · Collection time
- Primary
- Additive
- Derivative
- Sub additive/derivative
- Volume
- Storage location
- Original storage location

# **Receiving shipments**

Receiving a shipment is the process of importing specimen information from laboratory into your laboratory.

### **Background**

Once you have the shipping file from the laboratory sending you the specimens, you can import the specimen information into LDMS.

### Steps

- 1. From the menu bar, click **Shipping** > **Receive Shipments**.
- Click Select File.

A window will open to select the shipping file. The shipping file will generally be named ship-[your laboratory ID number].xml for LDMS shipping files or lab[your laboratory ID number].csv for CSV files.

- Note: Supported ID formats include Specimen IDs, Other Spec IDs, Global Spec IDs, and Database IDs.
- **3.** Optional: If you want specimens to automatically be assigned storage locations, select **Import directly into storage**.
- 4. Optional: To apply RPID to the specimens being imported, select Import as RPID. This will anonymize the imported specimen data. This feature is disabled by default and must be activated by LDMS User Support, and the feature can only be applied to specimens for the ACTG or IMPAACT projects.
- 5. Click Preview Shipment.
- **6.** On the **Confirm Temperature** list, select the temperature of the shipment as you received it.
- **7. Optional:** Click the **Edit Condition Codes** button to update information of specimens on the shipping file before receiving the shipment.
  - **7.1.** After clicking the **Edit Condition Codes** button, the **Set Condition Codes** window will open.
  - **7.2.** Select specimen(s) from the table.
  - **7.3.** Apply condition codes, set as unavailable, or add comments as needed.
  - **7.4.** Click **Update Selected Specimens**.
  - **7.5.** When finished, click **Save**.
- **8.** Review the shipment to ensure it is correct.

It is a good idea to check that the shipping destination is your laboratory, and that the specimens are what you were expecting. Click the **View** button next to shipment containers to see what specimens are in the shipment. By checking the shipment at this stage, you can avoid potential errors, such as receiving the wrong shipping file.

9. Click Receive Shipment.



**Note:** If the shipping temperature you indicated differs from the temperature indicated in the shipping file, you will be asked if you want to override the shipping temperature. This means the temperature you selected will be saved in your database instead of the temperature from the sending laboratory.

### Result

LDMS will assign the next available shipment number to the shipment.

### After you are finished

Go to the **Shipment History** page to view the newly received shipment and perform additional actions, such as QA/QC.

## Shipping and projects

During shipping import, users will be notified if the file contains a new project that does not exist in their database. If the new project is not expected, the import can be cancelled.

## Import as is

When receiving a shipping file, it is possible to take shipping and storage items in the shipment and automatically assign them to your laboratory's storage. The option to do this is provided when receiving a shipping file.

The items on the shipment will be placed in your storage based on what it is.

| Item               | Where it will go in storage   |
|--------------------|---|
| Storage unit       | Added as a new storage unit   |
| Level or container | In the <b>SHIPPING IMPORT FREEZER</b> , on a level with the shipment number |

If the item was a storage item at the sending laboratory, it will retain its original name in your storage. If the item was a storage contain created for the shipment, it will be placed in the **SHIPPING IMPORT FREEZER** on a level with the shipment number. These containers will generally have names like "Auto-generated box #76-1" in your storage. The shelf name also includes the sending lab ID.

Once the items are added to your storage, you are not restricted to how you use them. You can retain containers, move them to different storage units, and so forth.

## Handling non-LDMS shipping files

Shipping files that did not originate from LDMS may not contain the global specimen ID needed by LDMS. Such specimens will receive special handling when they are received.

- All specimens without a global specimen ID will be treated like aliquot specimens and assigned to a new primary specimen.
- All specimens without a global specimen ID will be assigned one
- Specimens with the same project, ID1, ID2, visit, collection date, collection time, primary type, and additive time will be grouped until the same primary specimen.

# **Shipment QA/QC**

Shipment QA/QC is the process of comparing actual specimens in a shipping container to the specimen that is expected in that container position by LDMS. This should be done for shipments before they are sent and after they are received.

You can view the current QA/QC status of a shipment on the **Pending Shipments** or **Shipment History** page.

| QA/QC Status         | Action Needed                                     |
|----------------------|---|
| Complete             | None  |
| In Progress          | Finish QA/QC                                      |
| Complete with Errors | Investigate the specimens that did not pass QA/QC |
| Not Performed        | Start QA/QC                                       |

To perform QA/QC, you need to look at the global specimen ID on a physical specimen, and then compare it to the global specimen ID in LDMS. If they match, the specimen has passed, otherwise it has failed. QA/QC should be performed by scanning the specimen's barcode, which will read the global specimen ID and do the comparison for you. This is the recommended process. If you do not have a barcode scanner, you can manually compare the two global specimen IDs instead.

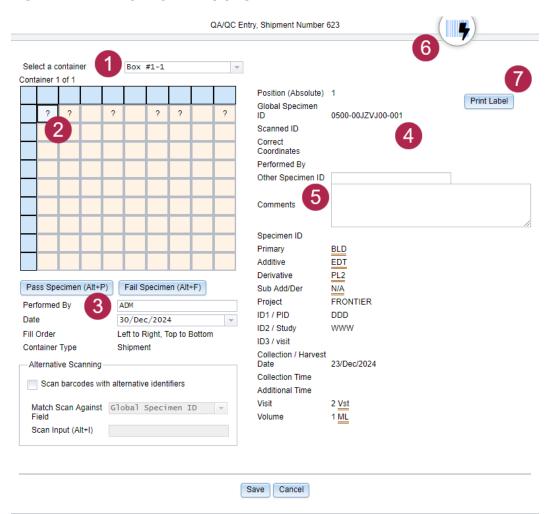


Figure 44: The QA/QC Entry page

(1) Containers in the shipment, (2) Currently selected specimen, (3) Manually pass or fail selected specimen, (4) Currently selected specimen's global specimen ID, (5) Modifiable boxes, (6) Indicator showing that you can scan a barcode to complete shipment QA/QC automatically, (7) Print the current label

If the shipment contains multiple containers, you will need to change containers during QA/QC, since each container must be reviewed separately.

While performing QA/QC, you have the option to modify each specimen's other specimen ID and comments. If you do so, these changes will be reflected in the specimen information on the **Specimen Management** page.

# **Performing QA/QC**

QA/QC is performed by scanning or comparing each specimen's label with the expected specimen information, and indicating if it is correct or incorrect.

### **Background**

QA/QC should be performed after creating a pending shipment but before creating the shipping file. It should also be performed on shipments from other laboratories after they have been received.

### **Steps**

- **1.** Do one of the following:
  - To QA/QC a pending shipment, on the navigation menu, hover over Shipping, and then click Pending Shipments.
  - To QA/QC a pending shipment, on the navigation menu, hover over Shipping, and then click Shipment History.
- 2. On the **Edit/Ship** (for pending) or **View** (for received) menu in the right column, click **QA/QC**.
- **3.** Optional: Enter the initials of the person that performed QA/QC and the date that it was completed in the **Performed By** and **Date** boxes.
- **4.** With the first specimen select (which is the default), do one of the following:
  - Scan the specimen barcode (recommended). After scanning the barcode, one of two sounds will play, depending on whether the sample passes or fails.
    - Note: Users can QA/QC shipments by scanning non-LDMS generated barcodes. The value in the scanned barcode must match to the value set in Other Specimen ID or ID1/PID field for QA to pass.
  - Manually compare the global specimen ID on the screen with the global specimen ID on the specimen label. If they match, click Pass Specimen, otherwise click Fail Specimen.
- (If there are multiple shipment containers) After each specimen in the container has been reviewed, select the next item from the Select a Container list.
- **6.** After all specimens in the shipment have been reviewed, click **Save** at the bottom of the page.

### After you are finished

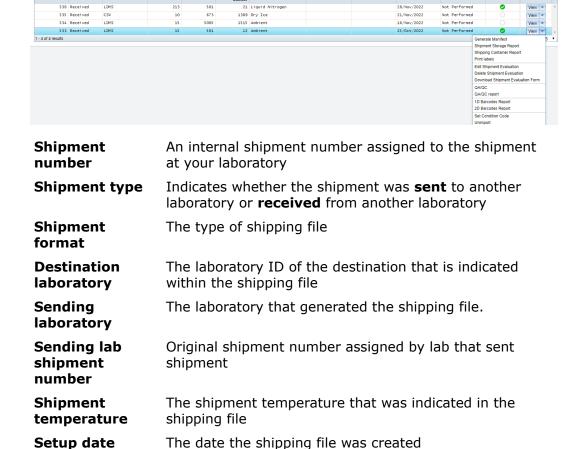
Confirm that QA/QC has been completed for the shipment by looking at the **QA/QC** column on the **Pending Shipments** or **Received Shipments** page.

# Shipment history

A record of shipments sent and received can be found on the **Shipment History** page.

In terms of LDMS, a shipment received is one for which you have uploaded a shipping data file from another laboratory in LDMS, and a shipment sent is when you have generated the shipping data file.

Figure 45: The Shipment History page



**QA/QC** This indicates whether or not QA/QC was performed on

The date that the shipping file was generated

The date that your laboratory loaded the shipping file in LDMS (only applicable to shipments that you received)

the shipping file

**Shipment** This Indicates whether or not a Shipment Evaluation has been performed for the shipment

#### Removing a received shipment

#### **Background**

Shipment date Received date

If a shipping file was received and loaded into LDMS in error, it may need to be removed. Because multiple laboratories may be affected and the quality of specimen data affected, the ability to unload a shipping file that was received requires you to contact LDMS User Support.

#### **Steps**

- Contact LDMS User Support.
   This should be done first because if your session is idle for too long and you are logged out, a new challenge code will be generated.
- 2. In the menu bar, click **Shipping** > **Shipment History**.
- **3.** On the **View** button to the right of the shipment, click the arrow and select **Unimport**.
- **4.** Provide LDMS User Support with the **Challenge Code**.
- 5. In the **Response Code** box, enter the code provided LDMS User Support.
- **6.** Click **Unimport**.

#### **Un-sending shipments**

After a shipping file has been generated and a shipment changed to sent in LDMS, it is possible to change the shipment back to a pending state.

#### **Background**

This would be necessary if you need to modify the shipment and re-generate the shipping file. This could happen, for example, if you discovered there were more specimens to be shipped and the shipment hasn't left your laboratory yet.



**CAUTION:** A shipment should not be un-sent if the receiving laboratory has received and loaded the shipping file. Doing so would cause the specimens to appear at both laboratories. Contact the receiving laboratory and ensure that have not yet loaded the shipping file before continuing.

#### **Steps**

- 1. In the menu bar, click **Shipping** > **Shipment History**.
- 2. On the **View** button to the right of the shipment, click the arrow and select **Unship**.
- 3. Click Unship.

#### Re-downloading the shipping file of a sent shipment

It is possible to re-download the shipping file of a previously sent shipment.

#### **Background**

When you send a pending shipment, you will be prompted to download the shipping file that you will need to supply to the receiving laboratory. If this file becomes lost or damaged, it is possible to download it again. This file will be identical to the shipping file that was previously downloaded.

#### Steps

1. In the menu bar, click **Shipping** > **Shipment History**.

- 2. On the **View** button to the right of the shipment, click the arrow and select **Download shipping file**.
- **3.** When prompted, save the shipping file.

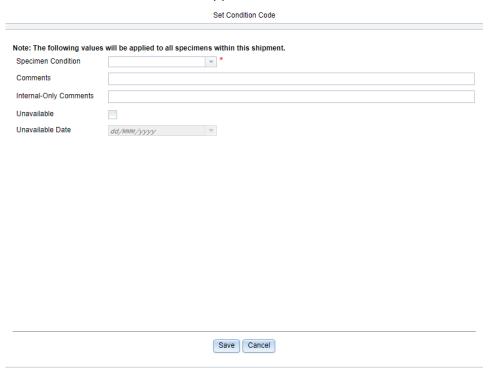
## **Bulk Updating Condition Code and Comments of Shipments**

Users have the ability to bulk update the Condition Code and Comments for all specimens in a shipment. This is accessed through the **Shipping** menu.

#### **Background**

#### Steps

- 1. From the menu bar, click **Shipping**, and then click **Shipment History**.
- **2.** Find the shipment that needs updating and click the arrow next to **View**.
- **3.** From the dropdown menu, click **Set Condition Code**. The **Set Condition Code** window will appear.

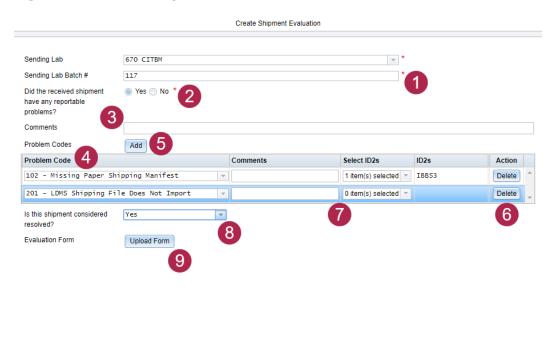


- **4.** In the new window, select the specimen condition from the dropdown menu.
- **5.** Add any comments or mark as unavailable as needed.
- 6. Click Save.

#### **Shipment Evaluations**

Users can create new shipment evaluations for received shipments and update existing ones.

Figure 46: Create Shipment Evaluation Screen





(1) Add or update the sending lab batch number, (2) indicate whether the shipment had any reportable problems, (3) add any comments, (4)\* select a problem code from a dropdown, (5)\* add more dropdowns for additional problem codes, (6)\* remove a listed problem code, (7)\* Associate comments and ID2s to individual problem codes, (8)\* indicate whether the shipment is considered resolved, (9) upload a shipment evaluation form, (10) create the shipment evaluation

#### **Creating a New Shipment Evaluation**

- 1. From the LDMS menu, navigate to **Shipping**, then select **Shipment History**.
- 2. In the dropdown menu of a received shipment, select **Add New Shipment Evaluation**.
- **3.** On the **Create Shipment Evaluation** screen, indicate whether the received shipment had any reportable problems, and enter a comment and attach an evaluation form as needed.

<sup>\*</sup>Only available if user selects Yes for the question Did the received shipment have any reportable problems?

- **4.** If the user indicates that the shipment did have reportable problems, at least one problem code must be added. The user may also indicate whether the shipment is considered resolved.
- 5. Click Create to create the evaluation.

#### **Editing an Existing Shipment Evaluation**

Users may edit or delete a shipment evaluation or evaluation form as needed.

- To update an existing Shipment Evaluation form, click Edit Shipment
   Evaluation in the dropdown next to a received shipment. Make changes as
   needed and then click Edit.
- To delete an existing Shipment Evaluation form, click **Delete Shipment Evaluation** in the dropdown next to a received shipment. In the bottom of the **Delete Shipment Evaluation** window, click **Delete**.

## **Reports**

The **Reports page** is where you can retrieve data that is stored in your laboratory's LDMS database.

While you can browse specimen information for individual participant visits on the **Specimen Management page**, you can't control what is displayed and how. For example, suppose you need a list of participant identifiers for all participants at your laboratory, and what project those participants belong to. You can't quickly get this information from the Specimen Management page.

This is where reports are helpful. The Reports page allows you to generate pre-defined reports about data in your laboratory's LDMS database. While the content of these reports are predetermined, you can often add criteria to filter what will be displayed. For example, you could generate a report that lets you see all participant identifiers at your laboratory, but only for a specific project.

## **Custom Report Builder**

The Custom Report Builder allows you to export specimen data from LDMS into a wide variety of formats.

The custom report builder allows you to do the following:

- specify what data to use to select specimens to appear on the report
- the information to appear on the report
- how to sort information on the report
- apply complex logic to reports
- save custom reports so they can be re-used

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Figure 47: The Custom Report Builder

#### Creating a custom report

Custom reports are created by indicating the information that should appear on the report, how the report should be sorted, and any special logic for selecting specimens.

Preview Generate Report

#### **Steps**

- 1. From the menu bar, click **Reports** > **Custom Report Builder**.
- 2. In the **All Fields** section, do the following:
  - For items that you want to *appear* on the report, select the check box under **Display**.
  - For items that you want to use to *sort* records on the report, select the check box under **Sort**.
- **3.** Optional: Use the **Add COUNT Field** button to provide aggregate counts on the report.
  - **3.1.** Click the **Add COUNT Field** button in the **All Fields** section. The **Add COUNT Field** window will open.
  - **3.2.** Select the field, enter the label, and mark as distinct as needed.
  - 3.3. Click **OK**.
- 4. In the **Fields to display** and **Sort results by** section, use the **Up** and **Down** buttons to set the order of items.

Fields displayed on the report will be shown from left-to-right in the order you specify. Results will be sorted in the order specified. An item can be removed by clicking the  $\mathbf{X}$  next to it or deselecting the check box.

#### **Example**

If you sort by **Destination Lab ID** and **Ship Date**, the records on the report will first be sorted by the destination laboratory, and *then* by the shipment date.

Optional: To remove duplicate records, under Fields to display, select Distinct.

Any rows on the report that have the same values across all displayed fields will only appear on the report once.

**6.** Optional: Apply a filter to your report in the **Selection criteria** section.

If no selection criteria is specified, all data from your laboratory will be included.

- **6.1.** In the **All Fields** section, click an item to select it.
- **6.2.** In the **Selection criteria** section, select an operator and value for your filter.
- 6.3. Click Add.
- **6.4.** In the box below the filters, enter logic for applying the filter using the numbers next to each item.

If you leave this box blank, filters will be applied as 1  $\,$  AND  $\,$  2  $\,$  AND  $\,$  . . . (etc.).

#### **Example**

For example, if you want to apply filter #1 and either filters #2 or #3, enter 1 AND (2 OR 3).

#### **Example**

To apply a filter that excludes specimens that originated from laboratory 500, select **Source Lab ID** in the **All Fields** section, the <> operator, and enter 500.

- Note: LDMS accepts wildcard values (\*). Wildcard values can be used for the Like and Not Like qualifiers.
- 7. In the **File Type** list, select the format for your report output.
- **8.** Optional: If you do not want a header row with the name of each column to appear on your report, deselect **Include Headers**.
- **9.** Do one of the follow:
  - To save your report so that it can be run later, enter a name for your report into the **Saved Queries** box at the top of the page, and then click **Save**.
  - To view the first 100 records of the report in your browser, click Preview.
  - To generate and save the report in the indicated file type, click **Generate Report**.

#### Running a saved report

Saved custom report can be run at any time.

#### **Steps**

- 1. From the menu bar, click **Reports** > **Custom Report Builder**.
- 2. In the **Saved Queries** box, select the report you want to run.
- 3. Optional: Make any changes needed to the report's fields, sort order, or selection criteria.
  - Changes will *not* be permanently saved unless you click **Save** at the top of the page.
- 4. Do one of the following:
  - To view the first 100 records of the report in your browser, click Preview.
  - To generate and save the report in the indicated file type, click **Generate Report.**

#### Modifying or deleting a saved report

A saved custom report can be modified and re-saved or deleted.

#### **Steps**

- From the menu bar, click **Reports** > **Custom Report Builder**. 1.
- 2. In the **Saved Queries** box, select the report you want to run.
- Do one of the following: 3.
  - To remove the saved report, click the **Delete** button.
  - To modify the report, make changes to its fields, sort order, or selection criteria, and then click **Save**.
  - To modify the report and save it as a new report, enter a new name into the **Saved Queries** box, and then click **Save**.

#### **Example custom report**

This is an example of a custom report for tracking specimens on shipments that have not been sent.

This report will find all specimens in your laboratory's database that are part of a shipment and are still available. It filters by specimens that are available and have a shipment setup date (all specimens with a shipment setup date will meet the "before 30/Sep/2099" criteria, since that date is in the future). This report would be useful for identifying specimens that you are expected to ship but still have in your possession.

- Fields to display Global specimen ID
  - Batch number
  - QA/QC status
  - Shipment setup date
  - Destination laboratory ID

#### Sort results by

- Destination laboratory ID
- Batch number

## Selection criteria

- Available = Yes
- Destination laboratory ID <> Your\_Laboratory\_ID
- Shipment setup date < 30/Sep/2099

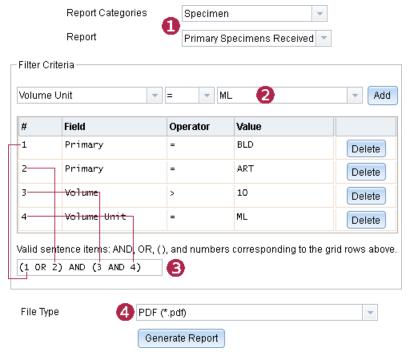
## Generating a report

All reports are generated by following this procedures

#### **Background**

By default, any report is generated using all available data in laboratory's database. If you want to narrow down the records that will appear on a report, you must apply filters. If more than one filter is applied, only records that met *all* of the filters will appear on the report.

Figure 48: The Reports page



In this example, the report that will be generated will contain all BLD or ART specimens that also have a volume greater than 10 mL. (1) Report categories, (2) Add criteria to table below, (3) Logic for generating report, (4) File format for the report

#### **Steps**

- 1. On the LDMS menu bar, click **Reports** > **Standard Reports**.
- 2. In the **Report categories** box, select a broad category for the report you want to generate.
- **3.** In the **Report** box select a report.

- **4.** Optional: Add one or more filter to narrow down the information displayed on the report.
  - **4.1.** Select a data item to filter by from the box below **Filter Criteria**.

Additional boxes will appear to specify the details for the filter.

- **4.2.** Select an operator from the middle box.
  - Note: The <> operator means "not equal to".
- **4.3.** Select a value for the filter from the third box.

The values available will be dependent on the filter. For example, if you are filtering by Additive, only valid additives in LDMS will be displayed.

- 4.4. Click the Add button.
  - Your criteria will be added to the list of filters. If you want to remove it after it has been added, click the **Delete** button to its right.
- **5.** Optional: Specify logic for the report in the box below the filter criteria.

The logic is formed by using the numbers in the # column in the filter criteria, along with the words AND, OR, or parenthesis.

- **6.** Select a format for the report from the **File Type** box.
- **7.** Click the **Generate** button.
  - Note: If no filter criteria is entered, LDMS will alert the user that the report may take a long time to generate.

#### Result

The report will be generated in the format that you specified. You may be asked to open or download the report, depending on your web browser's default behavior.

## Filter Reports Using a List of Specimens

You can use a list of identifiers to select a set of specimens.

The Specimen List feature is an available filter criteria on select reports. The filter criteria allows users to select specific specimens for display using a list of IDs. The file of identifiers must be a text file and must include one Global Specimen ID per line.

#### Figure 49: Global ID List File Example

| 0500-00ABCDE00-001 |
|--------------------|
| 0500-00ABCDE00-002 |
| 0500-00FGHIJ00-001 |
| 0500-00FGHIJ00-002 |
| 0500-00FGHIJ00-003 |
| 0500-00KLMNP00-001 |

## **Available report formats**

This section describes the file formats in which LDMS can generate reports.

- If you are generating a report from the **Exportable** reports category, the **Microsoft Excel Workbook Data-only** file type should be used.
- Reports in any category other than **Exportable** are designed for **PDF**.
- Use other file types for reports may result an unexpected formatting, depending on the report. There may also be variations in the file type's format, depending on the specific report.

Table 15: File types for LDMS reports

| File type                                  | Extension | Description   | Opened by  |
|--|-----------|---|--|
| Crystal Reports                            | .rpt      | Do not use  | N/A  |
| PDF  | .pdf      | A formatted report intended to be printed on letter size paper (preferred)  | A PDF viewer, such as Adobe Reader                       |
| Character separated values                 | .csv      | A text file with all information separated by comma characters  | A text editor, such as Notepad                           |
| Microsoft Excel<br>(97-2003)               | .xls      | A workbook for versions of Excel prior to Excel 2007; this version contains a header row for each column                    | Microsoft Excel or<br>LibreOffice Calc                   |
| Microsoft Excel<br>(97-2003) Data-<br>only | .xls      | Same as above, but usually without a header row   | Microsoft Excel or<br>LibreOffice Calc                   |
| Microsoft Excel<br>Workbook Data-<br>only  | .xlsx     | A workbook that will work in Excel 2007 or later (preferred for exportable reports)   | Microsoft Excel<br>2007 or later, or<br>LibreOffice Calc |
| Microsoft Word<br>(97-2003)                | .doc      | A Word document with formatting, such as tables and headers; can be difficult to modify, depending on the report            | Microsoft Word or<br>LibreOffice Writer                  |
| Microsoft Word<br>(97-2003) Editable       | .rtf      | A rich-text file that can be edited<br>by many word processors; uses less<br>formatting, which makes it easier to<br>modify | Microsoft Wordpad  |
| Rich Text Format                           | .rtf      | Same as Microsoft Word (97-2003) format, but compatible with RTF word processors  | Microsoft Wordpad  |
| XML  | .xml      | A Crystal Reports XML file. The scheme for this format <i>is available</i>  | An advanced text<br>editor, such as<br>Notepad++         |
| Tab-separated Text                         | .ttx      | A text file with information separated by tab characters.   | A text editor, such as Notepad                           |

## **Administrative reports**

#### **Anonymous Patients Map**

Provides a list of all anonymized specimens with their anonymized ID1s mapped to their source ID1s.

| Filter criteria   | Information displayed on report  |  |
|---|--|--|
| <ul> <li>Anonymized Date</li> <li>Anonymized ID1</li> <li>Original ID1</li> </ul> | <ul> <li>Source ID1</li> <li>Anon ID1</li> <li>Source Specimen ID</li> <li>Source Global Specimen ID</li> <li>Anon Global Spec ID</li> <li>Source Specimen Date</li> <li>Anon Specimen Date</li> </ul> |  |

## **Transaction Log Report**

On the Reports page, select **Report Categories > Admin**, then from the **Report** dropdown, select **Transaction Log Report**.

You may add start and end date filters as well as any others as desired.

| Filter criteria  | Information displayed on report   |  |
|--|---|--|
| <ul> <li>End Date</li> <li>Global Spec ID</li> <li>Start Date</li> <li>Transaction Type</li> <li>User</li> </ul> | <ul> <li>User</li> <li>Date/Time</li> <li>Table</li> <li>Transaction Type</li> <li>Old Data</li> <li>New Data</li> <li>LDMS Transaction ID</li> </ul> |  |

#### Figure 50: The Transaction Log Report

| User  | Date/Time                | Table        |        | Old Data  | New Data  | LDMS Transaction ID |
|-------|--------------------------|--------------|--------|---|---|---------------------|
| Moz   | 06/Nov/2023 12:36:42 EST | Patients     | Add    |   | ID1 = 123654; Group ID = 1300005                    | 32,549              |
| Moz   | 06/Nov/2023 12:36:42 EST | Enrollments  | Add    |   | ID2 = APRICOT                                       | 32,550              |
| Moz   | 06/Nov/2023 12:36:42 EST | Visits       | Add    |   | Visit Value = ; Visit Unit = ; ID3 = ; Clinic =     | 32,551              |
| Moz   | 06/Nov/2023 12:36:42 EST | VisitSamples | Add    |   | Global Specimen ID = KEQ0078L-00                    | 32,552              |
| Moz   | 06/Nov/2023 12:50:03 EST | Samples      | Update | Global Specimen ID = KEQ0078L-01; Specimen ID = 500V15000003; OriginalVolume    | OriginalVolume = 4                                  | 32,553              |
| Moz   | 06/Nov/2023 12:54:31 EST | VisitSamples | Delete | Global Specimen ID = KEQ0078L-06  |   | 32,554              |
| Moz   | 06/Nov/2023 12:54:31 EST | Samples      | Delete | Global Specimen ID = KEQ0078L-06; Specimen ID = 500V15000003                    |   | 32,555              |
| Moz   | 06/Nov/2023 12:54:41 EST | Samples      | Update | Global Specimen ID = KEQ0078L-04; Specimen ID = 500V15000003; OriginalVolume    | : OriginalVolume = 13                               | 32,556              |
| Moz   | 06/Nov/2023 12:55:12 EST | Samples      | Add    |   | Global Specimen ID = -NEW-GU-; Specimen ID =        | 32,558              |
| Moz   | 06/Nov/2023 12:55:12 EST | VisitSamples | Add    |   | Global Specimen ID = 0500-001MCA00-007              | 32,559              |
| Moz   | 06/Nov/2023 12:55:12 EST | Samples      | Update | Global Specimen ID = KEQ0078L-00; Specimen ID = 500V15000001; SubSamplesCre     | : SubSamplesCreated = 7                             | 32,560              |
| Moz   | 06/Nov/2023 12:55:12 EST | Samples      | Update | Global Specimen ID = -NEW-GU-; Specimen ID = ; Global Specimen Id = -NEW-GU-; A | GlobalSpecimenId = 0500-001MCA00-007; AliquotSeque  | 32,561              |
| Moz   | 06/Nov/2023 12:58:54 EST | VisitSamples | Delete | Global Specimen ID = 0500-001MCA00-007  |   | 32,562              |
| Moz   | 06/Nov/2023 12:58:54 EST | Samples      | Delete | Global Specimen ID = 0500-001MCA00-007; Specimen ID =                           |   | 32,563              |
| Moz   | 06/Nov/2023 12:59:28 EST | Samples      | Update | Global Specimen ID = KEQ0078L-04; Specimen ID = 500V15000003; OriginalVolume    | OriginalVolume = 11                                 | 32,564              |
| Moz   | 06/Nov/2023 12:59:38 EST | Samples      | Update | Global Specimen ID = KEQ0078V-00; Specimen ID = 500V15000001; OriginalVolume    | OriginalVolume = 45                                 | 32,565              |
| Moz   | 06/Nov/2023 13:01:34 EST | Samples      | Add    |   | Global Specimen ID = -NEW-GU-; Specimen ID =        | 32,566              |
| Moz   | 06/Nov/2023 13:01:34 EST | VisitSamples | Add    |   | Global Specimen ID = 0500-00257C00-000              | 32,567              |
| Moz   | 06/Nov/2023 13:01:34 EST | Samples      | Update | Global Specimen ID = -NEW-GU-; Specimen ID = ; RootSampleId = ; GlobalSpecimen  | RootSampleId = 10830; GlobalSpecimenId = 0500-0025  | 32,568              |
| admin | 06/Nov/2023 15:15:59 EST | Samples      | Update | Global Specimen ID = 0777-001MWK00-276; Specimen ID = ; Condition = 10412; Co   | Condition = 10424; Comments = Received_TESTING_AB   | 32,570              |
| admin | 06/Nov/2023 15:15:59 EST | Samples      | Update | Global Specimen ID = 0777-001MWK00-277; Specimen ID = ; Condition = 10412; Co   | Condition = 10424; Comments = Received_TESTING_AB   | 32,571              |
| admin | 06/Nov/2023 15:15:59 EST | Samples      | Update | Global Specimen ID = 0777-001MWK00-278; Specimen ID = ; Condition = 10412; Co   | Condition = 10424; Comments = Received_TESTING_AB   | 32,572              |
| admin | 06/Nov/2023 15:15:59 EST | Samples      | Update | Global Specimen ID = 0777-001MWK00-281; Specimen ID = ; Condition = 10412; Co   | Condition = 10424; Comments = Received_TESTING_AB   | 32,573              |
| admin | 06/Nov/2023 15:15:59 EST | Samples      | Update | Global Specimen ID = 0777-001MWK00-282; Specimen ID = ; Condition = 10412; Co   | Condition = 10424; Comments = Received_TESTING_AB   | 32,574              |
| admin | 06/Nov/2023 15:19:45 EST | Samples      | Update | Global Specimen ID = 0500-001NVC00-001; Specimen ID = ; Condition = 10412; Con  | Condition = 10413; Comments = Sent_TESTING_AJB; Int | 32,575              |
| admin | 06/Nov/2023 15:19:45 EST | Samples      | Update | Global Specimen ID = 0500-001NVC00-003; Specimen ID = ; Condition = 10412; Cor  | Condition = 10413; Comments = Sent_TESTING_AJB; Int | 32,576              |
|       |                          |              |        |   |   |                     |

#### **User Event Report**

Provides a report of user events, such as adding, updating, or deleting information in LDMS.

| Filter criteria  | Information displayed on report  |  |
|--|--|--|
| <ul> <li>Event Date</li> <li>Event Type</li> <li>Login ID</li> <li>Module</li> <li>Transaction ID</li> </ul> | <ul> <li>Login ID</li> <li>Event Date/Time</li> <li>Module</li> <li>Event Type</li> <li>Description</li> <li>Reason</li> <li>Item Name</li> <li>Item ID</li> <li>Min Transaction ID</li> <li>Max Transaction ID</li> </ul> |  |

#### **User permissions report**

Shows each user account associated with your LDMS laboratory, and the user's access to specific pages in LDMS.

#### **Example**

Figure 51: The User Permission Report

#### **User Permissions Report**

| jdoe           | Full | View |
|----------------|------|------|
| Specimen       | х    |      |
| DataRetrieval  | Х    |      |
| Reports        | Х    |      |
| Administration | Х    |      |
| Account        |      |      |
| Labels         | Х    |      |
| Shipping       | Х    |      |
| Storage        | Х    |      |

#### System Administrator

## **Barcode Reports**

#### **1D Barcodes Report**

Displays a line listing of specimens along with a series of 1D barcodes. Each specimen row will include a 1D barcode that encodes the specimens Global Spec ID, ID1, and Other Spec ID. This reports can be printed and used to scan groups of specimens into applications outside of LDMS that may use 1D barcodes to input information such as Global Spec ID, ID1, or Other Spec ID.

<sup>(1)</sup> The user's name. (2) The user's permissions by page. (3) Only present if the user has system administrator privileges.

| Filter criteria   | Information displayed on report  |
|---|--|
| <ul> <li>ACTG PID</li> <li>ACTG Protocol</li> <li>Additive</li> <li>Clinic ID</li> <li>Derivative</li> <li>Global Spec ID</li> <li>Non ACTG PID/ID1</li> <li>Non ACTG PID/ID2</li> <li>Primary</li> <li>Project</li> <li>Received Date</li> <li>Run ID</li> <li>Shipment Number</li> <li>SID/ID3</li> <li>Specimen Date</li> <li>Visit Unit</li> <li>Visit Value</li> </ul> | <ul> <li>Global Spec ID</li> <li>Project</li> <li>ID2</li> <li>Id1</li> <li>Collection Date</li> <li>Vid</li> <li>Vid Unit</li> <li>Other Spec ID</li> <li>Global Spec ID Barcode</li> <li>ID1 Barcode</li> <li>Other Spec ID Barcode</li> </ul> |

#### **2D Barcodes Report**

Displays a line listing of specimens along with a series of 2D barcodes. Each specimen row will include a 2D barcode that encodes the specimens Global Spec ID, ID1, and Other Spec ID. This reports can be printed and used to scan groups of specimens into applications outside of LDMS that may use 2D barcodes to input information such as Global Spec ID, ID1, or Other Spec ID.

| Filter criteria   | Information displayed on report  |
|---|--|
| ACTG PID     ACTG Protocol     Additive     Clinic ID     Derivative     Global Spec ID     Non ACTG PID/ID1     Non ACTG PID/ID2     Primary     Project     Received Date     Run ID     Shipment Number     SID/ID3     Specimen Date     Visit Unit     Visit Value | <ul> <li>Global Spec ID</li> <li>Project</li> <li>ID2</li> <li>Id1</li> <li>Collection Date</li> <li>Vid</li> <li>Vid Unit</li> <li>Other Spec ID</li> <li>Global Spec ID Barcode</li> <li>ID1 Barcode</li> <li>Other Spec ID Barcode</li> </ul> |

## **Exportable reports**

Exportable reports are designed to be generated for spreadsheet and CSV formats so that the data can be manipulated manually.

## **Abbott SARS-COV-2 Quant Export Report**

Displays specimen details and results from Abbott SARS-COV-2 Quant runs in the Test Results module.

| Sorting | Available filters   | Information on report  |
|---------|---|--|
| _       | <ul> <li>ACTG PID</li> <li>ACTG Protocol</li> <li>ACTG Protocol Type</li> <li>Additive</li> <li>Assay Date</li> <li>Clinic ID</li> <li>Derivative</li> <li>Global Spec ID</li> <li>Non ACTG PID/ID1</li> <li>Non ACTG Prot/ID2</li> <li>Other Specimen ID</li> <li>Primary</li> <li>Project</li> <li>Run ID</li> <li>SID/ID3</li> <li>Specimen Date</li> <li>Specimen ID</li> </ul> | <ul> <li>Group</li> <li>Protocol</li> <li>PID</li> <li>Secondary ID</li> <li>Qualitative Result</li> <li>Quantitative Result (Log copies/mL</li> <li>Dilution Factor</li> <li>Final Quantitative Result (Log copies/mL</li> <li>m2000 Error Code/ description</li> <li>Primary</li> <li>Additive</li> <li>Derivative</li> <li>Sub Add/Der</li> <li>Visit</li> <li>Clinic</li> <li>Collection Date/Time</li> <li>Other Specimen ID</li> <li>Global Specimen ID</li> <li>Run ID</li> <li>Assay Date</li> </ul> |

## **Abbott SARS-COV-2 Quant Export Report with Comments**

Displays specimen details and results from Abbott SARS-COV-2 Quant runs in the Test Results module. Report includes Specimen comments for each result.

| Sorting | Available filters   | Information on report  |
|---------|---|--|
|         | ACTG PID     ACTG Protocol     ACTG Protocol Type     Additive     Assay Date     Clinic ID     Derivative     Global Spec ID     Non ACTG PID/ID1     Non ACTG Prot/ID2     Other Specimen ID     Primary     Project     Run ID     SID/ID3     Specimen Date     Specimen ID | Group Protocol PID Secondary ID Qualitative Result Quantitative Result (Log copies/mL Dilution Factor Final Quantitative Result (Log copies/mL m2000 Error Code/description Primary Additive Derivative Sub Add/Der Visit Clinic |

| Sorting | Available filters | Information on report         |
|---------|-------------------|-------------------------------|
|         |                   | Collection Date/Time          |
|         |                   | Other Specimen ID             |
|         |                   | Global Specimen ID            |
|         |                   | Run ID                        |
|         |                   | Assay Date                    |
|         |                   | <ul> <li>Condition</li> </ul> |
|         |                   | Comments                      |
|         |                   |                               |

# **Abbott SARS-COV-2 Quant Export Report with Sample Location**

Displays specimen details and results from Abbott SARS-COV-2 Quant runs in the Test Results module. Report includes specimen location field from the Abbott result file.

| Sorting | Available filters   | Information on report   |
|---------|---|---|
| -       | ACTG PID     ACTG Protocol     ACTG Protocol Type     Additive     Assay Date     Clinic ID     Derivative     Global Spec ID     Non ACTG PID/ID1     Non ACTG Prot/ID2     Other Specimen ID     Primary     Project     Run ID     SID/ID3     Specimen ID     Specimen ID | Group Protocol PID Secondary ID Qualitative Result Quantitative Result (Log copies/mL Dilution Factor Final Quantitative Result (Log copies/mL m2000 Error Code/description Primary Additive Derivative Sub Add/Der Visit Clinic Collection Date/Time Other Specimen ID Run ID Assay Date Sample Location |

## Aliquot count by primary report

Shows the number of aliquots processed from each primary specimen.

| Sorting                              | Filter criteria  | Information displayed on report   |
|--------------------------------------|--|---|
| This report is sorted by ID1 and ID2 | <ul> <li>Additive</li> <li>Global spec ID</li> <li>Non ACTG PID/ID1</li> <li>Non ACTG Prot/ID2</li> <li>Other specimen ID</li> </ul> | <ul> <li>Additive</li> <li>Global specimen ID</li> <li>ID1</li> <li>ID2</li> <li>Other specimen ID</li> </ul> |

| Sorting | Filter criteria  | Information displayed on report   |
|---------|--|---|
|         | <ul><li>Primary</li><li>project</li><li>Specimen date</li><li>Visit unit</li><li>Visit value</li></ul> | <ul> <li>Primary</li> <li>project</li> <li>Specimen date</li> <li>Visit value and unit</li> <li>Primary volume</li> <li>Aliquot volume</li> <li>Number of aliquots</li> </ul> |

## **Aliquot Inventory Report**

Provides a specimen listing with a custom set of header values as shown below.

| Sorting | Available filters   | Information on report   |
|---------|---|---|
|         | ACTG PID ACTG Protocol Additive Clinic ID Derivative Global Spec ID Import Date Non ACTG PID/ID1 Non ACTG Prot/ID2 Other Specimen ID Primary Project Received Date Shipment Number Specimen Date Sub Add/Der Visit Unit Visit Value | PID Collection Date Collection Date Collection Date Collection Date Collection Date Collection time (min) Visit Value Visit Unit Study name Global Specimen ID Primary Type Additive Type Derivative Type Sub add/der type Volume Volume Volume Volume Frimary Volume Vol Unit Processing Time Processing Month Processing Day Processing Year Frz date Frz time Cell Count Condition Is sample Available Is sample stored Stored month Stored Day Stored Year Storage temp Is sample Comments Storage Freezer Storage Rack Storage Container |

| Sorting | Available filters | Information on report |
|---------|-------------------|-----------------------|
|         |                   | Storage Box position  |

## **CFAR** export report

Shows specimens stored at the laboratory.

| Sorting                              | Filter criteria | Information displayed on report  |
|--------------------------------------|-----------------|--|
| This report is sorted by specimen ID | • project       | <ul> <li>Specimen ID</li> <li>project</li> <li>Global specimen ID</li> <li>ID2</li> <li>OPID</li> <li>Visit value and unit</li> <li>Primary</li> <li>Additive</li> <li>Derivative</li> <li>Sub additive/derivative</li> <li>Volume</li> <li>Volume unit</li> <li>Stored date</li> <li>Specimen date</li> <li>Storage location</li> </ul> |

## **CFAR storage report**

Provides a summary of storage items at the laboratory.

| Sorting  | Filter criteria  | Information displayed on report   |
|--|--|---|
| This report is sorted by storage container and specimen ID | <ul> <li>Import date</li> <li>project</li> <li>Received date</li> <li>Specimen date</li> </ul> | <ul> <li>Container</li> <li>Specimen ID</li> <li>project</li> <li>ID2</li> <li>ID1</li> <li>Global specimen ID</li> <li>Visit</li> <li>Primary</li> <li>Additive</li> <li>Derivative</li> <li>Sub-additive/derivative</li> <li>Volume</li> <li>Specimen date</li> <li>Position</li> </ul> |

## **Database dump report**

A general report that provides information from all specimens in a laboratory's LDMS database.

| Sorting | Available filters   | Information on report  |
|---------|---|--|
|         | ACTG PID ACTG Protocol ACTG Protocol Type Additive Clinic ID Derivative Global Spec ID Non ACTG PID/ID1 Non ACTG Prot/ID2 Other Specimen ID Primary project Received Date Ship to Lab Number Specimen Date Specimen ID Sub Add/Der Visit Unit Visit Value Volume Volume Volume Unit | <ul> <li>project</li> <li>PID/ID1</li> <li>Protocol/ID2</li> <li>SID/ID3</li> <li>Visit</li> <li>Visit Unit</li> <li>Clinic</li> <li>Rec Date</li> <li>Rec Time</li> <li>Collection Date</li> <li>Collection Time</li> <li>Import Date</li> <li>Spec ID</li> <li>Global Spec ID</li> <li>PRI</li> <li>ADD</li> <li>DER</li> <li>SUB</li> <li>VOL</li> <li>VOL Unit</li> <li>Cond</li> <li>Time</li> <li>Time Unit</li> <li>Other Spec ID</li> <li>Test</li> <li>Comments</li> <li>Ship Date</li> <li>Ship Batch Number</li> <li>Ship to Lab</li> </ul> |

## **Database dump report 2**

A general report that provides information from all specimens in a laboratory's LDMS database.

| Sorting | Available filters  | Information on report  |
|---------|--|--|
| _       | ACTG PID     ACTG Protocol     Additive     Clinic ID     Derivative     Global Spec ID     Import Date     Non ACTG PID/ID1     Non ACTG Prot/ID2     Other Specimen ID     Primary     project     Received Date     Shipment Number     Specimen Date     Sub Add/Der | LDMS laboratory ID Primary unique ID Aliquot unique ID project ID1 Collection date (month, day, and year) Received date (month, day, and year) Collection time Visit (value and unit) ID2 ID3 Global specimen ID Specimen ID Other specimen ID |

| Sorting | Available filters | Information on report   |
|---------|-------------------|---|
|         | Visit Unit        | Primary code  |
|         | Visit Value       | Additive code   |
|         |                   | Derivative code   |
|         |                   | Sub-additive/derivative code                                      |
|         |                   | Enrollment  |
|         |                   | Aliquot volume (value and unit)                                   |
|         |                   | Primary volume (value and unit)                                   |
|         |                   | Processing time   |
|         |                   | Processing date (month,   |
|         |                   | day, and year)  |
|         |                   | Processing tech initials  |
|         |                   | Frozen date (month, day, year)                                    |
|         |                   | Frozen time   |
|         |                   | Total cell count  |
|         |                   | Specimen condition  |
|         |                   | Availability  |
|         |                   | Additional time (value and unit)                                  |
|         |                   | Status (not stored and<br>available, stored, and<br>unavailable)  |
|         |                   | Storage date (month, day, year)                                   |
|         |                   | Storage temperature   |
|         |                   | Shipping status   |
|         |                   | Destination laboratory  |
|         |                   | Shipment number   |
|         |                   | Shipment temperature  |
|         |                   | • Shipment date (month, day, year)                                |
|         |                   | <ul> <li>Sending laboratory</li> </ul>                            |
|         |                   | Received shipment number  |
|         |                   | Received temperature  |
|         |                   | <ul> <li>Date shipment received<br/>(month, day, year)</li> </ul> |
|         |                   | Clinic  |
|         |                   | Comments  |
|         |                   | <ul> <li>Whether a quick add<br/>template was used</li> </ul>     |
|         |                   | Reason specimen not collected                                     |
|         |                   | Storage unit  |
|         |                   | Storage level   |
|         |                   | Storage sub-level   |
|         |                   | Storage container   |
|         |                   | Position  |
|         |                   | System date   |
|         |                   | Date last changed   |
|         |                   | - Date last changed   |

## **Exportable Abbott Assay Report**

| Sorting | Available filters   | Information on report  |
|---------|---|--|
|         | ACTG PID/ID1     ACTG Protocol     ACTG Protocol Type     Assay Date     Global Spec ID     Non ACTG PID/ID1     Non ACTG Prot/ID2     Primary     Project     Run ID     Specimen Date     Specimen ID | <ul> <li>PID</li> <li>ID2/Pro</li> <li>ID3</li> <li>Visit</li> <li>PRI</li> <li>Specimen ID</li> <li>Global Spec ID</li> <li>Spec Date</li> <li>Rec Date</li> <li>Import Date</li> <li>Assay Date</li> <li>RUN ID</li> <li>Dilution</li> <li>Result</li> <li>System Censor</li> <li>User Censor</li> <li>Run System Censor</li> <li>Run User Censor</li> </ul> |

## Laboratory 081 billing report

Report request by laboratory 81 (but can be used for other purposes).

| Sorting   | Filter criteria   | Information displayed on report  |
|---|---|--|
| This report is sorted by project, ID2, and ID1. | <ul> <li>ACTG PID</li> <li>ACTG protocol</li> <li>ACTG protocol type</li> <li>Derivative</li> <li>Global specimen ID</li> <li>project</li> <li>Specimen date</li> </ul> | <ul> <li>project</li> <li>ACTG protocol</li> <li>PID/ID1</li> <li>Specimen date</li> <li>Derivative</li> <li>Global specimen ID</li> </ul> |

## Laboratory 81 CNICS by date report

Report requested by laboratory 81 (but can be used by any laboratory).

| Sorting                                | Filter criteria  | Information displayed on report   |
|--|--|---|
| This report is sorted by specimen date | <ul> <li>Non ACTG prot/ID2</li> <li>Primary</li> <li>project</li> <li>Received date</li> <li>Specimen date</li> <li>Volume</li> <li>Volume unit</li> </ul> | <ul> <li>project</li> <li>Non ACTG protocol/ID2</li> <li>Specimen date</li> <li>Received date</li> <li>Primary</li> <li>Volume</li> <li>Volume units</li> </ul> |

## Laboratory 081 CNICS general report

Report requested by laboratory 81 (but can be used by any laboratory).

| Sorting                      | Filter criteria  | Information displayed on report  |
|------------------------------|--|--|
| This report is sorted by ID1 | <ul> <li>Additive</li> <li>Derivative</li> <li>Non ACTG PID/ID1</li> <li>Non ACTG Prot/ID2</li> <li>Received date</li> <li>Specimen date</li> <li>Specimen time</li> <li>Sub additive/derivative</li> <li>Volume</li> <li>Volume unit</li> </ul> | <ul> <li>Non ACTG protocol/ID2</li> <li>PID/ID1</li> <li>Specimen date</li> <li>Specimen time</li> <li>Received date</li> <li>Additive</li> <li>Derivative</li> <li>Sub additive/derivative</li> <li>Volume</li> <li>Volume units</li> </ul> |

## **Laboratory 188 storage report**

Provides a summary a summary of stored and not stored specimens, useful for determining what specimens still needs to be assigned a storage location in LDMS.

| Sorting                                     | Filter criteria   | Information displayed on report  |
|---|---|--|
| This report is sorted by global specimen ID | ACTG PID/ID1 ACTG Protocol ACTG Protocol Type Additive Condition Derivative Global Spec ID Non ACTG PID/ID1 Non ACTG PID/ID1 Other Specimen ID Primary project SID/ID3 Specimen Date Specimen ID Specimen Time Time Time Time Unit Visit Unit Volume Volume Volume Unit | <ul> <li>Global specimen ID</li> <li>Other specimen ID</li> <li>Freezer</li> <li>Shelf</li> <li>Rack</li> <li>Container</li> <li>Position</li> </ul> |

## **Lab 40 LabKey report**

Short reference description.

| Sorting                              | Filter criteria                | Information displayed on report  |
|--------------------------------------|--------------------------------|----------------------------------|
| This report is sorted by specimen ID | ACTG PID/ID1     ACTG Protocol | GlobalSpecID     Laboratory Name |

| Sorting | Filter criteria   | Information displayed on report  |
|---------|---|--|
|         | ACTG Protocol Type     Additive     Clinic ID     Derivative     Global Spec ID     Import Date     Non ACTG PID/ID1     Non ACTG Prot/ID2     Primary     project     Received Date     Specimen Date     Specimen ID     Sub Add/Der     Visit Unit     Visit Value | Clinic PTID Specimen Date Receipt Date LDMS Spec ID project Visit Protocol Visit Unit Volume Volume Unit Storage Date Ship Date Ship Lab Sub additive/derivative Comments Primary Derivative Additive Condition Storage unit Level 1 Level 2 Container Position Frozen Time Processing Tech Initials Processing Date Total Cell Count Other Spec ID ID3 OPID Processing time Frozen date Primary Volume Primary Volume Primary Vol Units Received Time Mark for Shipping |

## **Laboratory 48 billing report**

Report that provides specimen information.

| Filter criteria  | Information displayed on report   |
|--|---|
| <ul> <li>ACTG PID</li> <li>ACTG Protocol</li> <li>ACTG Protocol Type</li> <li>Additive</li> <li>Clinic ID</li> <li>Non ACTG PID/ID1</li> <li>Non ACTG Prot/ID2</li> <li>project</li> </ul> | <ul> <li>Specimen ID</li> <li>project/Protocol</li> <li>PID</li> <li>Visit/Unit</li> <li>Specimen Date</li> <li>Additive</li> </ul> |

| Filter criteria | Information displayed on report |
|-----------------|---------------------------------|
| Specimen Date   |                                 |
| Specimen ID     |                                 |
| Visit Unit      |                                 |
| Visit Value     |                                 |
|                 |                                 |

## Lab 485 aliquot report

Similar to the Specimen Export Report, but sorts the results differently.

| Sorting  | Filter criteria   | Information displayed on report  |
|--|---|--|
| <ol> <li>project</li> <li>Non-ACTG Protocol</li> <li>Non-ACTG PID</li> <li>Visit Value</li> <li>Primary</li> <li>Additive</li> <li>Derivative</li> </ol> | ACTG PID/ID1 ACTG Protocol ACTG Protocol Type Additive Derivative Non ACTG PID/ID1 Non ACTG Prot/ID2 Primary project Sub Add/Der Visit Unit Visit Value | <ul> <li>project</li> <li>PID/ID1</li> <li>Protocol</li> <li>SID/ID3</li> <li>Visit</li> <li>Visit [Unit]</li> <li>Clinic</li> <li>Received Date</li> <li>Collection Date</li> <li>Collection Time</li> <li>Import Date</li> <li>Specimen ID</li> <li>Global Specimen ID</li> <li>Primary</li> <li>Additive</li> <li>Derivative</li> <li>Sub Additive/Derivative</li> <li>Volume</li> <li>Volume Unit</li> <li>Condition</li> <li>Time</li> <li>Time Unit</li> <li>Other Specimen ID</li> <li>Test</li> <li>Comments</li> <li>Ship Date</li> <li>Ship Batch</li> <li>Ship to Laboratory</li> </ul> |

## Lab 485 specimen count report

Provides the number of specimens with a unique primary-additive-derivative-sub  $\mbox{A/D}$  combination.

| Sorting   | Filter criteria   | Information displayed on report   |
|---|---|---|
| <ol> <li>project and study combination</li> <li>Participant</li> <li>Visit</li> </ol> | <ul> <li>ACTG PID/ID1</li> <li>ACTG Protocol</li> <li>ACTG Protocol Type</li> <li>Additive</li> <li>Derivative</li> </ul> | <ul><li>PID</li><li>Visit</li><li>Visit Unit</li><li>Primary</li><li>Additive</li></ul> |

| Sorting | Filter criteria  | Information displayed on report  |
|---------|--|--|
|         | <ul> <li>Non ACTG PID/ID1</li> <li>Non ACTG Prot/ID2</li> <li>Primary</li> <li>project</li> <li>Sub Add/Der</li> <li>Visit Unit</li> <li>Visit Value</li> <li>Volume</li> <li>Volume unit</li> </ul> | <ul><li>Derivative</li><li>Sub Additive/Derivative</li><li>Count</li></ul> |

## Lab 485 specimen count with volume report

Provides the number of specimens with a unique primary-additive-derivative-sub A/D combination, along with total volume.

| Sorting   | Filter criteria  | Information displayed on report  |
|---|--|--|
| <ol> <li>project and study combination</li> <li>Participant</li> <li>Visit</li> </ol> | <ul> <li>ACTG PID/ID1</li> <li>ACTG Protocol</li> <li>ACTG Protocol Type</li> <li>Additive</li> <li>Derivative</li> <li>Non ACTG PID/ID1</li> <li>Non ACTG Prot/ID2</li> <li>Primary</li> <li>project</li> <li>Sub Add/Der</li> <li>Visit Unit</li> <li>Visit Value</li> </ul> | <ul> <li>PID</li> <li>Visit</li> <li>Visit Unit</li> <li>Primary</li> <li>Additive</li> <li>Derivative</li> <li>Sub Additive/Derivative</li> <li>Volume</li> <li>Volume Unit</li> <li>Count</li> </ul> |

#### Primary specimen database dump report

Provides a list of specimen ID numbers for primaries, along with draw dates and received dates.

| Sorting                              | Filter criteria | Information displayed on report   |
|--------------------------------------|-----------------|---|
| This report is sorted by specimen ID | None            | <ul><li>Specimen ID</li><li>Other specimen ID</li><li>Collection date</li><li>Received date</li></ul> |

## Sample counts for specified project report

Shows how many specimens for a given derivative type are available for each project locally.

| Sorting                             | Filter criteria        | Information displayed on report |
|-------------------------------------|------------------------|---------------------------------|
| This report is organized by project | Derivative     project | Derivative type                 |

| Sorting | Filter criteria                 | Information displayed on report                                |
|---------|---------------------------------|--|
|         | Received date     Specimen date | Description of derivative type     Count (number of specimens) |

## **Specimen export report**

Designed for pulling information for local groups that have been shipped.

| Sorting   | Filter criteria      | Information displayed on report   |
|---|----------------------|---|
| This report is sorted by project, then by participant identifier (ID1), and then by specimen ID | Shipped batch number | <ul> <li>Protocol</li> <li>project</li> <li>Primary type</li> <li>Derivative type</li> <li>Additive</li> <li>Sub additive/derivative type</li> <li>Collection date</li> <li>Visit value</li> <li>Visit unit</li> <li>Global specimen ID</li> <li>Original volume</li> <li>Original volume</li> <li>Available volume</li> <li>Available volume</li> <li>Elinic</li> <li>Time</li> <li>Received date</li> <li>Participant identifier (ID1)</li> <li>Protocol identifier (ID2)</li> <li>Source laboratory</li> <li>Draw time</li> <li>Received time</li> </ul> |

## Storage export report

Provides a summary of specimens and their storage location in a laboratory's local database.

| Sorting | Filter criteria  | Information displayed on report   |
|---------|--|---|
| _       | <ul> <li>ACTG PID/ID1</li> <li>ACTG protocol</li> <li>ACTG protocol type</li> <li>Additive</li> <li>Derivative</li> <li>Non ACTG PID/ID1</li> <li>Non ACTG Prot/ID2</li> <li>OPID</li> <li>Other specimen ID</li> <li>Primary</li> </ul> | <ul> <li>Specimen ID</li> <li>project/Prot</li> <li>PID/ID1</li> <li>OPID</li> <li>Primary</li> <li>Additive</li> <li>Derivative</li> <li>Sub additive/derivative</li> <li>Specimen Date</li> <li>Visit ID</li> </ul> |

| Sorting | Filter criteria  | Information displayed on report                                       |
|---------|--|---|
|         | <ul> <li>project</li> <li>Specimen date</li> <li>Specimen ID</li> <li>Sub add/der</li> <li>Visit unit</li> <li>Visit value</li> <li>Volume</li> <li>Volume unit</li> </ul> | Volume Time Shipped no. Other specimen ID Storage location + position |

## **Westat PK Export Report**

Used to transfer PK results to Westat data management team.

| Filter criteria  | Information displayed on report   |
|--|---|
| Assay Date     Non ACTG PID/ID1     Non ACTG Prot/ID2     Project     Run ID     Specimen ID | <ul> <li>Specimen ID</li> <li>PID/ID1</li> <li>Other Spec ID</li> <li>VID</li> <li>Spec Date</li> <li>Spec Time</li> <li>Time/Unit</li> <li>Assay Date</li> <li>Run ID</li> <li>Add</li> <li>Der</li> <li>Drug</li> <li>Conc</li> <li>Units</li> <li>Censors</li> <li>Reviewed</li> </ul> |

## **Miscellaneous reports**

## **Clinic contact report**

Provides contact information for clinics by clinic ID number.

| Sorting                            | Filter criteria | Information displayed on report   |
|------------------------------------|-----------------|---|
| This report is sorted by clinic ID | Clinic ID       | <ul> <li>Clinic ID</li> <li>Clinic name</li> <li>Address</li> <li>Telephone number</li> <li>Fax number</li> <li>Contact(s) [including name, phone number, fax number, and email address]</li> </ul> |

#### LDMS abbreviated codes report

Provides a list of LDMS unit, visit, time, specimen condition codes used in LDMS and their meaning.

#### Information displayed on report



**Note:** The information on this report is identical to the hover text that appears throughout LDMS.

- Measurement codes
- Visit unit codes
- Time unit codes
- · Specimen condition codes

#### LDMS assay censor codes report

Provides a list of assay censor codes.

#### Information displayed on report

- · Censor codes
- · Code descriptions

## LDMS primary, additive, derivative, sub additive/ derivative codes report

Provides a list of LDMS primary, additive, derivative, and sub additive/derivative codes throughout LDMS.

#### Information displayed on report



**Note:** The information on this report is identical to the hover text that appears throughout LDMS.

- · Primary codes
- · Additive codes
- · Derivative codes
- · Sub additive/derivative codes

#### **MWCCS**

## **MWCCS Processing Report**

| Filter criteria  | Information displayed on report  |
|--|--|
| Non-ACTG PID/ID1     Project     Specimen Date     Specimen ID | <ul><li>Global Spec ID</li><li>Der</li><li>Other Spec ID</li><li>Volume</li></ul>  |
|  | <ul><li>Number of Aliquots</li><li>Study</li><li>Processing Instructions</li></ul> |

| Filter criteria | Information displayed on report  |
|-----------------|--|
|                 | <ul> <li>Cell count</li> <li>Amt</li> <li>Storage</li> <li>Processor's Initials</li> <li>Date/Time Processed/Frozen</li> <li>Unused Tubes</li> </ul> |

## **Participant reports**

#### **Participant identifiers report**

Provides a list of participants in your database by project.

| Sorting   | Filter criteria   | Information displayed on report   |
|---|---|---|
| This report is sorted by project, and then by ID1 | ACTG PID     ACTG protocol     ACTG protocol type     Non ACTG PID/ID1     Non ACTG Prot/ID2     OPID     project     SID/ID3 | <ul><li>project</li><li>ID1</li><li>ID2</li><li>ID3</li><li>OPID</li><li>Clinic</li></ul> |

## **PK reports**

## **Pharmacology Drug Count**

Displays a result count by analyte per study/ protocol.

#### **Filter Criteria**

- ACTG Protocol
- ACTG Protocol Type
- Assay Date
- Clinic ID
- Project
- Specimen Date

## **Pharmacology Drug List**

List of pharmacology drugs and their associated codes.

## **Pharmacology Proficiency Results**

Displays results of Pharmacology proficiency testing

#### **Filter Criteria**

• Round Number

#### **PK Drug Limits By Run**

PK drug limits sorted by run.

| Filter criteria  | Information displayed on report   |
|--|---|
| <ul><li>Assay Date</li><li>Drug</li><li>Run ID</li></ul> | <ul> <li>Run ID</li> <li>Drug</li> <li>Drug Lower Limit</li> <li>Drug Upper Limit</li> <li>Run Lower Limit</li> <li>Run Upper Limit</li> <li>Units</li> </ul> |

## **PK Participant Report**

An individualized report per specimen showing associated participant details along with detailed test results including censor codes, run limits, and a final calculated result.

| Filter criteria   | Information displayed on report   |
|---|---|
| ACTG PID/ID1 ACTG Protocol Assay Date Clinic ID Derivative Global Spec ID Non ACTG PID/ID1 Non ACTG Prot/ID2 Other Specimen ID Primary Project Received Date Run ID SID/ID3 Specimen Date Specimen ID | <ul> <li>Participant</li> <li>Project / ID2</li> <li>ID3</li> <li>Collection Date</li> <li>Visit</li> <li>Clinic Name</li> <li>Fax</li> <li>Testing Lab</li> <li>Specimen ID</li> <li>Global Spec ID</li> <li>Other Spec ID</li> <li>Received Date</li> <li>Received Time</li> <li>Sample Condition</li> <li>Primary</li> <li>Additive</li> <li>Derivative</li> <li>Run ID</li> <li>Detection Platform</li> <li>Run Type</li> <li>Test Date</li> <li>Tech Initials</li> <li>Analyte</li> <li>Description</li> <li>Unit</li> <li>Concentration</li> <li>Lower Limit</li> <li>Upper Limit</li> <li>Censors</li> <li>Review Comment</li> </ul> |

| Filter criteria | Information displayed on report |  |
|-----------------|---------------------------------|--|
|                 | Sample Comment                  |  |

## **PK Summary Report**

PK Summary Report sorted by protocol

|  | Information displayed on report  |
|--|--|
| <ul> <li>ACTG PID</li> <li>ACTG Protocol</li> <li>ACTG Protocol Type</li> <li>Additive</li> <li>Assay Date</li> <li>Derivative</li> <li>Drug</li> <li>Global Spec ID</li> <li>Non ACTG PID/ID1</li> <li>Non ACTG Prot/ID2</li> <li>OPID</li> <li>Other Specimen ID</li> <li>Primary</li> <li>Project</li> <li>Received Date</li> <li>Run ID</li> <li>Specimen ID</li> <li>Specimen ID</li> <li>Specimen Time</li> <li>Time</li> <li>Time Unit</li> <li>Visit Unit</li> </ul> | <ul> <li>PID/ID1</li> <li>Specimen ID</li> <li>Global Specimen ID</li> <li>Other Spec ID</li> <li>Spec Date</li> <li>VID</li> <li>Spec Time</li> <li>Time/Unit</li> <li>Assay Date</li> <li>Run ID</li> <li>Pri</li> <li>Add</li> <li>Der</li> <li>Drug</li> <li>Conc</li> <li>Cens</li> <li>Rev by</li> <li>Rev Date</li> </ul> |

## **PK Summary with Assay Name**

Same as PK Summary Report but with additional Assay Name column

| Filter criteria    | Information displayed on report |
|--------------------|---------------------------------|
| ACTG PID           | PID/ID1                         |
| ACTG Protocol      | Specimen ID                     |
| ACTG Protocol Type | Global Specimen ID              |
| Additive           | Assay Name                      |
| Assay Date         | Spec Date                       |
| Derivative         | VID                             |
| Drug               | Spec Time                       |
| Global Spec ID     | Time/Unit                       |
| Non ACTG PID/ID1   | Assay Date                      |
| Non ACTG Prot/ID2  | Run ID                          |
| OPID               | • Pri                           |
| Other Specimen ID  | Add                             |
| Primary            | Der                             |
| Project            | Drug                            |
| Received Date      | Conc                            |
| Run ID             | Units                           |

| Filter criteria | Information displayed on report |  |
|-----------------|---------------------------------|--|
| Specimen Date   | Cens                            |  |
| Specimen ID     | Reviewed                        |  |
| Specimen Time   |                                 |  |
| Time            |                                 |  |
| Time Unit       |                                 |  |
| Visit Unit      |                                 |  |
| Visit Value     |                                 |  |

## PK Summary with Assay Name (exportable)

Exportable version of PK Summary with Assay Name report

| Filter criteria   | Information displayed on report   |
|---|---|
| ACTG PID ACTG Protocol ACTG Protocol Type Additive Assay Date Derivative Drug Global Spec ID Non ACTG PID/ID1 Non ACTG Prot/ID2 OPID Other Specimen ID Primary Project Received Date Run ID Specimen ID Specimen ID Specimen ID Specimen Time Time Time Time Time Unit Visit Unit | <ul> <li>Group/Prot</li> <li>Global Spec ID</li> <li>Assay Name</li> <li>Assay Date</li> <li>Run ID</li> <li>Drug</li> <li>Conc</li> <li>Units</li> <li>Lower Limit</li> <li>Upper Limit</li> <li>Censors Reviewed</li> </ul> |

## **Quick Add Templates**

## **Quick Add Template List Report**

This report provides a list of Quick Add templates available to your laboratory.

| Sorting                       | Available filters  | Information on report  |
|-------------------------------|--|--|
| Alphabetical by template name | <ul><li>ID2/protocol</li><li>project</li><li>Quick Add Template Name</li><li>Quick Add template Type</li></ul> | <ul> <li>Quick add template name</li> <li>Type (local or preset by<br/>Frontier Science</li> <li>[list of enrollment and<br/>specimen data that will be</li> </ul> |

| Sorting | Available filters | Information on report                                     |
|---------|-------------------|---|
|         |                   | populated by the template;<br>this will vary by template] |

## **RPID Reports**

#### **Random PID Report**

Provides a link between the original ACTG/IMPAACT PID and the RPID ID assigned during shipment import. This report is sensitive and is only available to the RPID importing lab.

| Sorting | Available filters   | Information on report  |
|---------|---|--|
| -       | <ul> <li>Import Date</li> <li>Original Specimen ID</li> <li>Received Shipment<br/>Number</li> <li>Shipment Number</li> <li>Specimen ID</li> </ul> | <ul> <li>PID</li> <li>RPID</li> <li>Other Specimen ID</li> <li>Spec Date</li> <li>Received Batch</li> <li>Sending Lab</li> <li>Contact</li> <li>Phone</li> </ul> |

#### **RPID Specimen Request Report**

Report displaying the RPID values assigned to various request numbers in the RPID Requests module.

| Sorting | Available filters  | Information on report  |
|---------|--|--|
| -       | <ul> <li>Import Date</li> <li>Received Shipment<br/>Number</li> <li>Request Number</li> <li>Shipment Number</li> </ul> | <ul> <li>PID</li> <li>RPID</li> <li>Other Specimen ID</li> <li>Shipment Number</li> <li>Received Batch</li> <li>Sending Lab</li> <li>Contact</li> <li>Phone</li> </ul> |

## **Shipping reports**

## Daily imported specimen log report

Shows a list of specimens for a project-protocol combination that have been received from another laboratory in a shipping file.

| Sorting  | Filter criteria                                    | Information displayed on report  |
|--|--|--|
| This report is sorted by project-protocol combination, and then by specimen date | ACTG protocol     ACTG protocol type     Clinic ID | <ul><li>Specimen ID</li><li>First global specimen ID</li><li>Second global specimen ID</li></ul> |

| Sorting | Filter criteria   | Information displayed on report  |
|---------|---|--|
|         | Condition Global specimen ID Import date Non ACTG PID/ID1 Non ACTG Prot/ID2 project SID/ID3 Sending lab ID Specimen date Specimen ID Visit unit Visit value | Clinic PID/ID1 SID/ID3 Visit Specimen date Import date Sending laboratory Specimen condition |

## **Detailed imported specimen report**

Shows specimens received through a shipping file from another laboratory, sorted by shipment number.

| Sorting   | Filter criteria  | Information displayed on report  |
|---|--|--|
| This report is sorted by shipment number, and then by specimen ID | ACTG PID     ACTG protocol     ACTG protocol type     Additive     Condition     Derivative     Import date     Laboratory ID     Non ACTG Prot/ID2     Other specimen ID     Primary     project     Shipment number     Specimen date     Specimen ID     Specimen time     Sub additive/derivative     Time     Time unit     Visit unit     Visit value     Volume     Volume unit | Import batch no. (the shipment number) Import date Sending laboratory Specimen ID project/Prot Primary Additive Derivative Sub additive/derivative PID/ID1 VID Condition Volume Specimen date Specimen time Other specimen ID Time/Time unit |

## **Imported specimen report - summary**

Shows the number of a specific type of specimen received as part of a shipment from another laboratory.

| Sorting                                  | Filter criteria  | Information displayed on report  |
|--|--|--|
| This report is sorted by shipment number | <ul> <li>ACTG protocol</li> <li>ACTG protocol type</li> <li>Additive</li> <li>Derivative</li> <li>Import date</li> <li>Non ACTG prod/ID2</li> <li>Primary</li> <li>project</li> <li>Sending laboratory ID</li> <li>Shipment number</li> <li>Sub additive/derivative</li> </ul> | <ul> <li>Sending laboratory</li> <li>Batch number (shipment number)</li> <li>project-protocol combination</li> <li>Import date</li> <li>Primary</li> <li>Additive</li> <li>Derivative</li> <li>Sub additive/derivative</li> <li>Count</li> </ul> |

## **Lab 263 Summary Detail of Shipped Specimens**

Provides a summary of shipped specimens for lab 263.

| Filter criteria   | Information displayed on report  |
|---|--|
| <ul> <li>Additive</li> <li>Derivative</li> <li>Destination Lab</li> <li>Primary</li> <li>Project</li> <li>Received Date</li> <li>Shipment Date</li> <li>Specimen Date</li> <li>Sub Add/Der</li> </ul> | <ul> <li>Project</li> <li>Destination Lab</li> <li>Primary</li> <li>Additive</li> <li>Derivative</li> <li>Sub A/D</li> <li>Tube count</li> <li>Total Volume</li> </ul> |

## Shipped specimen report - detail report

Lists the specimens within a shipment, organized by shipment number.

| Sorting  | Filter criteria  | Information displayed on report   |
|--|--|---|
| This report is sorted by shipment number, and then by global specimen ID | ACTG Protocol ACTG Protocol Type Additive Clinic ID Condition Derivative Destination Lab NON ACTG PID/ID1 NON ACTG PROT/ID2 Other Spec ID Primary project Shipment Date Shipment Number Shipping Temp. Spec ID Spec ID Specimen Time | <ul> <li>Shipment batch number</li> <li>Shipment date</li> <li>Laboratory shipped to</li> <li>Clinic</li> <li>Specimen ID</li> <li>Other specimen ID</li> <li>project/prot</li> <li>Global specimen ID</li> <li>PID/ID1</li> <li>VID</li> <li>Specimen Date</li> <li>Specimen time</li> <li>Primary</li> <li>Additive</li> <li>Derivative</li> <li>Sub additive/derivative</li> <li>Volume</li> </ul> |

| Sorting | Filter criteria   | Information displayed on report |
|---------|---|---------------------------------|
|         | <ul><li>Sub Add/Der</li><li>Visit Unit</li><li>Visit Value</li><li>Volume</li><li>Volume Unit</li></ul> | • Condition                     |

## **Shipped specimen report - summary report**

Shows the number of specimens of a specific derivative type on a shipment that was sent.

| Sorting  | Filter criteria  | Information displayed on report   |
|--|--|---|
| This report is sorted by shipment number, and then grouped by derivative | <ul> <li>ACTG protocol</li> <li>ACTG protocol type</li> <li>Clinic ID</li> <li>Derivative</li> <li>Destination laboratory</li> <li>Non ACTG prot/ID2</li> <li>project</li> <li>Shipment date</li> <li>Shipment number</li> <li>Shipping temperature</li> </ul> | <ul> <li>Batch number</li> <li>Ship date</li> <li>Destination</li> <li>project/protocol</li> <li>Derivative</li> <li>Number of specimens shipped</li> </ul> |

## **Shipping laboratory contact report**

Shows contact information for a given laboratory.

| Sorting                                | Filter criteria | Information displayed on report  |
|--|-----------------|--|
| This report is sorted by laboratory ID | Laboratory ID   | <ul> <li>Laboratory ID</li> <li>Laboratory name</li> <li>Contact (with telephone number and email address)</li> <li>Address</li> <li>Room number</li> <li>Country</li> <li>Laboratory telephone number</li> <li>Laboratory fax number</li> </ul> |

## **Specimens Marked for Shipping**

Provides a list of specimens that are marked for shipping.

| Filter criteria                 | Information displayed on report |
|---------------------------------|---------------------------------|
| ACTG PID     ACTG Protocol      | PID/ID1     Specimen ID         |
| ACTG Protocol Type              | Global Specimen ID              |
| ACTG Protocol Type     Additive | Global Specimen ID     VID      |

| Filter criteria    | Information displayed on report |
|--------------------|---------------------------------|
| Derivative         | • Pri                           |
| Intended Ship Date | Add                             |
| Marked in Storage  | • Der                           |
| Non ACTG PID/ID1   | Sub A/D                         |
| Non ACTG Prot/ID2  | Spec Date                       |
| Primary            | Category                        |
| Project            | Intended Ship Date              |
| Shipment Number    | Intended Destination            |
| Specimen ID        | Batch Number                    |
| Sub Add/Der        | Overdue?                        |
| Visit Unit         |                                 |
| Visit Value        |                                 |

# **Specimen reports**

### Cell yield QA/QC summary report

Provides a summary cell yield for viable PBMCs, along with a graph of cell yield by date, mean, standard deviation, and range.

| Sorting                                   | Filter criteria   | Information displayed on report  |
|---|---|--|
| This report is sorted by collection date. | <ul> <li>ACTG protocol</li> <li>ACTG protocol type</li> <li>Non ACTG prot/ID2</li> <li>project</li> <li>Specimen date</li> <li>Technician initials</li> </ul> | <ul> <li>Patid</li> <li>Collection date</li> <li>Collection time</li> <li>Tech</li> <li>Additive</li> <li>Total cell count (×10<sup>6</sup>)</li> <li>Total volume</li> <li>Comments (from Specimen Management)</li> <li>Cell yield (×10<sup>6</sup>)</li> </ul> |

### **Lab 263 Processing Report**

Provides processing information (processing date, amount aliquoted, etc.) for primaries and aliquots for lab 263.

| Sorting  | Filter Criteria  | Information Displayed on Report  |
|--|--|--|
| This report is sorted by project-protocol combination, then by collection date, and then grouped by primary. | <ul> <li>ACTG Protocol</li> <li>Additive</li> <li>Available</li> <li>Clinic ID</li> <li>Condition</li> <li>Derivative</li> <li>Import Date</li> <li>Non ACTG Prot/ID2</li> <li>Primary</li> <li>Project</li> </ul> | <ul> <li>Global Spec ID</li> <li>Add</li> <li>Der</li> <li>Other Spec ID</li> <li># of Tubes</li> <li>Volume</li> <li># of Aliq.</li> <li>Study</li> <li>Visit</li> <li>Processing Instructions</li> </ul> |

| Sorting | Filter Criteria  | Information Displayed on Report  |
|---------|--|--|
|         | <ul> <li>Received Date</li> <li>Shipment Date</li> <li>SID/ID3</li> <li>Specimen Date</li> <li>Specimen Type</li> <li>Stored</li> <li>Sub Add/Der</li> </ul> | <ul> <li>Cell Count</li> <li>Amt Aliq'd</li> <li>Processor's Initials</li> <li>Date/Time Processed</li> <li># of Unused Tubes</li> </ul> |

# **Lab 263 Summary of Specimens**

| Filter criteria  | Information displayed on report  |
|--|--|
| <ul> <li>ACTG PID</li> <li>ACTG Protocol</li> <li>Additive</li> <li>Available</li> <li>Clinic ID</li> <li>Condition</li> <li>Derivative</li> <li>Import Date</li> <li>Non ACTG PID/ID1</li> <li>Non-ACTG Prot/ID2</li> <li>Primary</li> <li>project</li> <li>Received Date</li> <li>Received Time</li> <li>Shipment Date</li> <li>SID/ID3</li> <li>Specimen Date</li> <li>Specimen Time</li> <li>Specimen Type</li> <li>Stored</li> <li>Sub additive/derivative</li> </ul> | <ul> <li>project</li> <li>Primary</li> <li>Additive</li> <li>Derivative</li> <li>Sub additive/derivative</li> <li>Participant count</li> <li>Tube count</li> <li>Total Volume</li> </ul> |

# **Primary specimens received report**

Provides a summary of all primary specimens within a laboratory's database, and the total number of primaries.

| Sorting  | Filter criteria  | Information displayed on report   |
|--|--|---|
| This report is sorted by project-protocol combination, and then by specimen ID | ACTG PID     ACTG protocol     ACTG protocol type     Additive     Clinic ID     Condition     Import date     Non ACTG PID/ID1     Non ACTG Prot/ID2     OPID | Specimen ID     project/protocol     PID/ID1     Clinic     Global specimen ID     Visit     Specimen date     Received date     Primary     Additive |

| Sorting | Filter criteria  | Information displayed on report  |
|---------|--|--|
|         | Other specimen ID Primary project Received date SID/ID3 Specimen date Specimen ID Visit unit Visit value Volume Volume Volume unit | Specimen condition     Volume     Other specimen ID     Comments (from the Specimen Management page) |

### **Specimen count report**

Should the number of aliquots entered, stored, and shipped by your laboratory by project-protocol combination, as well as the total across all projects and protocols.

| Sorting   | Filter criteria  | Information displayed on report   |
|---|--|---|
| This report is sorted by project-protocol combination | <ul> <li>ACTG protocol</li> <li>ACTG protocol type</li> <li>Clinic ID</li> <li>Derivative</li> <li>project</li> <li>Specimen date</li> </ul> | <ul> <li>project/protocol<br/>combination</li> <li>Number of aliquots entered<br/>into LDMS</li> <li>Number of currently stored<br/>aliquots</li> <li>Number of shipped aliquots</li> </ul> |

### Specimen log report

Provides detailed information about every primary and aliquot within a laboratory's database.

| Sorting  | Filter criteria  | Information displayed on report  |
|--|--|--|
| This report is sorted by project-protocol combination, then by collection date, and then grouped by primary. | ACTG PID     ACTG protocol     ACTG protocol type     Additive     Clinic ID     Condition     Derivative     Global specimen ID     Import date     Non ACTG PID/ID1     Non ACTG Prot/ID2     OPID     Other specimen ID     Primary     project     Received date | <ul> <li>PID/ID1</li> <li>project/prot</li> <li>SID/ID3</li> <li>VID</li> <li>Clinic</li> <li>OPID</li> <li>Primary specimen ID</li> <li>Global specimen ID</li> <li>Specimen time</li> <li>Specimen date</li> <li>Received date</li> <li>Primary volume</li> <li>Time/time unit</li> <li>Other specimen ID</li> <li>Comments [for primary specimen]</li> <li>Aliquot specimen ID</li> </ul> |

| Sorting | Filter criteria   | Information displayed on report   |
|---------|---|---|
|         | SID/ID3 Shipped Specimen date Specimen ID Specimen time Sub add/der System entry date Test ordered Time Time Time unit Visit unit Visit value Volume Volume Volume unit | Global specimen ID Other specimen ID Primary/additive Derivative/Sub-additive/derivative Current volume Condition project/prot Test(s) ordered Shipped Comments |

# **Specimen processing report**

Provides processing information (processing date, frozen date, etc.) for primaries and aliquots in a laboratory's database.

| Sorting  | Filter criteria  | Information displayed on report  |
|--|--|--|
| This report is sorted by project-protocol combination, then by collection date, and then grouped by primary. | ACTG PID/ID1     ACTG protocol     ACTG protocol type     Additive     Derivative     Global specimen ID     Import date     Non ACTG PID/ID1     Non ACTG Prot/ID2     Other specimen ID     Primary     project     Received date     SID/ID3     Specimen ID     Visit unit     Visit value | <ul> <li>PID/ID1</li> <li>project/Prot</li> <li>SID/ID3</li> <li>VID</li> <li>Clinic</li> <li>OPID</li> <li>Primary specimen ID</li> <li>Global specimen ID</li> <li>Other specimen ID</li> <li>Primary</li> <li>Additive</li> <li>Volume</li> <li>Specimen date</li> <li>Specimen time</li> <li>Received date</li> <li>Received time</li> <li>Total cell count</li> <li>Proc[essing] date</li> <li>Proc[essed] by</li> <li>Aliquot specimen ID</li> <li>Global specimen ID</li> <li>Other specimen ID</li> <li>Other specimen ID</li> <li>Derivative</li> <li>Sub-additive/derivative</li> <li>Volume</li> <li>project/prot</li> <li>Frozen date</li> <li>Frozen time</li> <li>Proc[essing] date</li> </ul> |

| Sorting | Filter criteria | Information displayed on report                             |
|---------|-----------------|---|
|         |                 | <ul><li>Proc[essing] time</li><li>Proc[essing] by</li></ul> |

### Specimens for a given project report

Shows the aliquots for a given project-protocol combination.

| Sorting  | Filter criteria  | Information displayed on report  |
|--|--|--|
| This report is grouped by project-protocol combination, and then sorted by ID1, and then by specimen ID. | ACTG PID     ACTG protocol     ACTG protocol type     Additive     Clinic ID     Derivative     Import date     Non ACTG PID/ID1     Non ACTG prot/ID2     OPID     Other specimen ID     Primary     project     Received date     SID/ID3     Specimen date     Specimen ID     Specimen ID     Specimen time     Sub additive/derivative     Visit unit     Visit value | <ul> <li>Protocol (and project)</li> <li>PID/ID1</li> <li>OPID</li> <li>Specimen ID</li> <li>Global specimen ID</li> <li>Specimen date</li> <li>Specimen time</li> <li>Primary</li> <li>Additive</li> <li>Derivative</li> <li>Sub additive-derivative</li> <li>Received date</li> <li>Received time</li> <li>Import date</li> <li>VID (visit value and unit)</li> <li>Other specimen ID</li> </ul> |

# Specimens for a given project 2

Shows the aliquots for a given project-protocol combination in aggregate form, with the number of aliquots with a given specimen ID.

| Sorting   | Filter criteria   | Information displayed on report   |
|---|---|---|
| This report is grouped by project-protocol combination, then grouped by Specimen ID, and then sorted by ID1, and then by specimen ID. | ACTG PID     ACTG protocol     ACTG protocol type     Additive     Clinic ID     Derivative     Import date     Non ACTG PID/ID1     Non ACTG prot/ID2     OPID     Other specimen ID     Primary     project | Protocol (and project) PID/ID1 OPID Specimen ID Count Specimen date Specimen time SID/ID3 Primary Additive Derivative Sub additive-derivative Received date |

| Sorting | Filter criteria   | Information displayed on report  |
|---------|---|--|
|         | <ul> <li>Received date</li> <li>SID/ID3</li> <li>Specimen date</li> <li>Specimen ID</li> <li>Specimen time</li> <li>Sub additive/derivative</li> <li>Visit unit</li> <li>Visit value</li> </ul> | Received time Import date VID (visit value and unit) Other specimen ID |

### Time to freeze QA/QC summary report

Shows the time to freeze for aliquots, along with a graph and summary of overall time to freeze.

| Sorting <sup>1</sup> | Filter criteria   | Information displayed on report  |
|----------------------|---|--|
| -                    | <ul> <li>ACTG protocol</li> <li>ACTG protocol type</li> <li>Non ACTG prot/ID2</li> <li>project</li> <li>Specimen date</li> <li>Tech initials</li> </ul> | <ul> <li>Patid</li> <li>Collection date</li> <li>Collection time</li> <li>Frozen date</li> <li>Frozen time</li> <li>Time to freeze (in minutes)</li> <li>Tech</li> <li>Additive</li> <li>Derivative</li> <li>Comments</li> </ul> |

### Time to process QA/QC summary report

Shows the time to process for aliquots, along with a graph and summary of overall time to process.

| Sorting <sup>2</sup> | Filter criteria   | Information displayed on report  |
|----------------------|---|--|
| -                    | <ul> <li>ACTG protocol</li> <li>ACTG protocol type</li> <li>Non ACTG prot/ID2</li> <li>project</li> <li>Specimen date</li> <li>Tech initials</li> </ul> | <ul> <li>Patid</li> <li>Collection date</li> <li>Collection time</li> <li>Process time</li> <li>Frozen time</li> <li>Time to process (in minutes)</li> <li>Tech</li> <li>Additive</li> <li>Derivative</li> <li>Comments</li> </ul> |

You must specify a sample type to report on to generate this report. You can generate the report for PBMCs, plasma, or both.

You must specify a sample type to report on to generate this report. You can generate the report for PBMCs, plasma, or both.

# Storage reports

### Specimens in storage per PID report

Shows the number for a given project-participant combination that have a storage location assigned in a laboratory's database.

| Sorting  | Filter criteria  | Information displayed on report  |
|--|--|--|
| This report is sorted by project, and then by participant. | <ul> <li>ACTG PID/ID1</li> <li>ACTG protocol</li> <li>ACTG protocol type</li> <li>Non ACTG PID/ID1</li> <li>Non ACTG prot/ID2</li> <li>OPID</li> <li>Primary</li> <li>project</li> </ul> | <ul><li>project</li><li>PID</li><li>Count (number of aliquots)</li></ul> |

### Specimens not in storage report

Shows specimens that are available in a laboratory's database, but have not had a storage location assigned on the **Storage** page.

| Sorting   | Filter criteria   | Information displayed on report  |
|---|---|--|
| This report is grouped by project-protocol combination, and then by specimen ID, and then sorted by global specimen ID. | ACTG protocol     ACTG protocol type     Additive     Condition     Derivative     Non ACTG PID/ID1     Non ACTG prot/ID2     Primary     project     Specimen ID     Sub additive/derivative     Visit unit     Visit value     Volume     Volume unit | <ul> <li>project/prot</li> <li>Specimen ID</li> <li>Global specimen ID</li> <li>PID/ID1</li> <li>Specimen date</li> <li>Pri[mary]</li> <li>Add[itive]</li> <li>Der[ivative]</li> <li>Sub a[dditive]/d[erivative]</li> <li>VID (visit unit and value)</li> <li>Volume</li> <li>Condition</li> <li>Status</li> </ul> |

### Specimens remaining in storage report

Shows the number of aliquots for a given participant with a storage location assigned in a laboratory's database.

| Sorting  | Filter criteria  | Information displayed on report                                       |
|--|--|---|
| This report is grouped and sorted by project, ID1, and protocol. | <ul><li>ACTG PID/ID1</li><li>ACTG protocol</li><li>ACTG protocol type</li><li>Additive</li></ul> | <ul><li>project</li><li>PID</li><li>Protocol</li><li>Volume</li></ul> |

| Sorting | Filter criteria                       | Information displayed on report |
|---------|---------------------------------------|---------------------------------|
|         | Non ACTG PID/ID1                      | Volume unit                     |
|         | <ul> <li>Non ACTG prot/ID2</li> </ul> | Derivative                      |
|         | OPID                                  | Aliquots remaining in           |
|         | Primary                               | storage                         |
|         | project                               |                                 |
|         | Specimen date                         |                                 |

### **Storage container location report**

Shows the storage location for all containers in a laboratory's database.

| Sorting   | Filter criteria  | Information displayed on report  |
|---|--|--|
| This report is sorted by storage unit name, and then by container position. | <ul> <li>ACTG protocol</li> <li>ACTG protocol type</li> <li>Freezer name</li> <li>Level name</li> <li>Non ACTG prot/ID2</li> <li>project</li> <li>Sublevel name</li> </ul> | <ul><li>Storage location</li><li>Container name</li><li>Position</li></ul> |

# Storage count report by freezer

Shows the number of specimens stored in a given storage unit for a given project-protocol combination.

| Sorting  | Filter criteria   | Information displayed on report  |
|--|---|--|
| This report is sorted by storage unit, and then by project-protocol combination, and then by derivative. | <ul> <li>ACTG PID/ID1</li> <li>ACTG protocol</li> <li>ACTG protocol type</li> <li>Derivative</li> <li>Non ACTG PID/ID1</li> <li>Non ACTG prot/ID2</li> <li>project</li> </ul> | <ul> <li>Freezer</li> <li>project/protocol</li> <li>Primary</li> <li>Additive</li> <li>Derivative</li> <li>Sub a[dditive]/d[erivative]</li> <li>Count</li> </ul> |

# Storage detail report

Shows detailed information about the contents of containers in storage units.

| Sorting  | Filter criteria   | Information displayed on report   |
|--|---|---|
| This report is sorted by storage location, and then by storage position. | ACTG PID     ACTG protocol     ACTG protocol type     Additive     Derivative     Import date     Non ACTG PID/ID1     Non ACTG Prot/ID2     OPID | <ul> <li>Storage location</li> <li>Specimen ID</li> <li>Global spec[imen] ID</li> <li>project/prot</li> <li>PID/ID1</li> <li>VID/unit</li> <li>Prim[ary]</li> <li>Add[itive]</li> <li>Der[ivative]</li> </ul> |

| Sorting | Filter criteria  | Information displayed on report  |
|---------|--|--|
|         | Other specimen ID Primary project Received date SID/ID3 Shipment number Specimen date Specimen ID Specimen time Stored date Sub add[itive]/der[ivative] Visit unit Visit value Volume Volume Volume unit | <ul> <li>Sub a[dditive]/d[erivative]</li> <li>Volume</li> <li>Ship</li> <li>Storage date</li> <li>Spec[cimen] date</li> <li>Time</li> <li>Clinic</li> <li>Pos[ition]</li> <li>Other specimen ID</li> </ul> |

# **Test Result Reports**

### **Abbott Realtime HIV1 Assay Report**

| Filtering Criteria   | Information Displayed on Report   |
|--|---|
| <ul> <li>ACTG PID/ID1</li> <li>ACTG/Protocol</li> <li>ACTG Protocol Type</li> <li>Assay Date</li> <li>Clinic ID</li> <li>Global Spec ID</li> <li>Non-ACTG PID/ID1</li> <li>Non-ACTG Prot/ID2</li> <li>Other Specimen ID</li> <li>Primary</li> <li>Project</li> <li>Run ID</li> <li>SID/ID3</li> <li>Specimen ID</li> </ul> | Group/protocol PID SID Visit PRI Specimen ID Other Spec ID Spec Date Assay Date RUN ID Dilution Result System Censor Run System Censor Run User Censor Group/Pro Total Grand Total  PID Grand Final System Final Sys |

# **Abbott Realtime HIV1 Patient Report**

| Filtering Criteria   | Information Displayed on Report |
|--|---------------------------------|
| The state of the s |                                 |

| ACTG PID/ID1       | • Patient                          |
|--------------------|------------------------------------|
| ACTG/Protocol      | • SID                              |
| ACTG Protocol Type | <ul> <li>Group/Protocol</li> </ul> |
| Assay Date         | Specimen Date                      |
| Derivative         | Visit                              |
| Global Spec ID     | Clinic Info                        |
| Non-ACTG PID/ID1   | • Fax                              |
| Non-ACTG Prot/ID2  | Testing Lab Info                   |
| Other Specimen ID  | Specimen ID                        |
| Primary            | Received Date                      |
| Received Date      | Primary                            |
| Run ID             | Global Spec ID                     |
| • SID/ID3          | Received Time                      |
| Specimen Date      | Additive                           |
| Specimen ID        | Other Spec ID                      |
|                    | Sample Condition                   |
|                    | Derivative                         |
|                    | Type of Assay                      |
|                    | Assay Date                         |
|                    | Sample Prep Tech                   |
|                    | Amplification Tech                 |
|                    | Input Volume                       |
|                    | Data Transfer Tech                 |
|                    | Results:                           |
|                    | Run Comment                        |
|                    | Sample Comment                     |
|                    | Reportable Page                    |
|                    | . top of cable 1 age               |

### **Abbott Repeat and Censored Run/Samples**

| Filtering Criteria | Information Displayed on Report |
|--------------------|---------------------------------|
| ACTG PID/ID1       | Group/Prot                      |
| ACTG/Protocol      | System Censor                   |
| ACTG Protocol Type | Run System Censor               |
| Assay Date         | User Censor                     |
| Global Spec ID     | Run User Censor                 |
| Non-ACTG PID/ID1   | Specimen ID                     |
| Non-ACTG Prot/ID2  | Global Spec ID                  |
| Other Specimen ID  | • PID                           |
| Primary            | Spec Date                       |
| Received Date      | RUN ID                          |
| Run ID             | Run Valid/Invalid               |
| • SID/ID3          | Assay Date                      |
| Specimen Date      | Dilution                        |
| Specimen ID        | Result                          |

# **Abbott SARS-COV-2 Quant Assay Report**

Displays specimen details and results from Abbott SARS-COV-2 Quant runs in the Test Results module.

| Filter criteria  | Information displayed on report  |
|--|--|
| <ul> <li>ACTG PID/ID1</li> <li>ACTG Protocol</li> <li>ACTG Protocol Type</li> <li>Assay Date</li> <li>Clinic ID</li> <li>Global Spec ID</li> <li>Non ACTG PID/ID1</li> <li>Non ACTG Prot/ID2</li> <li>Other Specimen ID</li> <li>Primary</li> <li>Project</li> <li>Run ID</li> <li>SID/ID3</li> <li>Specimen ID</li> <li>Specimen ID</li> <li>Specimen List</li> </ul> | Group/Prot PID SID Visit PRI Specimen ID Other Spec ID Global Spec ID Spec Date Assay Date RUN ID Result System Censor Run System Censor Run user Censor |

# **Abbott SARS-COV-2 Quant Patient Report**

| ilter criteria Information displayed on report |                    |
|--|--------------------|
| ACTG PID/ID1                                   | Participant        |
| ACTG Protocol                                  | • SID              |
| <ul> <li>ACTG Protocol Type</li> </ul>         | Group/Protocol     |
| Assay Date                                     | Specimen Date      |
| Derivative                                     | • Visit            |
| Global Spec ID                                 | Clinic Info        |
| Non ACTG PID/ID1                               | • Fax              |
| Non ACTG Prot/ID2                              | Testing Lab Info   |
| Other Specimen ID                              | Specimen ID        |
| • Primary                                      | Global Spec ID     |
| Project  | Other Spec ID      |
| Received Date                                  | Received Date      |
| Run ID   | Received Time      |
| SID/ID3  | Sample Condition   |
| Specimen Date                                  | Primary            |
| Specimen ID                                    | Additive           |
| Specimen List                                  | Derivative         |
|  | Type of Assay      |
|  | Assay Date         |
|  | Input Volume       |
|  | Sample Prep Tech   |
|  | Amplification Tech |
|  | Data Transfer Tech |
|  | Results            |
|  | Log copies/mL      |
|  | Run Comment        |
|  | Sample Comment     |

| Filter criteria | Information displayed on report                     |
|-----------------|---|
|                 | Assay Reportable Range and Dilution     Information |

### **IQA Cryopreservation Patient Report**

An individualized report per specimen showing associated participant details along with detailed test results including censor codes, run limits, and a final calculated result.

| Filter criteria              | Information displayed on report |
|------------------------------|---------------------------------|
| • ACTG PID/ID1               | Group/Study                     |
| ACTG Protocol                | Run ID                          |
| ACTG Protocol Type           | Assay Tech                      |
| Assay Date                   | • PID                           |
| • Derivative                 | Spec No                         |
| Global Spec ID               | GlobalSpec ID                   |
| Non ACTG PID/ID1             | Spec Date                       |
| Non ACTG Prot/ID2            | VID                             |
| On Cryopreservation Test Run | Primary                         |
| Other Specimen ID            | Additive                        |
| Primary                      | Derivative                      |
| Project                      | Sub A/D                         |
| Received Date                | Data Entered By                 |
| Run ID                       | Data Entry Date                 |
| SID/ID3                      |                                 |
| Specimen Date                |                                 |
| Specimen ID                  |                                 |
| Specimen List                |                                 |

### **TaqMan HCV Repeat and Censored Run/Samples**

| Filtering Criteria  | Information Displayed on Report  |
|---|--|
| <ul> <li>ACTG PID/ID1</li> <li>ACTG/Protocol</li> <li>ACTG Protocol Type</li> <li>Assay Date</li> <li>Global Spec ID</li> <li>Non-ACTG PID/ID1</li> <li>Non-ACTG Prot/ID2</li> <li>Project</li> <li>Run ID</li> <li>Specimen Date</li> <li>Specimen ID</li> </ul> | <ul> <li>Group/Prot</li> <li>System Censor</li> <li>Run System Censor</li> <li>User Censor</li> <li>Run User Censor</li> <li>Specimen ID</li> <li>Global Spec ID</li> <li>PID/ID1</li> <li>Spec Date</li> <li>RUN ID</li> <li>Run Valid/Invalid</li> <li>Result</li> <li>Assay Date</li> </ul> |

### TaqMan HIV-1 Repeat and Censored Run Samples

| Filtering Criteria | Information Displayed on Report |
|--------------------|---------------------------------|

| <ul> <li>ACTG PID/ID1</li> <li>ACTG/Protocol</li> <li>ACTG Protocol Type</li> <li>Assay Date</li> <li>Global Spec ID</li> <li>Non-ACTG PID/ID1</li> <li>Non-ACTG Prot/ID2</li> <li>Project</li> <li>Run ID</li> <li>Specimen Date</li> <li>Specimen ID</li> </ul> | <ul> <li>Group/Prot</li> <li>System Censor</li> <li>Run System Censor</li> <li>User Censor</li> <li>Run User Censor</li> <li>Specimen ID</li> <li>Global Spec ID</li> <li>PID/ID1</li> <li>Spec Date</li> <li>RUN ID</li> <li>Run Valid/Invalid</li> <li>Result</li> <li>Assay Date</li> </ul> |
|---|--|
|---|--|

# **TaqMan Realtime HCV Assay Report**

| Filtering Criteria  | Information Displayed on Report  |
|---|--|
| ACTG PID/ID1 ACTG/Protocol ACTG Protocol Type Assay Date Clinic ID Global Spec ID Non-ACTG PID/ID1 Non-ACTG Prot/ID2 Other Specimen ID Primary Project Run ID SID/ID3 Specimen ID Specimen ID Specimen ID | <ul> <li>Group/Prot</li> <li>SID/ID3</li> <li>VID</li> <li>PRI</li> <li>Specimen ID</li> <li>Other Spec ID</li> <li>Global Spec ID</li> <li>Spec Date</li> <li>Assay Date</li> <li>RUN ID</li> <li>Ver</li> <li>Result</li> <li>System Censor</li> <li>Run System Censor</li> <li>Run System Censor</li> </ul> |
| l '   |  |

# **TaqMan Realtime HCV Patient Report**

| 1 | Filtering Criteria | Information Displayed on Report |
|---|--------------------|---------------------------------|
|   | Filtering Criteria | Information Displayed on Report |

# TaqMan Realtime HIV-1 Assay Report

| Filtering Criteria | Information Displayed on Report |
|--------------------|---------------------------------|
| ACTG PID/ID1       | PID/ID1                         |
| ACTG/Protocol      | • SID/ID3                       |
| ACTG Protocol Type | • VID                           |
| Assay Date         | • PRI                           |
| Clinic ID          | Specimen ID                     |
| Global Spec ID     | Other Spec ID                   |
| Non-ACTG PID/ID1   | Global Sec ID                   |
| Non-ACTG Prot/ID2  | Spec Date                       |
| Other Specimen ID  | Assay Date                      |
| Primary            | RUN ID                          |
| Project            | • Ver                           |
| Run ID             | Result                          |
| SID/ID3            | System Censor                   |
| Specimen Date      | Use Censor                      |
| Specimen ID        | Run System Censor               |
| Version            | Run User Censor                 |

# **TaqMan Realtime HIV-1 Patient Report**

|                    | Î                               |
|--------------------|---------------------------------|
| Filtering Criteria | Information Displayed on Report |

| ACTG PID/ID1 ACTG/Protocol ACTG Protocol Type Assay Date Derivative Global Spec ID Non-ACTG PID/ID1 Non-ACTG Prot/ID2 Other Specimen ID Primary Project Received Date Run ID SID/ID3 Specimen ID Version | <ul> <li>Patient</li> <li>SID</li> <li>Group/Protocol</li> <li>Specimen Date</li> <li>Visit</li> <li>Clinic Info</li> <li>Fax</li> <li>Testing Lab Info</li> <li>Specimen ID</li> <li>Global Spec ID</li> <li>Other Spec ID</li> <li>Received Date</li> <li>Received Time</li> <li>Sample Condition</li> <li>Primary</li> <li>Additive</li> <li>Derivative</li> <li>Type of Assay</li> <li>Assay Date</li> </ul> |
|--|--|
| Specimen ID  | <ul><li>Additive</li><li>Derivative</li><li>Type of Assay</li></ul>  |

# **Taqman Realtime Qual Assay Report**

# **TaqMan Realtime Qual Patient Report**

|                    | Î                               |
|--------------------|---------------------------------|
| Filtering Criteria | Information Displayed on Report |

### **WIHS**

# **Processing Log - Lab 263**

Displays specimen processing details and instructions based on WIHS processing codes.

| Filter criteria  | Information displayed on report  |
|--|--|
| <ul> <li>Non ACTG PID/ID1</li> <li>Specimen Date</li> <li>Specimen ID</li> </ul> | <ul> <li>Global Spec ID</li> <li>Der</li> <li>Other Spec ID</li> <li>Volume</li> <li>Number of Aliquots</li> <li>Study</li> <li>Processing Instructions</li> <li>Cell Count</li> <li>Amt</li> <li>Storage</li> <li>Processor's Initials</li> <li>Date/Time Processed/Frozen</li> <li>Unused Tubes</li> </ul> |

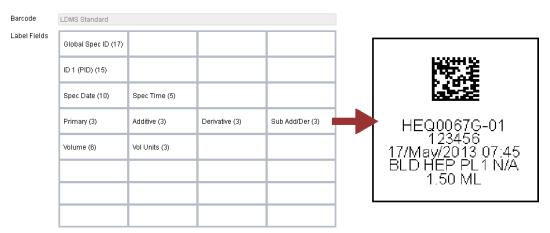
# **Labels**

# **Label formats**

Labels are setup like grids, with items that appear on the label occupying one cell.

Items that can appear are most of the information that can be found on the **Specimen Management** page, such as the primary type and the received date and time. Each row on a label can hold up to four items, and a label can have up to 8 rows.

Figure 52: Label definition and corresponding output



A label as it is set up in LDMS (left) compared to a generated label (right). The colored background shows how each item in the label definition matches up to the item on the label.

#### **Barcodes**

LDMS supports generating 2D barcodes on labels

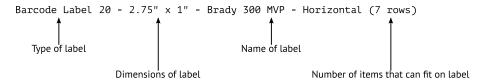
If a label has a barcode, it will appear as the first item on the label. The barcodes generated by LDMS contain the specimen's *global specimen ID*.

#### Label sizes

Labels can be generated in a variety of common sizes.

The label's name in LDMS indicates information about its size, manufacturer, and use.

Figure 53: Example label name



There are three types of labels available:

**Laser labels** For printing using a desktop printer with special 8.5-

by-11-inch paper

**Barcode labels** For printers specifically made for printing labels

**Dot matrix** For use with dot matrix printers

labels



**Note:** If you choose a label format that has a barcode, only barcode labels can be selected.

For laser and dot matrix labels, the size of the label refers to the label itself, not the paper it will be printed on. These labels are generated in US letter size paper (8.5-by-11-inches), with the assumption that the user is using special label paper that can be pealed apart. Barcode labels, on the other hand, require special printers and label paper that are designed exclusively for printing labels.

#### How to use a barcode reader with LDMS

LDMS supports using any Windows compatible barcode reader.

Barcode readers that are compatible with Windows will extract the global specimen ID from the 2D barcodes that are generated by LDMS. You can test this yourself by opening Notepad and scanning a barcode. Doing so will cause the global specimen ID from the barcode to appear in Notepad.

In any place in where you would need to type a global specimen ID, you can use a barcode reader to input the ID instead. Select the box where you want the global specimen ID to appear, and then scan the barcode. This can help avoid errors when trying to type out a global specimen ID using the keyboard.

# Printing labels anywhere

Specimen labels can be printed in many places in LDMS, not just the Labels page.

Labels can be printed almost anywhere where you can interact with specimens.

- On the Specimen Management page, click the down arrow next to any of the Edit buttons, and then select Print Labels. This will print all labels associated with that item. For example, if you select Edit Participant > Print labels, you will print the labels for all specimens associated with that participant. Likewise, if you click Edit [primary] > Print Labels, you will print all labels associated with that primary.
- Also on the Specimen Management page, when adding a participant using Quick Add, after a record is added a success message appears at the top of the page. Below it, click the button that says **Print labels**.
- On the Storage page, click the down arrow next to **Edit** button next to any storage item, and then click **Print Labels**. This will allow you to print labels for all specimens stored at that location.
- On the Pending Shipments page, click the down arrow next to the Edit/ Ship button next to a shipment, and then click Print Labels.

When creating labels, specimens that are not available will not be included.

# **Support for Labelscape-generated Barcodes**

The Labelscape LDMS-supported barcode is a data matrix 2D barcode that includes the ID1/PID & specimen date in the barcode.

In LDMS, there are two ways of scanning Labelscape barcodes. One method is to scan a barcode in **Specimen Management** and search for primary records. Another method is to scan a label into **Quick Add** which will populate the PID and specimen date.

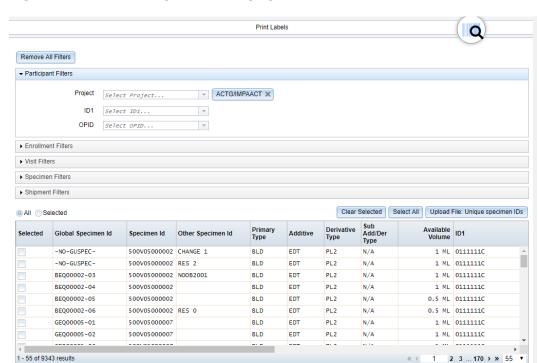
# Generating labels

While labels can be generated from many places, the **Print Labels** page allows you to generate labels from multiple sources.

#### **Background**

If you want to generate labels, you generally want to do so on either the **Specimen Management**, **Storage**, or **Shipping** page. On these pages, you can create labels for a specific participant, a particular shipping or storage container, or an individual shipment. These are the most common scenarios for which you will need to generate labels. You can also choose to upload a list of unique specimen IDs, which will set a specimen criteria to locate any sample with a unique specimen ID appearing in the list.

There may be times where you need to generate labels from a variety of sources. For example, you may want to print labels for all specimens received on a specific day.



Primary/Aliquot Creation Order

Figure 54: The label generation page



#### Note:

You may select up to 500 more specimens.

Labels can be generated for up to 500 specimens.

Sort Order

#### Steps

1. On the navigation bar, hover over **Labels**, and then click **Print Labels**.

Print Labels

- **2.** Optional: At the top of the **Print Labels** page, use filters to narrow down the list of specimens displayed.
- **3.** For each specimen you want to print a label, select the check box in the **Selected** column.
- Optional: Use dropdown next to Sort Order to specify the order in which labels are generated. This option is also available in the Print Labels window.
- 5. Click Print Labels.
- **6.** In the **Print Labels** window, select the following.

| Option  |  |
|---------|--|
| Project | Only shown if specimens from different projects were selected. |

**Option** 

**Use Defaults** If the project has a default label

format defined, selecting this check box will re-select that

default

**Format** The label format to use

**Size** The size of your label stock

Skip

**Sort Order** Order in which labels are sorted

#### 7. Click Generate Labels.

A PDF with the labels will be generated, which may open or you may be prompted to save it, depending on your browser's settings.

If generating labels for specimens from more than one project, a check will appear in the **Generated** column for that project. Repeat the previous steps with the next project.

# **Defining new label formats**

You can create new label formats.

#### **Steps**

- In the menu bar, hover over Labels, and then click Define Custom Labels.
- 2. Optional: Use the **Project** and **Format** lists to select a label as the basis for your new label.

This is not needed if you want to create a new label from scratch.

- 3. Click Create Format.
- **4.** In the **Name** box, enter a name for your new format.

If your new label format is based on an existing format, you must also select the check box next to **Copy Fields**.

5. Click Create Format.

#### Result

Your new label format will be created and will be available for selection.

#### After you are finished

You will want to customize your new label format and define the information that will appear on it.

# **Customizing label formats**

You can customize that information that appears on label formats created by your laboratory.

#### **Background**

New label formats are blank when they are created. You must customize the format to show the information that you want displayed. New labels and existing labels are modified the same way. Only label formats that your laboratory created can be modified. Label formats that are pre-defined in LDMS cannot be changed.

Name FRONTIER Barcode LDMS\_Standard Fields Global Spec ID (17) ID 1 (PID) (15) Spec Date (10) Spec Time (5) Primary (3) Additive (3) Derivative (3) Sub Add/Der (3) Volume (6) Vol Units (3) Column 1 Set Field Clear Field Ship Batch No. (8)

Figure 55: The label modification page

#### **Steps**

- In the menu bar, hover over Labels, and then click Define Custom Labels.
- 2. In the **Project** and **Format** lists, select a locally defined project and label format.
- 3. Click Modify Format.
- **4.** Modify the label format as needed.
  - To change the barcode options, select from the **Barcode** dropdown. There are various 1D and 2D options for the user to define.
  - To add a field to a label, select a box from the Fields section where
    you want the information to appear. Select an item from the list
    below the Fields section, and then click Set Field.

- To remove a field, select it from the Fields section, and then click Clear Field.
- 5. Click Modify Format.

# Setting the default label format for a project

The default label format and stock size can be set for individual projects.

#### **Background**

This means that when you print labels for that project, the default format and size will automatically be selected. If you typically use the same format and size for a specific project's labels, then you can save time by defining these defaults.

#### Steps

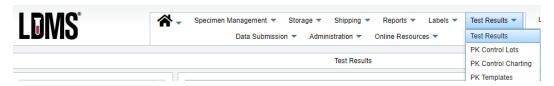
- 1. On the navigation menu, hover over **Labels**, and then click **Default Label Setup**.
- **2.** In the **Project** box, select the project to set defaults.
- **3.** In the **Format** box, select the applicable default label format.
- **4.** In the **Label** box, select the applicable default label stock size.
- 5. Click Add Default.
- **6.** Click **Save** to save the changes.

# **Test Results**

#### **Test Results**

The **Test Results** page is where you can manage test results from the various available assays. The application captures the complete audit history of test runs. The **Test Results** page shows all test runs for the lab, users can access this history and view the run from each phase.

Figure 56: Test Results Page



Users can create, edit, and delete committed and uncommitted test results on the Test Results page.

When viewing the Test Results page, you'll notice two main features on the page. First is the Main Window, second is the filters window. There is a third, less noticeable yet incredibly useful feature, and that is the Add Pending Results bar. We'll examine these features one by one after a note on assays themselves.

### **Test Supported in LDMS**

Below is a list of all tests supported in LDMS:

- Abbott RealTime HIV-1 RNA
- Abbott SARS-COV-2
- COBAS TaqMan HCV
- COBAS TaqMan HIV-1
- COBAS TaqMan HIV-1 Qual
- PK Assay

### Running an assay means reading data

The method of getting assay data into LDMS will vary, depending on the assay and available equipment at your laboratory. There are two ways to get assay data into LDMS:

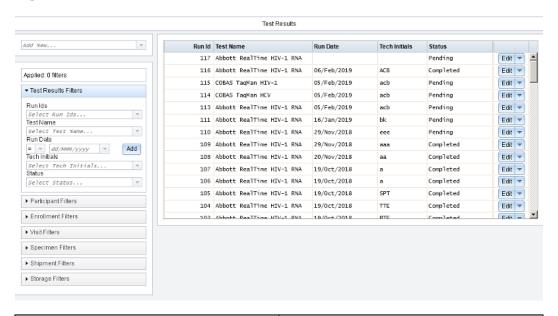
- 1. Read an output file that was created when the assay was run
- 2. Enter the assay results data by hand

LDMS does not really run your assay; it is more accurate to say that LDMS reads assay data, and then stores it. Reading assay output using LDMS is also sometimes referred to as "resulting" an assay in LDMS, since the intent is to get assay results into LDMS's database for long-term storage.

### **Test Results page**

The **Test Results** page is where you can access all test result runs and create new runs.

Figure 57: Test Results Main Window



| Column        | Description   |
|---------------|---|
| Run ID        | ID number assigned automatically  |
| Test name     | Name of assay selected when run was created   |
| Run date      | Date that the assay was run (or is expected to be run)  |
| Tech Initials | The assay technician who ran (or will run) the assay; not necessarily the data entry technician |
| Status        | Current status of the run   |

### Locating assay runs

Additional filters are available for locating assays on the **Test Results** page.

In additional to the regular filters, **Test Results Filters** are available on the **Test Results** page. These provide additional filters specifically for assays, such as locating tests by assay name.



**Note:** Deleted tests are not displayed by default. To see them, you must apply a **Status** filter set to Deleted.

#### **Censor Codes**

Censor codes can be assigned to specimens, controls and assay runs to indicate an issue or special condition.

For example, a censor code might indicate a contaminated specimen or equipment failure. *User censors* are those that are added by a user. *System censors* are those that are added automatically by LDMS when certain conditions are met.

There are two types of user and system censors:

specimen censors

These censor a single specimen on a run; other

specimens on the run are unaffected.

run censors

These censor the results of an entire assay run. They indicate that something went wrong with the entire test.

#### **Run Statuses**

Each run has a status that indicates what state the run is at and what action is needed next.

| Status    | Meaning  |  |
|-----------|--|--|
| Pending   | The run has been created, but is awaiting date entry (or data entry is in progress)  |  |
| Completed | Data entry is complete, but the run has not been reviewed  |  |
| Reviewed  | Data entry is complete and the run has been reviewed   |  |
| Deleted   | Run was created in error and has been removed. (Deleted runs are not displayed by default. You must apply a <b>Test Results Filter</b> for <b>Status</b> as <code>Deleted</code> to see them.) |  |

### **Creating New Test Result Runs**

Adding assays results involves selecting specimens that were tested, uploading a results file, and then matching results to specimens.

#### **Background**

There are some differences between supposed assay, such as additional assay-specific fields.

You can create an assay run and then click **Save** without adding specimens or results. The run will be saved with the status of pending and can be updated later.

#### **Steps**

- 1. From the LDMS menu, click **Test Results**.
- **2.** From the action menu, click **Add Pending Results**.
- 3. In the **Test Name** box, select an assay, and then click **Continue**

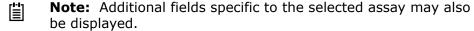


**Warning:** The assay cannot be changed later. If the wrong assay was selected, the run must be deleted and a new run created.

**4.** On the **General** tab, complete the details of the run.

| Field    | Usage  |
|----------|--|
| Run Date | The date that the test was run in the laboratory |

| Field              | Usage  |
|--------------------|--|
| Tech Initials      | The initials of the laboratory technician who completed the test |
| Data Transfer Tech | The initials of the person entering this data into LDMS          |



- **5.** Click the **Results** tab.
- **6.** (If needed) To add controls to the run, click **Add Control**.
- 7. Click **Add Specimens**.

Use the filters to narrow down the list of specimens. You can also upload a list of unique specimen IDs.

- **Tip:** Under **Specimen Filters**, there is a filter for **Assigned Test**. If you set the anticipate test for specimens when entering new specimens in LDMS, you can use this filter to quickly find and add specimens to test runs.
- **8.** Optional: To add additional specimens to the run that have not been entered into LDMS click the arrow next to **Add Specimens**, and then click **Add non-logged specimens**.

You can enter any global specimen ID for non-logged specimens for the purposes of matching assay results, however non-logged specimens will *not* be available to view on the **Specimen Management** page; they are only found in this assay run.

- 9. Optional: Set Dilution. Some tests allow you to indicate if a specimen was diluted before being tested. The dilution will default to 1 for each specimen. If the specimen was diluted you can change this value as needed. For example, a dilution of 1:5 should be entered as dilution= 5. A dilution of 1:20 should be entered as dilution= 20.
  - Note: LDMS will adjust the final result according to the dilution and the adjusted final result will be reflected on any test result reports. This does not apply to the PK test.
- **10.** Click **Upload File**, and then select your assay results file.

LDMS will attempt to match results in your file to specimens added to the run based on global specimen ID, other specimen ID, or ID1.

- Note: If any results were matched incorrectly, you can drag and drop them to the correct specimen, or drag and drop them into the **Unmatched Results from File** section to remove them.
- (If necessary) Match unmatched results to specimens.Click and drag unmatched results in the Unmatched Results from file section to appropriate specimens.
- **12.** (If necessary) To manual change a result, select the result, and then click **Set Manual Result**.
- **13.** Do one of the following:

- To save the run and change the status to complete, click Complete.
- To save the run and change the status to *pending*, click **Save**.

#### **Adding Cryopreservation Test Results**

One of the test results is the Cryopreservation test. This feature allows the IQA lab to assign confirmatory post-thaw results to cryopreserved specimens and is only available at the IQA lab.

- 1. Under the **Test Results** dropdown, click **Test Results**.
- **2.** From the action menu, select **Add Pending Results** The **Select Test** window will open.
- From the Test Name dropdown, select Cryopreservation and click Continue.
- **4.** On the **General** tab, define information about the test.
- **5.** On the **Results** tab, add specimens that are already in LDMS by using the **Add Specimens** button. The user can also add specimens from a file using the **Upload File** button.
- **6.** To save the test and complete it later, click **Save**.
- **7.** To complete the test, click **Complete**.

### **Assigning Results to Blinded Controls (Pellets)**

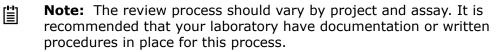
- Note: The steps below apply only to the TaqMan Qual assay.
- 1. On the **Test Results** page, select **Add Pending Results** from the **Add New...** drop down.
- 2. Select the COBAS TagMan HIV-1 Qual from the Test Name drop down.
- 3. Choose either the 24 or 48 radio button for the Number of Items.
- 4. Click Continue.
- 5. In the **Results** tab, choose pellets from the **Select Pellet** drop down.
- 6. Fill out the remaining field and click Complete.
- Note: Pellets can only be consumed once and will no longer be available once used.

### Reviewing a Run

Once a run has the status of completed, it must be reviewed for quality control purposes.

#### **Background**

Runs should typically be reviewed by someone other than the Data Transfer Tech who entered the results. The reviewer can check the calculated results, pass-fail status of specimens and the run, and apply censors as needed.



#### **Steps**

- 1. From the LDMS menubar, click **Test Results**.
- **2.** Locate the completed run of interest.

Use **Test Results Filters** to help locate the run.

- **3.** Next to the run, click **Edit**.
- Review the results.

Consult any procedures or documentation applicable to your laboratory to complete this review.

**5.** (If applicable) Apply censor codes.

To apply a censor to a specimen, locate the specimen, and then in the **User Censor** column, select the censor.

To apply a censor to the entire run, on the **General** tab, in the **User Censor** box, select a censor.

For details about a censor, hover over it with the mouse pointer.

System censors are automatically applied and cannot be removed.

- 6. On the **General** tab, complete the **Review Date**, **Reviewer Initials**, and (optionally) **Reviewer Comments** boxes.
- 7. Click Review.

### **Test History**

Test results can be changed between completed and pending statuses. It is possible to view a snapshot of test results at the time the status changed.

To view the test result history, on the **Test Result** page, locate the run. Next to **Edit** for that run, click the arrow, and then click **View History**. Next to the even, click **View** to see the run at that point in time.

Only certain events are shown in the history. For example, making changes to a run while its status is pending will not be shown in the history.

### **Deleting assay runs**

Assay runs that were erroneously created can be deleted.

#### **Background**

Deleted runs are not shown by default on the **Test Results** page. To display them, in the **Test Results Filters**, set the **Status** filter to Deleted.

#### **Steps**

- 1. On the **Test Results** page, locate the run to be deleted.
- 2. Click the arrow next to the run's **Edit** button, and then click **Delete**.
- **3.** Verify that the run you are about to delete is the correct run.
- Complete the History Date, History Tech Initials, and History Comments boxes.

The comments should indicate why the run is being deleted.

#### 5. Click **Delete**.

#### Restoring deleted runs

If an assay run is deleted in error, it can be changed back to pending without data loss.

#### **Steps**

- 1. On the **Test Results** page, in the **Test Results Filters**, set the **Status** filter to Deleted.
- **2.** Locate the run to be restored.
- 3. Click the arrow next to the run's **Edit** button, and then click **Reset to Pending**.
- 4. Complete the **History Date**, **History Tech Initials**, and **History Comments** boxes.
- 5. Click Reset.

#### **Test result reports**

Several tests can be run for assay result runs.

All reports are run by clicking the arrow next to the **Edit** button next to the run, and then clicking the desired reports.

| Report                      | Purpose  |  |
|-----------------------------|--|--|
| Completed Test Run Report   | Shows details of the specimens on a run, including plat positions, result filename, results for each specimen, and other result details.                       |  |
| Participant Report          | Provides detailed results broken down by participant and specimen (one per page)   |  |
| Pending Test Results Report | Shows details about specimens on the run and where they are located on testing plates (useful for setting up equipment to match a run already set up in LDMS). |  |

# **Entering PK Test Results**

Users can enter PK test results by navigating to the **Test Results** tab, clicking the **Test Results** module, selecting a test, and then clicking **Edit**.

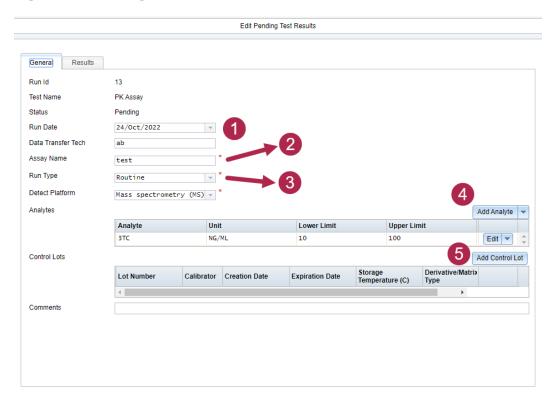


Figure 58: Editing Test Results - General Tab

On the General Tab, the user must define some required information about the PK Test run, including (1) a Run Date, (2) Assay Name, (3) Run Type, and (4) Analytes. (5) The user may optionally add a Control Lot to a run.

Complete Save Close

**Assay Name:** The Assay Name is a free entry field that the user can define. This name will be used to group assays together on certain reports and in the PK Control Charting Module.

**Run Type:** Certain Run Types are used for different scenarios. For example, Routine and Routine/Proficiency are used to report network results whereas Calibration and Stability may be used when validating a new method.

**Add Analyte:** When adding an analyte the user may choose to add these individually for each run or they can select an Analyte Setup that was previously defined in the PK Templates module.

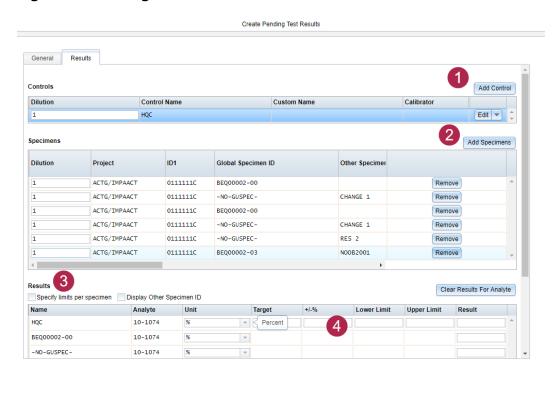


**Note:** The analyte setup can be modified for each run and for each item on the run. For example, a PK Template may have defined the analyte range as 10-1,000 but a special run today may need the range to be 100-100,000. The user can modify the range for any run as needed. Individual specimens and controls may also need range adjustments. For example a specimen analyte range may use ng/mL

but the controls may use a range in ng/sample. The LDMS will allow the user to change the limits for each individual item on the Results tab.

**Add Control Lot:** This will allow a user to add a Control Lot to this run. The user can either add a predefined set of controls created in the PK Control Lots module or, if a preset lot of controls have not previously been defined in the PK Control Lots module, the user can create it for this individual run. If the user chooses to not add or create the entire lot of controls here, they can add individual controls on the Results tab.

Figure 59: Editing Test Results - Results Tab



On the Results Tab the user can (1) add individual controls, (2) add specimens, (3) change the result grid to allow editing of the limits for each individual item, (4) review, define, or edit the limits, target, or range of each control and specimen

Complete Save Close

**Add Control:** The user can click Add Control to add additional controls to this run. Even if the user added a Control Lot on the general tab they can use this feature to include additional controls now.

**Add Specimens:** The user can choose to add specimens onto this run.

Note: Dilution will default to 1 for all specimens added to a PK Run. If the specimen was tested with a dilution the user can choose to change

the value here. Any result that has a dilution modified from 1 will be assigned a U censor.

**Specify Limits Per Specimen:** If any specimen on the run needs to use a limit range other than what was defined at the analyte level this option can be used to change the limit for each individual specimen.

**Setting Limits in the Result Grid:** The user can set the Target and +/- for each control or they can set a specific Lower and Upper limit. If the user has checked to Specify Limits per Specimen they can also set the lower and upper limit for each specimen.



**Note:** If a specimen was diluted the LDMS will account for the dilution when comparing the input result to the limits. The user should not change the standard limits for a specimen to account for dilution as the LDMS will do this automatically.



**Important:** If a result was not obtained for a specific specimen, leave the result field for that row Blank/Null. LDMS will allow the user to apply a User Censor to this result when Reviewing the run. Leaving a result Blank/Null may be necessary in scenarios such as a template including an analyte that was not tested for, a sample on the run having insufficient volume, or an issue in interpreting the final result.

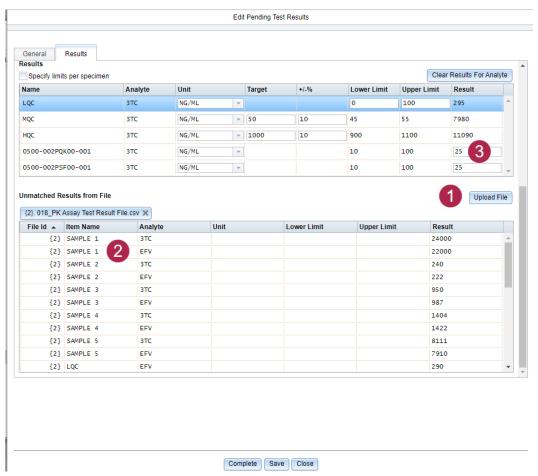


Figure 60: Editing Test Results - Results Tab (Continued)

After the run setup is complete and results are available, the user can set these results on the Results tab by (1) uploading a file, (2) auto matching or drag and drop matching file results to the results grid, or (3) by skipping result file upload and manually setting results.

**Upload File:** An example PK assay test result file is shown below.

Figure 61: Example PK Assay Test Result File

| 4  | A         | В       | С     | D |
|----|-----------|---------|-------|---|
| 1  | ID (1)    | зтс (4) | EFV   |   |
| 2  | SAMPLE 1  | 24000   | 22000 |   |
| 3  | SAMPLE 2  | 240     | 222   |   |
| 4  | SAMPLE 3  | 950     | 987   |   |
| 5  | SAMPLE 4  | 1404    | 1422  |   |
| 6  | SAMPLE 5  | 8111    | 7910  |   |
| 7  | LQC 2     | 295     | 290   |   |
| 8  | LQC 2     | 310     | 315   |   |
| 9  | MQC       | 7980    | 7990  |   |
| 10 | MQC 2     | 8025    | 8055  |   |
| 11 | HQC       | 11090   | 11080 |   |
| 12 | HQC 2     | 12050   | 12060 |   |
| 13 | SAMPLE 6  | 24000   | 22000 |   |
| 14 | SAMPLE 7  | 3 240   | 222   |   |
| 15 | SAMPLE 8  | 950     | 987   |   |
| 16 | SAMPLE 9  | 1404    | 1422  |   |
| 17 | SAMPLE 10 | 8111    | 7910  |   |
| 18 |           |         |       |   |
| 10 |           |         |       |   |

When uploading a result file the LDMS expects (1) a file that has a defined ID. The ID in the file will auto match to LDMS based on (2) the Control Name, (3) Global Specimen ID, Other specimen ID, or ID1 value. The LDMS also expects a result column named (4) as the Specific analyte being tested for.

**Unmatched Results From File:** Any result from the file that could not be automatically matched to an item on the run in LDMS will be available in this grid to drag-and-drop to match.

**Manually Setting Results:** If a file is not used or if the LDMS contains more results than are found in the result file, the user can manually enter results to LDMS by typing directly into the field.



**Note:** Results from diluted specimens must be entered to LDMS precalculated to account for dilution. LDMS will not change the input result to account for dilution. LDMS will adjust the limits based on dilution for purposes of calculating the A and B censors but LDMS expects the user to input the final result pre-calculated to account for the dilution.

### **Completing PK Test Results**

After all required information has been defined for a run the user may click **Complete**. When clicking complete, the LDMS will calculate system censors such as A- above upper limit, B- below lower limit, and U- specimen diluted. The LDMS will also verify any controls on the run fall within the expected

range and may set a pass/fail censor for the run depending on the success of any defined controls.

### Reviewing a PK Test Run

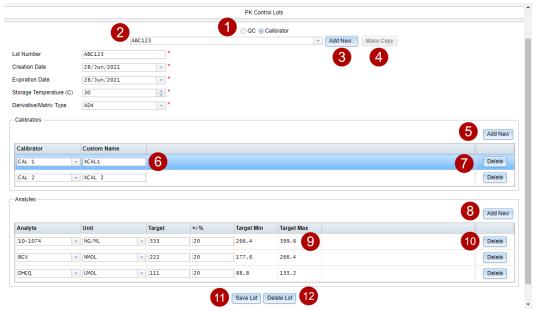
When reporting PK Test results for certain network specimens, the networks expect the run to pass a Review. After completing the run, the application allows for the run to be reviewed from the main test results grid and set information such as Reviewer Initials, Reviewed Date, Review Comments, and define user censors.

#### **PK Control Lots**

The PK Control Lot Module allows the user to pre-define the details of a control or calibrator lot in advance of using them on a run. Once a lot is added in the PK control lots module, it will be available in the Test Results module to quickly and easily place on any PK Assay runs for which it is used. When initially added to a specific run in the Test Result module, the lot will default to the details defined here.

#### Figure 62: PK Control Lots

(1) QC and Calibrator radio buttons, (2) choose existing PK lot, (3) add new PK lot, (4) make copy of PK lot, (5) add new calibrator/control, (6) list of calibrators/controls, (7) delete calibrator/control, (8) add new analyte, (9) list of analytes, (10) delete analyte, (11) save lot, (12) delete lot



### Adding a New Lot

- 1. Select QC or Calibrator.
- 2. Click Add New.
- **3.** Enter the following information:

- 3.1. Lot Number
- 3.2. Creation Date
- 3.3. Expiration Date
- **3.4.** Storage Temperature
- 3.5. Derivative/Matrix Type
- **4.** To add a control/calibrator, do the following:
  - Note: If the user selected QC in the first step, they will have the ability to add a control. If the user selected Calibrator in the first step, they will have the ability to add a calibrator.
  - 4.1. Click Add New in the Controls/Calibrator section.
  - **4.2.** Select a control/calibrator from the dropdown.
  - **4.3.** Enter a custom name for the control/calibrator.
    - Note: When matching results from an uploaded file, the LDMS will first try to match using this Custom Name.
  - **4.4.** Repeat step 4 as needed for additional controls/calibrators.
  - 4.5. To delete a control/calibrator, click the **Delete** button next to it.
- 5. To add an analyte, do the following:
  - **5.1.** Click **Add New** in the **Analytes** section.
  - **5.2.** Select an analyte from the dropdown.
  - **5.3.** Enter a unit.
  - **5.4.** Enter a target.
  - **5.5.** Enter a **+/-%**.
    - Note: LDMS will calculate the **Target Min** and **Target Max**. If necessary, these values can be modified for clarity during run set up when using this lot.
  - **5.6.** Repeat step 5 as needed for additional analytes.
  - **5.7.** To delete an analyte, click **Delete** next to the analyte.
- 6. Click Save Lot.

## **Editing or Deleting a Lot**

To Edit an existing lot, do the following:

- 1. Select QC or Calibrator
- **2.** Select the lot from the dropdown.
- 3. Optional: a lot can be copied by clicking Make Copy.
- **4.** Edit the lot information, controls/calibrators, and analytes as needed.
- 5. Click Save Lot.

To Delete a lot, select it and click **Delete Lot**.

## **PK Control Charting**

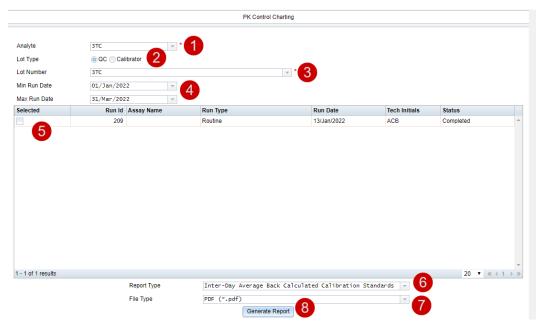
The PK Control Charting module allows the user to report on specific PK Control or Calibrator lots to review their details over time. Various reports

are made available for the user to run, including Inter-Day Average Back Calculated Calibration Standards, Stability, Accuracy and Precision, and more. Generation of these reports requires a user to first define PK Control or Calibrator lots in the PK Control Charting Module and to then also make use of these lots on PK Assay runs in the Test Results Module.



**Note:** In some scenarios the LDMS may be unable to generate PK Control Charting reports if the user chose to modify or manipulate certain lot details that result in mathematical queries and equations that cannot be properly executed.

Figure 63: PK Control Charting



(1) select Analyte, (2) QC and Calibrator radio buttons, (3) Lot Number, (4) Min Run Date and Max Run Date, (5) Selected runs, (6) report type to generate, (7) report file type, (8) Generate Report

## **Generating a Report**

- 1. Enter the following information:
  - **1.1.** Select an analyte from the dropdown.
  - 1.2. Select Lot Type (QC or Calibrator).
  - **1.3.** Select **Lot Number** from the dropdown.
  - 1.4. Add Min Run Date and Max Run Date.
- 2. Tick boxes next to runs to include them in the report.
- **3.** Choose the type of report to generate from the **Report Type** dropdown.
- **4.** Select a file type for the report.
- 5. Click Generate Report.

## **PK Templates**

PK templates are predefined, specific analyte scenarios that can be used to help set up a Pending Test Results. Templates can be created for PK analyses that happen frequently for a specific set of analytes. Templates can be created and modified by users. User-created templates are only available at the laboratory where they were created; they are not sharable with other laboratories.

Figure 64: PK Templates



(1) Currently selected template, (2) add a new template, (3) template name, (4) add new analyte, (5) delete selected analyte, (6) save template, (7) delete template

### **Adding a New Template**

- 1. Name the template in the Name field.
- 2. Add analytes:
  - 2.1. Click Add Analyte.
  - 2.2. Select Analyte and Unit.
  - **2.3. Optional**: Add **Upper Limit** and **Lower Limit**.
  - **2.4.** To delete an analyte, click the **Delete** button.
- 3. Click Save Template.

### **Updating an Existing Template**

PK Templates can be modified at any time, even if they have previously been used on a live test run.



**Note:** PK Templates can be selected for use on a live test run and be further modified to meet the needs of that specific run. For example, additional analytes can be added or removed, the limit units can be changed, or the specified upper or lower limit can be updated as needed for the live run.

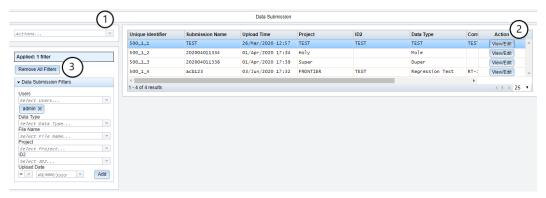
- **1.** Select the template from the drop down menu.
- **2.** Update the name or analyte information as needed.
- 3. Click Save Template.
- **4.** To delete the template that is currently open, click **Delete Template**.

# **Data Submission**

The Data Submission Module is used to upload data files to Frontier Science within web LDMS without having to log into the Portal or other Frontier Science systems. This is accessed through the **Data Submission** menu.

Note: Users must contact Frontier Science if access to this module is needed.

Figure 65: The Data Submission Page

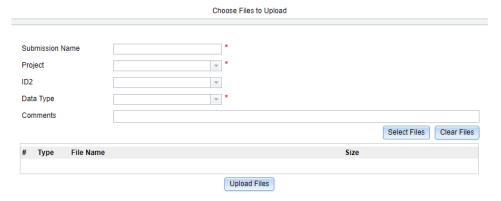


(1) The dropdown list used to create new data submissions. (2) The View/Edit button used to edit data submissions. (3) Filters to narrow results.

## **Adding Submissions**

To add a data submission, follow the steps below.

- From the menu bar, click **Data Submission**, and then click **Data Submission**.
- 2. In the **Actions** dropdown menu on the left side of the screen, click **Upload Files**. The **Choose Files to Upload** window will appear.

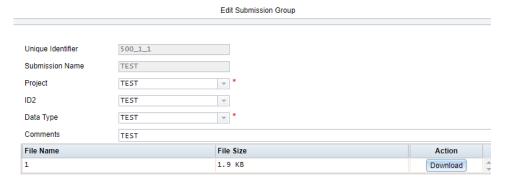


- **3.** Add the submission name to the **Submission Name** field.
- **4.** Select the Project, ID2, and Data Type from the dropdown menus.
- **5.** Add any comments as needed.
- **6.** Below the **Comments** text box, click the **Select Files** button.
- **7.** Select the necessary files.
- **8.** To remove selected files, click **Remove Files**.
- **9.** To complete the submission, click **Upload Files**.

## **Editing Submissions**

To edit a data submission, follow the steps below.

- 1. From the menu bar, click **Data Submission**, and then click **Data Submission**.
- 2. To make changes to submissions, click **View/Edit** next to the submission in question. The **Edit Submission Group** window will appear.





- **3.** Make any changes to the project, ID2, Data Type, or Comments as needed by using the appropriate dropdown menus.
- **4.** Files in the submission can also be downloaded individually by clicking the **Download** button next to the file in question at the bottom of the window.
- 5. Click Save.

## **Data Submission Filters**

Filters can be applied to narrow results in the **Data Submission** menu.

#### **Background**

The following filters are available for the **Data Submission** module:

- Users
- Data Type
- File Name
- Project
- ID2 Upload Date

Select any filters as needed and click **Add**.

# **Printers**

This section will describe how to setup various printers that are known to work with LDMS.

## Setting up the Brady IP300 printer

These instructions will help ensure that your printer is set up to work correctly with LDMS.

#### **Prerequisites**

 Install the driver for your printer. For assistance obtaining and installing the driver, contact your laboratory's local IT department or the manufacturer's customer support.

#### **Background**

This printer is known to work with the following label and stock settings in LDMS:

label stock THT-183-461

LDMS label Barcode label 7

- Open the printer settings from the Windows Start menu.
   These are typically found in Start > Devices and Printers.
- 2. Right-click the **IP300** icon, and then click **Printing preferences**.
- 3. Click Advanced.

- 4. Next to Paper Size, click Properties.
- **5.** In the Paper Size box, select the appropriate paper size.
- On the control panel in front of the printer, change the printer head to +10mm for the X axis and -3.0mm for the Y axis

These head adjustments have been tested with the THT-183-461 label stock. Other label stocks may require different printer head adjustments.

## Setting up the Brady® MVP 300 printer

These instructions will help ensure that your printer is set up to work correctly with LDMS.

#### Steps

- Open the printer settings from the Windows Start menu.
   These are typically found in Start > Devices and Printers.
- **2.** Right click the icon for your printer, and click **Printing preferences**.
- **3.** Set the stock size.
  - **3.1.** Expand the **Paper/Output** settings
  - **3.2.** In the **Paper Size** box, select **User Defined**.
  - 3.3. Click Customize.
  - **3.4.** In the **Width** and **Height** boxes, entered the dimensions of your label stock.
  - **3.5.** In the **Unit of Measure** box, select **Inches**.
  - **3.6.** Click **OK**.
- **4.** In the **Devices and Printers** window, right click the printer icon, and click **Properties**.
- **5.** On the **Printer** tab, change the following settings:
  - **5.1.** In the **Device Options** > **Head Settings** > **Print Darkness** section, change the **Print Darkness** setting to 22.
  - **5.2.** In the **Speed Settings**, enter the following settings:

Print speed 2.00 in/sec
Slew speed 2.00 in/sec
Back feed 2.00 in/sec
speed

**6.** On the **Ports** tab, click **Configure Port** and enter the following settings:

**Bits per** 115200

second

**Data bits** 8

Parity None

#### Stop bits 1

Flow control Xon/Xoff

- **7.** Configure the printer hardware.
  - **7.1.** Open the printer and remove the protective plate that is covering the ribbon sensor.
  - **7.2.** Slide the transmissive sensor protector to the outside of the adjustable transmissive sensor.
  - **7.3.** Place the ribbon on the ribbon supply spindle.
    - Note: The shiny side of the ribbon must be facing up.
  - **7.4.** Feed the ribbon through the ribbon sensor and around the print head assembly.
  - **7.5.** Wrap the ribbon around the ribbon take-up spindle until it is tight.
    - **Tip:** Attach a piece of tape to the end of the ribbon to help keep it tight during the loading process.
  - **7.6.** Place the labels on the media supply spindle.
    - The printable side of the label must be face-up.
  - **7.7.** Place the labels through the media guide and align the notch between labels with the tear-off plate of the printer.
  - **7.8.** Adjust the ribbon sensor so that the notch of the label will cross the ribbon sensor.
    - The ribbon sensor is the red light under the label.
  - **7.9.** Close the print head and ensure that the ribbon is aligned with the left side of the label.
    - This adjustment may require you to slide the ribbon across the print head assembly.
  - **7.10.**Close the printer.
- **8.** On the control pad on front of the printer, do the following:
  - **8.1.** Press the **Setup/Exit** button.
  - **8.2.** Press the + button until the printer displays **Manual Calibrate**.
  - **8.3.** Press the **Select** button.
  - **8.4.** Press the + button.
    - The printer will re-calibrate its label settings.
  - **8.5.** Press the **Setup/Exit** button, and then press the + button to save the changes.

## Setting up the LabXpert printer

These instructions will help ensure that your printer is set up to work correctly with LDMS.

#### Steps

**1.** Open the printer settings from the Windows **Start** menu.

These are typically found in **Start** > **Devices and Printers**.

- **2.** Right click the icon for your printer, and click **Printing preferences**.
- **3.** Expand the **Paper/Output** settings, and click **Properties** text to Paper Size.
- **4.** In the Paper Size box, select the appropriate paper size.
- **5.** In the **Devices and Printers** window, right click the LabXpert icon, and click **Properties**.
- **6.** On the **Ports** tab, enter the following settings:

**Bits per** 115200

second

Data bits 8

Parity None

Stop bits 1

Flow control Xon/Xoff

- **7.** Configure the printer to accept input from your LDMS computer.
  - **Important:** This step must be completed every time the printer is turned on.
  - **7.1.** On the printer, press the Menu button.
  - **7.2.** Press the Next button three times.
  - **7.3.** Press the PC button.

#### After you are finished

After the printer is set up, you should print a set of test labels to ensure that it is working correctly.

## Setting up the Zebra® GX 430t printer

These instructions will help ensure that your printer is set up to work correctly with LDMS.

### **Prerequisites**

- Install the driver for your printer. For assistance obtaining and installing the driver, contact your laboratory's local IT department or the manufacturer's customer support.
- Load the printer with label stock and ribbon according to the printer's manual.

- Open the printer settings from the Windows Start menu.
   These are typically found in Start > Devices and Printers.
- **2.** Right click the icon for your printer, and click **Printing preferences**.

- **3.** On the **Options** tab, change the **Speed** to 2.
- 4. Change the **Print Darkness** to about 27.

This setting should be adjusted up or down, depending on whether labels are too light or too dark.

- **5.** In Paper format, select inches.
- **6.** In **Paper Size**, set the **Width** to 4.00 and the **Height** to 1.75.
- 7. On the **Advanced Setup** tab, click **Calibrate**.
- 8. On the **Advanced Setup** tab, verify that the **Tracking Mode** is set to Web Sensing.

#### After you are finished

After the printer is set up, you should print a set of test labels to ensure that it is working correctly.

## **Setting up the Brady BBP11-34L printer**

These instructions will help ensure that your printer is set up to work correctly with LDMS.

### **Background**

This printer is known to work with the following label and stock settings in LDMS:

label stock THT-183-461-0.5-SC LDMS label Barcode Label 9

#### Steps

- Open the printer settings from the Windows Start menu.
   These are typically found in Start > Devices and Printers.
- **2.** Right click the icon for your printer, and click **Printing preferences**.
- 3. On the Page Setup tab, under Stock click New.
- **4.** Enter the dimensions  $1.75" \times 1.00"$ .

These are the dimensions for the THT-183-461-0.5-SC label stock. If you are using a different stock, enter the appropriate dimensions.

- **5.** Click **Advanced Options**, and verify that the horizontal and vertical offset options are set to 0".
- **6.** On the **Stock** tab, enter the following into the **Media settings**:

Method Thermal transfer

Type Labels with marks

Mark height 0.08 in Mark offset 0.00 in

**7.** On the **Options** tab, enter the following into the **Printer options**:

**Print speed** 2.00 in/sec

Darkness 10

8. Click OK.

#### After you are finished

After the printer is set up, you should print a set of test labels to ensure that it is working correctly.

## Setting up the Brady® BBP33 printer

These instructions will help ensure that your printer is set up to work correctly with LDMS.

#### **Prerequisites**

• Install the driver for your printer. For assistance obtaining and installing the driver, contact your laboratory's local IT department or the manufacturer's customer support.

#### **Background**

This printer is known to work with the following label and stock settings in LDMS:

label stock B33-179-482 (1"x1")

**LDMS label** Barcodelabel19

#### Steps

- Open the printer settings from the Windows Start menu.
   These are typically found in Start > Devices and Printers.
- **2.** Right click the icon for your printer, and click **Printing preferences**.
- 3. Click Advanced.
- 4. Next to Paper Size, click Properties.
- **5.** In the Paper Size box, select the appropriate paper size.
- **6.** In LDMS, set the printer as your label printer.

#### After you are finished

After the printer is set up, you should print a set of test labels to ensure that it is working correctly.

## Setting up the Brady BMP53 printer

These instructions will help ensure that your printer is set up to work correctly with LDMS.

#### **Prerequisites**

 Install the driver for your printer. For assistance obtaining and installing the driver, contact your laboratory's local IT department or the manufacturer's customer support.

#### **Background**

This printer is known to work with the following label and stock settings in LDMS:

Cartridge M-156-492

**LDMS label** Barcode Label 16

#### **Steps**

- Open the printer settings from the Windows Start menu.
   These are typically found in Start > Devices and Printers.
- **2.** Right click the BMP51(53) icon, and click **Printing preferences**.
- 3. Click Advanced.
- 4. Next to Paper Size, click Properties.

#### After you are finished

After the printer is set up, you should print a set of test labels to ensure that it is working correctly.

## Setting up the Brady BP-PR 300 printer

These instructions will help ensure that your printer is set up to work correctly with LDMS.

#### **Prerequisites**

• Install the driver for your printer. For assistance obtaining and installing the driver, contact your laboratory's local IT department or the manufacturer's customer support.

- Open the printer settings from the Windows Start menu.
   These are typically found in Start > Devices and Printers.
- **2.** Right click the Brady PR 300 PLUS icon, and click **Properties**.

3. In the **Size** section, change the **Width** and **Height** to the size of your label stock.

### After you are finished

After the printer is set up, you should print a set of test labels to ensure that it is working correctly.

## **Setting up the Zebra ZD620 Printer**

These instructions will help ensure that your printer is set up to work correctly with LDMS.

#### **Prerequisites**

- Install the driver for your printer. For assistance obtaining and installing the driver, contact your laboratory's local IT department or the manufacturer's customer support.
- Load the printer with label stock and ribbon according to the printer's manual.

#### Steps

- Open the printer settings from the Windows Start menu.
   These are typically found in Start > Devices and Printers.
- **2.** Right click the icon for your printer and click **Printing Preferences**.
- **3.** On the **Print Options** tab, change the **Speed** to 4. This setting should be adjusted accordingly.
- 4. Change the **Print Darkness** to about 20. This setting should be adjusted up or down, depending on whether labels are too light or too dark.
- 5. In the **Page Setup**, set the **Width** and **Height** according to labels used. Ex.: Width: 2.75" Height: 1"

### After you are finished

After the printer is set up, you should print a set of test labels to ensure that it is working correctly.

## Setting up other printers

Label printers that do not have specific instructions provided in this manual may still work with LDMS, however they will not have been tested by LDMS User Support.

### Steps (completed in any order)

- Contact LDMS User Support for assistance.
- Consult the most current list of supported printers.

• If you are using a printer not listed in this manual, let LDMS User Support know so that it can be added.

# **Preset projects**

Preset projects are projects that are built into LDMS that have special rules and requirements.

Preset projects and their requirements cannot be modified or removed by users. These projects and designed based on the needs of a specific project working with Frontier Science. An example of a preset project is ACTG/IMPAACT. Specimens created until this project have specific requirements, which are defined by ACTG and IMPAACT. These requirements are implemented in LDMS to accommodate the needs of the project.

The rules for preset projects fall into two categories:

- Validation rules that can be enforced by LDMS, such as requiring that ID1 be formatted a certain way
- Requirements that cannot be enforced by LDMS, such as requiring specimens to be entered within a certain period of time after they are received by your laboratory.



**Important:** The information for individual projects provided here are for convenience only. Always refer to guidance provided by the project for the most up-to-date instructions.

### **ACTIV**

The ACTIV (Accelerating COVID-19 Therapeutic Interventions and Vaccines) preset project must meet these requirements.

| Field         | Required? | Note  |
|---------------|-----------|---|
| ID1 (PID)     | Required  | Must be 9 numeric digits                    |
| ID2 (Study)   | Required  | "ACTIV2" in ID2 field; not editable         |
| ID3 (ID3)     | Optional  | Warning generated if other spec ID is blank |
| Visit         | Required  |   |
| Clinic        | Optional  |   |
| Specimen Time | Required  |   |
| Received Time | Required  |   |

## **ACTG/IMPAACT**

The ACTG/IMPAACT preset project has these requirements.

### **Data entry requirements**

| Field          | Required? | Note  |
|----------------|-----------|---|
| ID1 (PID)      | Required* | 8 characters long, 7 numbers and one letter<br>between A and L; if the PID does not have 7<br>numbers, add leading zeros to the front of it.  |
|                |           | *This requirement can be disabled by Frontier Science for certain studies.  |
| ID2 (Protocol) | Required  | Select from the pre-defined list of protocols.  |
| ID3            | Optional  | LDMS will remember previously entered ID3s and automatically populate ID3 when the same project-PID-protocol combination is entered.  |
| Visit          | Required  | Select from pre-defined list of visit units.  |
| OPID           | Available |   |
| Clinic         | Required  | LDMS will automatically populate the clinic using one previously entered for the same PID, protocol, and SID combination.   |
| Specimen Time  | Depends   | If you do not have a specimen time recorded on your CRF, contact the clinic that collected the specimen from the participant. This field is required for specimens with a specimen date of 01 January 2004 or later; it is optional for specimens prior to that date. This requirement can be overridden for specimens after this date, if necessary. |
| Received Time  | Optional  |   |

#### **Notes**

- ACTG/IMPAACT uses the standard LDMS Shipping File format to ship between ACTG and IMPAACT laboratories using LDMS.
- ACTG/IMPAACT requires specimens to be labeled with a 2D barcode and label from LDMS; LDMS comes with label formats for ACTG setup and ready to use.
- ACTG and IMPAACT are listed as a combined project throughout LDMS (ACTG/IMPAACT).
- Preloads are permitted for the ACTG/IMPAACT project, but users are not permitted to create their own. All ACTG preloads are created by Frontier Science in collaboration with network leadership.

## **AERAS**

The AERAS preset project has these requirements.

| Field         | Required? | Note  |
|---------------|-----------|---|
| ID1 (PID)     | Required  | 11-digits of any combination of letters and numbers.  |
| ID2 (Study)   | Required  | Selected from pre-populated list. Once selected, the LDMS will check that the PID is associated with the selected protocol. |
| ID3           | Disabled  | This field is not used by Aeras   |
| Visit         | Required  | Select from pre-defined list of visit units.  |
| OPID          | Disabled  | This field is not used by AERAS   |
| Clinic        | Required  | AERAS clinics are in the format ARSXX, where XX are the 5th and 6th digits from the PID                                     |
| Specimen Time | Required  |   |
| Received Time | Optional  |   |

### Notes

Quick Add templates can be used for Aeras.

## **AIEDRP**

The AIEDRP preset project has these requirements.

## **Data entry requirements**

| Field          | Required? | Note  |
|----------------|-----------|---|
| ID1 (PID)      | Required  | Must either pass the ACTG/IMPAACT or HPTN ID1 validation check, depending on the protocol |
| ID2 (Protocol) | Required  | Will determine what ID1 format should be used   |
| ID3 (SID)      | Required  | Must be NOSID or pass the ID1 validation  |
| Visit          | Required  |   |
| OPID           | Optional  |   |
| Clinic         | Optional  |   |
| Draw Time      | Optional  |   |
| Received Time  | Optional  |   |

### **Protocol and ID1 check used**

| Protocol | ID1 validation used |
|----------|---------------------|
| 103005   | ACTG/IMPAACT        |
| 103006   | ACTG/IMPAACT        |
| 103007   | ACTG/IMPAACT        |

| Protocol | ID1 validation used |
|----------|---------------------|
| 103008   | ACTG/IMPAACT        |
| I03010   | Either              |
| I03011   | Either              |
| CORE01   | Either              |
| AIN501   | Either              |
| AIN502   | Either              |
| AIN503   | Either              |
| AIN504   | Either              |

## **AIS**

The AIS (Nigeria State Level ART Impact Survey) preset project has these requirements.

### **Data entry requirements**

| Field                              | Required? | Note  |
|------------------------------------|-----------|---|
| ID1 (PTID)                         | Required  | Must be length 9. First 4 characters are letters only and last 5 characters are numeric only (e.g., ABCD12345)  |
| ID2 (Country)                      | Required  | "Nigeria" is the only available option  |
| ID3 (HIVRT)                        | Required  | Pick list of specific options   |
| Collection Time                    | Required  |   |
| Received Date and<br>Received Time | Required  |   |
| Clinic                             | Required  | Must require the clinic be formatted like NG### where the first 2 characters are the letters NG and the next 3 characters are numbers. (e.g., NG000 or NG031) |

## **AMC**

The AMC preset project has these requirements.

| Field          | Required? | Note |
|----------------|-----------|------|
| ID1 (ID1)      | Required  |      |
| ID2 (PROTOCOL) | Optional  |      |
| ID3 (ID3)      | Optional  |      |
| Visit          | Optional  |      |

| Field         | Required? | Note |
|---------------|-----------|------|
| OPID          | Optional  |      |
| Clinic        | Optional  |      |
| Draw Time     | Optional  |      |
| Received Time | Optional  |      |

## **ATN**

The ATN preset project has these requirements.

## **Data entry requirements**

| Field          | Required? | Note   |
|----------------|-----------|--|
| ID1 (PID)      | Required  | A 6-digit number between 10001 and 999999. If the PID is less than 6 digits, leading zeros will automatically be added. LDMS will check that the PID is valid.   |
| ID2 (Protocol) | Required  | Must be a 3 digit number between 004 and 999   |
| ID3 (SID)      | Required  | An 8-digit number between 40001 and 999999999. If the SID is less than 8-digits, leading zeros will automatically be added. 7-digit SIDs can still be imported by LDMS. If the same PID-protocol combination has been entered, LDMS will automatically default to the previously used SID. NOSID is a valid entry for ATN. |
| Visit          | Required  | Select from pre-defined list of visit units.   |
| OPID           | Available |  |
| Clinic         | Optional  |  |
| Specimen Time  | Required  |  |
| Received Time  | Optional  |  |

#### **Notes**

Quick Add templates can be used for ATN.

## **BHP**

The BHP preset project has these requirements.

| Field         | Required? | Note   |
|---------------|-----------|--|
| ID1 (PID)     | Required  | 8 characters long, 7 numbers and one letter between A and L; if the PID does not have 7 numbers, add leading zeros to the front of it. |
| ID2 (Study)   | Required  | The only available choice in the drop down menu is 'Tatelo'.   |
| ID3 (Step)    | Required  | Available choices in drop down menu are 0, 1, 2, or 3.   |
| Visit         | Required  | Select from pre-defined list of visit units.   |
| OPID          | Optional  |  |
| Clinic        | Required  | Any clinic may be set.   |
| Specimen Time | Required  |  |
| Received Date | Required  |  |
| Received Time | Required  |  |

### Notes

• User Quick Add templates are not permitted for BHP.

## **BM**

The BM preset project has these requirements.

### **Data entry requirements**

| Field         | Required? | Note                             |
|---------------|-----------|----------------------------------|
| ID1 (studyID) | Required  | Must be in the BM studyID format |
| ID2 (ID2)     | Optional  |                                  |
| ID3 (ID3)     | Optional  |                                  |
| Visit         | Optional  |                                  |
| OPID          | Optional  |                                  |
| Clinic        | Optional  |                                  |
| Draw Time     | Optional  |                                  |
| Received Time | Optional  |                                  |

#### **BM ID1 format**

BM-XXX-Y

**BM** Must be "BM" **XXX** Any 3 digits

Y 0 for mother; 1 to 9 for child

### **Botswana MOH**

The Botswana MOH preset project has these requirements.

### **Data entry requirements**

| Field       | Required? | Note  |  |
|-------------|-----------|---|--|
| ID1 (PID)   | Required  | Requires 2 letters followed by 8 numbers.         |  |
| ID2 (Study) | Required  | Default option is IBBS3                           |  |
| ID3 (HIVRT) | Required  | Select from a list of predefined HIV status value |  |
| Clinics     | Optional  |   |  |

#### **Notes**

• The application must allow FSTRF-defined preloads for this project.

#### **Botswana MOH PID format**

XXYYNNNNNN

XX District (alphabet, length 2)

**YY** KP type (numeric, length 2)

**NNNNN** Participant (numeric, length 6)

## **CEMALB**

The CEMALB preset project must meet these requirements.

| Field          | Required? | Note                 |
|----------------|-----------|----------------------|
| ID1 (PID)      | Required  | Must be 5 characters |
| ID2 (Protocol) | Required  |                      |
| ID3 (Visit)    | Optional  |                      |
| Visit          | Optional  |                      |
| OPID           | Optional  |                      |
| Clinic         | Optional  |                      |
| Draw Time      | Optional  |                      |
| Received Time  | Optional  |                      |

## **CHAVI**

The CHAVI preset project must meet these requirements.

## **Data entry requirements**

| Field          | Required? | Note   |  |
|----------------|-----------|--|--|
| ID1 (PID)      | Required  | Must pass HPTN ID1 validation and must be 9 characters in length   |  |
| ID2 (Protocol) | Required  | Validated using ID1, where ID2 is embedded into ID1 as either the 4th or 4th and 5th digits, depending on the protocol |  |
| ID3 (ID3)      | Optional  |  |  |
| Visit          | Required  |  |  |
| OPID           | Optional  | Must be empty until the protocol is 008A   |  |
| Clinic         | Optional  |  |  |
| Draw Time      | Optional  |  |  |
| Received Time  | Required  |  |  |

## **CIPRA-HT**

The CIPRA-HT preset project has these requirements.

## **Data entry requirements**

| Field          | Required? | Note   |
|----------------|-----------|--|
| ID1 (PID)      | Required  | Must pass HPTN ID1 validation                        |
| ID2 (Protocol) | Required  |  |
| ID3 (SID)      | Required  | Must pass the historical ACTG/IMPAACT ID3 validation |
| Visit          | Optional  |  |
| OPID           | Optional  |  |
| Clinic         | Required  |  |
| Draw Time      | Optional  |  |
| Received Time  | Optional  |  |

## **CIPRA-ZA**

The CIPRA-ZA project has these requirements.

| Field         | Required? | Note |
|---------------|-----------|------|
| ID1 (PID)     | Required  |      |
| ID2 (CP#)     | Optional  |      |
| ID3 (SID)     | Optional  |      |
| Visit         | Optional  |      |
| OPID          | Optional  |      |
| Clinic        | Optional  |      |
| Draw Time     | Optional  |      |
| Received Time | Optional  |      |

## **CIPRA**

The CIPRA preset project must meet these requirements.

### **Data entry requirements**

| Field          | Required? | Note   |  |
|----------------|-----------|--|--|
| ID1 (PID)      | Required  | Must be 9 characters in the CIPRA ID1 format |  |
| ID2 (Protocol) | Required  | In the format P[0-10]                        |  |
| ID3 (ID3)      | Optional  |  |  |
| Visit          | Required  |  |  |
| OPID           | Optional  |  |  |
| Clinic         | Optional  |  |  |
| Draw Time      | Optional  |  |  |
| Received Time  | Optional  |  |  |

#### **CIPRA ID1 format**

AABBCCCCD

**AA** 01 to 99 **BB** 01 to 99

**CCCC** 0001 to 9999

**D** 0 to 9

## **CONTROL**

The CONTROL preset project must meet these requirements.

| Field             | Required? | Note |
|-------------------|-----------|------|
| ID1 (CONTROLNAME) | Required  |      |
| ID2 (PROTOCOL)    | Optional  |      |
| ID3 (ID3)         | Optional  |      |
| Visit             | Optional  |      |
| OPID              | Optional  |      |
| Clinic            | Optional  |      |
| Draw Time         | Optional  |      |
| Received Time     | Optional  |      |

## **CoVPN**

The CoVPN (COVID-19 Vaccine and Prevention Network) preset project must meet these requirements.

### **Data entry requirements**

| Field         | Required? | Note   |
|---------------|-----------|--|
| ID1 (PATID)   | Required  | Study 3008 has a predefined ID1/PATID check. |
| ID2 (Study)   | Required  |  |
| ID3 (ID3)     | Optional  |  |
| Visit         | Required  |  |
| Clinic        | Required  |  |
| Specimen Time | Required  |  |
| Received Time | Required  |  |

#### **Notes**

- Information for study CoVPN 5001 should be logged under VTN study 5001.
- Labels are restricted to all supported VTN formats.
- CoVPN has a checkbox for **Collected Outside Protocol Requirements**.

## **CPCRA**

The CPCRA preset project has these requirements.

| Field     | Required? | Note |
|-----------|-----------|------|
| ID1 (PID) | Required  |      |

| Field         | Required? | Note |
|---------------|-----------|------|
| ID2 (Study)   | Optional  |      |
| ID3 (Bth/Sex) | Optional  |      |
| Visit         | Optional  |      |
| OPID          | Optional  |      |
| Clinic        | Optional  |      |
| Draw Time     | Optional  |      |
| Received Time | Optional  |      |

## **CPQA**

The CPQA preset project has these requirements.

### **Data entry requirements**

| Field         | Required? | Note   |
|---------------|-----------|--|
| ID1 (PanelID) | Required  | Must be five characters in length in the format [Round_#][Panel_alpha_ID][2_numbers], such as 22A00        |
| ID2 (RoundNo) | Required  | This is the same as the first 2 digits of the PaneIID. This field will automatically be populated by LDMS. |
| ID3 (ExpDate) | Optional  |  |
| Visit         | Optional  | Select from pre-defined list of visit units.   |
| OPID          | Optional  |  |
| Clinic        | Optional  |  |
| Specimen Time | Optional  |  |
| Received Time | Optional  |  |

#### **Notes**

- CPQA use the LDMS shipping file format
- There are CPQA barcode options available in LDMS.
- Quick Add templates can be used for CPQA, however users are not expected to use this feature.
- The *Pharmacology Proficiency Results* report can be generated from LDMS for Clinical Laboratory Improvement Amendments (CLIA). This is sometimes referred to as the "CLIA Report".

## **CP-CTNET**

The CP-CTNET (Cancer Prevention Clinical Trials Network) preset project must meet these requirements.

| Field       | Required? | Note |
|-------------|-----------|------|
| ID1 (PID)   | Required  |      |
| ID2 (Study) | Required  |      |
| ID3 (ID3)   | Optional  |      |

## **FACTS**

The FACTS preset project has these requirements.

### **Data entry requirements**

| Field         | Required? | Note  |
|---------------|-----------|---|
| ID1 (PID)     | Required  | A 7-digit number. LDMS will verify that the PID is valid. |
| ID2 (Study)   | Required  | Select from pre-populated list                            |
| ID3           | Disabled  | This field is not used by FACTS                           |
| Visit         | Required  | Select from pre-defined list of visit units.              |
| OPID          | Disabled  | This field is not used by FACTS                           |
| Clinic        | Disabled  | This field is not used by FACTS                           |
| Specimen Time | Required  |   |
| Received Time | Optional  |   |

### **Notes**

• Quick Add templates can be used for FACTS.

### HN

The HN preset project must meet these requirements.

| Field         | Required? | Note |
|---------------|-----------|------|
| ID1 (studyID) | Required  |      |
| ID2 (ID2)     | Optional  |      |
| ID3 (ID3)     | Optional  |      |
| Visit         | Optional  |      |
| OPID          | Optional  |      |
| Clinic        | Optional  |      |
| Draw Time     | Optional  |      |

| Field         | Required? | Note |
|---------------|-----------|------|
| Received Time | Optional  |      |

## **HPTN**

The HPTN preset project has these requirements.

### **Data entry requirements**

| Field          | Required? | Note  |
|----------------|-----------|---|
| ID1 (PID)      | Required  | LDMS performs a checks to make sure the PID is valid. Based on the protocol, LDMS will determine whether a 9-digit or 10-digit PID must be used.  |
| ID2 (Protocol) | Required  | Special PID checks are enforced if the protocol is 043.0. For 043.0, the visit will also be set to 2.0 VST, and the user will not be able to modify it. This field allows dashes (e.g., 084-01)   |
| ID3            | Optional  | Special ID3 checks are enforced if the protocol is 043.0. For 043.0, this field allows up to 12 characters, and should be scanned from the blood ID number.   |
| Visit          | Required  | Defaults to the visit unit of VST. Interim, unscheduled visits should be entered by incrementing the visit value (e.g. 1.10 for the first unscheduled visit, 1.20 for the second, etc.). If the protocol is 043.0, the visit will also be set to 2.0 VST, and the user will not be able to modify it. |
| OPID           | Disabled  |   |
| Clinic         | Optional  | Any clinic may be set.  |
| Specimen Time  | Required  |   |
| Received Time  | Optional  |   |

### **Notes**

- There are several HPTN label formats available.
- Primary and aliquot volumes with default to ML.
- Quick Add templates are not permitted for HPTN.
- Old records may have a clinic set retroactively if not set at the time of entry

## **IDCRC**

The IDCRC (Infectious Diseases Clinical Research Consortium) preset project must meet these requirements.

| Field         | Required? | Note  |
|---------------|-----------|---|
| ID1 (PTID)    | Required  | LDMS checks that the PID is valid. Must<br>be 9 or 10 characters. A PID cannot be<br>changed without approval from VTN Laboratory<br>Leadership.  |
| ID2 (Study)   | Required  | Drop down menu  |
| ID3 (ID3)     | Optional  |   |
| Visit         | Required  |   |
| Clinic        | Required  | The clinic is automatically populated based on the PID; the same clinical location can be associated with multiple clinic IDs (e.g. for Phase I/II or Phase II/III). IDCRC clinics start with the letter V (such as V101 instead of 101). LDMS will verify that an IDCRC clinic was selected. |
| Specimen Time | Required  |   |
| Received Time | Required  |   |

## **IPREX**

The IPREX preset project has these requirements.

## **Data entry requirements**

| Field          | Required? | Note   |
|----------------|-----------|--|
| ID1 (PID)      | Required  | Must by 5 to 7 characters long and a valid IPREX PID; NOPID is a valid PID, unless the SID is NOSID.                                       |
| ID2 (Protocol) | Required  |  |
| ID3 (SID)      | Required  | Must be 11 characters long, with the first 2 digits being the clinic and the last five being the screening number in the range 80001-99999 |
| Visit          | Required  |  |
| OPID           | Optional  |  |
| Clinic         | Required  | Must be 3 characters long in the format I[86 to 97] (such as 192)  |
| Draw Time      | Optional  |  |
| Received Time  | Optional  |  |

#### **Notes**

- IPREX using the LDMS Shipping File format.
- When entering sero-conversion visits, use the visit unit sc. The visit value should be the same as the originally scheduled visit.

• When entering sero-positive visits, use the visit unit SP. The visit value should be the number of weeks after the seroconversion visit.

## **IQA**

The IQA preset project has these requirements.

### **Data entry requirements**

| Field         | Required? | Note |
|---------------|-----------|------|
| ID1 (PID)     | Required  |      |
| ID2 (ID2)     | Optional  |      |
| ID3 (ID3)     | Optional  |      |
| Visit         | Optional  |      |
| OPID          | Optional  |      |
| Clinic        | Optional  |      |
| Draw Time     | Optional  |      |
| Received Time | Optional  |      |

## **IRC**

The IRC preset project has these requirements

### **Data entry requirements**

| Field           | Required? | Note  |
|-----------------|-----------|---|
| ID1 (PID)       | Required  |   |
| ID2 (Protocol)  | Required  |   |
| ID3 (SID)       | Required  |   |
| Visit           | Required  |   |
| OPID            | Optional  |   |
| Clinic          | Required  |   |
| Collection Time | Required  | Only required if the collection date is after 01-<br>Jan-2004 |
| Received Time   | Optional  |   |

## **KENPHIA2**

The KENPHIA2 preset project has these requirements.

| Field                              | Required? | Note   |
|------------------------------------|-----------|--|
| ID1 (PTID)                         | Required  | Must be length 8. First 2 characters are letters only (KE) and last 6 characters are numeric only (e.g., KE123456)               |
| ID2 (Country)                      | Required  | "KENYA" is the only available option   |
| ID3 (HIVRT)                        | Required  | Pick list of specific options  |
| Collection Time                    | Required  |  |
| Received Date and<br>Received Time | Required  |  |
| Clinic                             | Required  | Must be formatted like XX### where the first 2 characters are the letters KE and the next 3 characters are numeric (e.g., KE101) |

## **MACS**

The MACS preset project has these requirements.

### **Data entry requirements**

| Field         | Required? | Note  |
|---------------|-----------|---|
| ID1 (MACSID)  | Required  | MACS Identifier   |
| ID2 (Study)   | Required  |   |
| ID3           | Optional  | Previously entered ID3s will be automatically populated by LDMS for the MACSID and study combination    |
| Visit         | Optional  | Select from pre-defined list of visit units.  |
| OPID          | Optional  |   |
| Clinic        | Optional  | Previously entered clinics will be automatically populated by LDMS for the MACSID and study combination |
| Specimen Time | Optional  |   |
| Received Time | Optional  |   |

#### **Notes**

- Quick Add templates are permitted for MACS.
- MACS shipments being sent to Precision Bioservices (formally SeraCare) use the SeraCare shipping file format.

## **MATRIX**

The MATRIX preset project has these requirements.

| Field           | Required? | Note |
|-----------------|-----------|------|
| ID1 (PTID)      | Required  |      |
| ID2 (Study)     | Required  |      |
| ID3             | Optional  |      |
| Collection Time | Required  |      |

## **MAVRC**

The MAVRC preset project must meet these requirements.

### **Data entry requirements**

| Field          | Required? | Note |
|----------------|-----------|------|
| ID1 (PID)      | Required  |      |
| ID2 (Protocol) | Optional  |      |
| ID3 (ID3)      | Optional  |      |
| Visit          | Optional  |      |
| OPID           | Optional  |      |
| Clinic         | Optional  |      |
| Draw Time      | Optional  |      |
| Received Time  | Optional  |      |

## **MOSAIC**

The MOSAIC preset project has these requirements.

### **Data entry requirements**

| Field           | Required? | Note |
|-----------------|-----------|------|
| ID1 (PTID)      | Required  |      |
| ID2 (Study)     | Required  |      |
| ID3             | Optional  |      |
| Collection Time | Required  |      |

# **mStudy**

The mStudy preset project must meet these requirements.

| Field         | Required? | Note                              |
|---------------|-----------|-----------------------------------|
| ID1 (PID)     | Required  | 6 digit numeric format            |
| ID2 (Study)   | Required  | Pre-populated with value = MSTUDY |
| ID3 (ID3)     | Optional  |                                   |
| Visit         | Required  |                                   |
| OPID          | Optional  |                                   |
| Clinic        | Required  |                                   |
| Draw Time     | Required  |                                   |
| Received Time | Required  |                                   |

## **MTCT**

The MTCT preset project has these requirements

### **Data entry requirements**

| Field          | Required? | Note |
|----------------|-----------|------|
| ID1 (PID)      | Required  |      |
| ID2 (Protocol) | Optional  |      |
| ID3 (ID3)      | Optional  |      |
| Visit          | Optional  |      |
| OPID           | Optional  |      |
| Clinic         | Optional  |      |
| Draw Time      | Optional  |      |
| Received Time  | Optional  |      |

## **MTN**

The MTN preset project has these requirements.

| Field          | Required? | Note   |
|----------------|-----------|--|
| ID1 (PID)      | Required  | 9 characters long                                      |
| ID2 (Protocol) | Required  | Select from a pre-populated list                       |
| ID3            | Disabled  | Not used for MTN                                       |
| Visit          | Required  | Visit unit defaults to VST, but this can be overridden |
| OPID           | Disabled  | Not used for MTN                                       |

| Field         | Required? | Note  |
|---------------|-----------|---|
| Clinic        | Optional  |   |
| Specimen Time | Optional  | This field is not required, but recommended |
| Received Time | Optional  | This field is not required, but recommended |

#### **Notes**

- Primary and aliquot volume will default to ML for MTN specimens.
- There are two label options available for MTN, one with a barcode and one without.
- Quick Add templates are permitted for MTN.

### **MWCCS**

The MWCCS preset project has these requirements.

#### **Data entry requirements**

| Field         | Required? | Note   |
|---------------|-----------|--|
| ID1 (PID)     | Required  | 8 digits long  |
| ID2 (Study)   | Required  | Select from a pre-populated list (e.g., MWCCS)                               |
| ID3           | Optional  |  |
| Visit         | Optional  |  |
| OPID          | Optional  |  |
| Clinic        | Required  | Select from pre-populated list. Correct options defined as XNNN (e.g., X206) |
| Specimen Time | Optional  | This field is not required, but recommended                                  |
| Received Time | Optional  | This field is not required, but recommended                                  |

#### **Notes**

• The cross-LIMS shipping file format should be used for all shipments going to Precision Biosciences (lab 512).

## **NICHD-Westat**

The NICHD-Westat has the same requirements the ACTG/IMPAACT project.

### **PHACS**

The PHACS preset project has these requirements

| Field          | Required? | Note  |
|----------------|-----------|---|
| ID1 (PID)      | Required  | 8 characters long, 7 numbers with a letter at the end   |
| ID2 (Protocol) | Required  | Select from a pre-populated list; all protocols are in the format PHXXX (e.g. PH200).   |
| ID3            | N/A       | Not used by PHACS   |
| Visit          | Required  | Select from pre-defined list of visit units.  |
| OPID           | Optional  |   |
| Clinic         | Required  | Will automatically populate using the previous clinic for the project, PID, and protocol combination entered; PHACS clinics range between 1 and 24. LDMS will check that a valid clinic was selected. |
| Specimen Time  | Required* | *Required for HOPE, PH700, and all future PHACS studies. Optional but recommended for all previous or historical studies.   |
| Received Time  | Optional  | This field is optional, but recommended   |

#### Notes

- PHACS uses the standard LDMS shipping file format.
- There are two label options for PHACS specimens, one with a barcode and one without.
- Quick Add templates can be applied for PHACS, but users are not permitted to create their own templates. All PHACS templates are created by Frontier Science in collaboration with PHACS leadership.

## **PHIA**

The PHIA preset project has these requirements

| Field         | Required? | Note  |
|---------------|-----------|---|
| ID1 (PTID)    | Required  | [Country_Code][6_digits] (Example: ZW123456)  |
| ID2 (Country) | Required  | Automatically populated based on country code in PTID; SW (Swaziland) differs from the ISO 3166 country code (SZ) |
| ID3 (HIVRT)   | Required  | Testing result (POS, NEG, or IND)   |
| Visit         | Required  |   |
| OPID          | Optional  |   |
| Clinic        | Required  | A warning will appear if the country codes in the PTID and clinic do not match                                    |

| Field         | Required? | Note |
|---------------|-----------|------|
| Specimen Time | Required  |      |
| Received Time | Required  |      |

#### **Notes**

- Quick Add templates *cannot* be created for this project.
- Quick Add templates can be used for this project.

## **REPRIEVE**

The REPRIEVE preset project has the same requirements as the ACTG/IMPACCT project.

## **SHIMS**

The SHIMS preset project must meet these requirements.

### **Data entry requirements**

| Field            | Required? | Note   |
|------------------|-----------|--|
| ID1 (PID)        | Required  | Nine characters long that meets the SHIMS ID1 requirements |
| ID2 (Study)      | Required  |  |
| ID3 (Rapid Test) | Required  |  |
| Visit            | Required  |  |
| OPID             | Optional  |  |
| Clinic           | Optional  |  |
| Draw Time        | Required  |  |
| Received Time    | Required  |  |

#### **SHIMS ID1 format**

RAAAhhcpp

| R   | 1 to 4      |
|-----|-------------|
| AAA | 001 to 599  |
| hh  | 01 to 26    |
| С   | Check digit |
| рр  | 01 to 99    |

## **SNRP**

The SNRP preset project has these requirements

## **Data entry requirements**

| Field         | Required? | Note |
|---------------|-----------|------|
| ID1 (PID)     | Required  |      |
| ID2 (Study)   | Optional  |      |
| ID3 (SID)     | Optional  |      |
| Visit         | Optional  |      |
| OPID          | Optional  |      |
| Clinic        | Optional  |      |
| Draw Time     | Optional  |      |
| Received Time | Optional  |      |

## **TIES**

The TIES preset project has these requirements.

## **Data entry requirements**

| Field          | Required? | Note   |
|----------------|-----------|--|
| ID1 (PID)      | Required  | 8 characters long, 7 numbers and one letter<br>between A and L; if the PID does not have 7<br>numbers, add leading zeros to the front of it. |
| ID2 (Protocol) | Required  | Select from the pre-defined list of protocols.   |
| ID3 (ID3)      | Available | ID3 is not used by this project  |
| Visit          | Required  | Select from pre-defined list of visit units.   |
| OPID           | Available |  |
| Clinic         | Required  | In the format T[Clinic_Number]   |
| Specimen Date  | Required  |  |
| Specimen Time  | Required  |  |
| Received Date  | Required  |  |
| Received Time  | Optional  |  |

# **VQA**

The VQA preset project has these requirements.

| Field         | Required? | Note  |
|---------------|-----------|---|
| ID1 (PanelID) | Required  | LDMS will determine if this field is valid.<br>Specimens entered or received in a shipping<br>file prior to 01-July-2007 will not be validated. |
| ID2           | N/A       | This field is not used by VQA   |
| ID3           | N/A       | This field is not used by VQA   |
| Visit         | N/A       | This field is not used by VQA   |
| OPID          | N/A       | This field is not used by VQA   |
| Clinic        | N/A       | This field is not used by VQA   |
| Specimen Time | N/A       | This field is not used by VQA   |
| Received Time | N/A       | This field is not used by VQA   |

#### **PanelID format**

[Assay\_Category][Panel\_Number][Panel\_Subtype].[Num]Panel\_Configuration

Assay\_Category RNA, DNA, or CUL

Panel\_Number A number between 000 and 999

Panel\_Subtype

| UM | Ultrasensitive RNA          |
|----|-----------------------------|
| SM | Standard RNA                |
| RT | DNA proficiency panel       |
| PQ | DNA pre-qualification panel |
| CC | Culture panel               |

**Num** A number between 01 and 99

Panel\_Configuration,nB, or C

### **VTN**

The VTN preset project has these requirements.

| Field          | Required? | Note   |
|----------------|-----------|--|
| ID1 (PID)      | Required  | LDMS checks that the PID is valid. Must<br>be 9 or 10 characters. A PID cannot be<br>changed without approval from VTN Laboratory<br>Leadership. |
| ID2 (Protocol) | Required  | LDMS checks that the PID and protocol combination are valid.   |
| ID3            | Disabled  |  |

| Field         | Required? | Note   |
|---------------|-----------|--|
| Visit         | Required  | Use VST for routine HIV diagnostic visits, otherwise use the appropriate visit unit. This visit unit is used by certain laboratories for driving testing algorithms.   |
| OPID          | Disabled  |  |
| Clinic        | Required  | The clinic is automatically populated based on the PID; the same clinical location can be associated with multiple clinic IDs (e.g. for Phase I/II or Phase II/III). VTN clinics start with the letter V (such as V101 instead of 101). LDMS will verify that a VTN clinic was selected. |
| Specimen Time | Required  |  |
| Received Time | Required  |  |

#### **Notes**

- Throughout LDMS, HVTN is listed as VTN.
- The volume for VTN specimens will default to ML.
- Within VTN, LDMS Shipping files are sometimes called "e-manifests".
- Barcodes must be printed on labels for VTN specimens.
- Quick Add templates are not permitted for VTN specimens.
- Information for study CoVPN 5001 should be logged under VTN study 5001.

# **WHIN**

The WHIN preset project must meet these requirements.

### **Data entry requirements**

| Field               | Required? | Note |
|---------------------|-----------|------|
| ID1 (ParticipantID) | Required  |      |
| ID2 (Protocol)      | Optional  |      |
| ID3 (ID3)           | Optional  |      |
| Visit               | Optional  |      |
| OPID                | Optional  |      |
| Clinic              | Optional  |      |
| Draw Time           | Optional  |      |
| Received Time       | Optional  |      |

# **WIHS**

The WIHS preset project has these requirements.

### **Data entry requirements**

| Field          | Required? | Note  |
|----------------|-----------|---|
| ID1 (PID)      | Required  | Must meet WIHS PID format requirements  |
| ID2 (Protocol) | Required  | Select from pre-populated list  |
| ID3 (SID)      | Optional  |   |
| Visit          | Required  | The visit unit will default to VST, and the visit number will be automatically populated based on an algorithm that uses the current date |
| OPID           | Optional  |   |
| Clinic         | Optional  |   |
| Specimen Time  | Optional  |   |
| Received Time  | Optional  |   |

#### **WIHS PID format**

[Site\_Number][Recruitment\_Number][Participant\_Number][Check\_Digit]

### Site\_Number

| 1 | NYC (Bronx)      |
|---|------------------|
| 2 | Brooklyn         |
| 3 | Washington, D.C. |
| 4 | Los Angeles      |
| 5 | San Francisco    |
| 6 | Chicago          |

#### Recruitment\_Number

| 2B | New recruit      |
|----|------------------|
| 0B | Original recruit |
| 1B | Original recruit |

### Check\_Digit

Any character; an algorithm will determine if the character is valid based on the read of the PID.

#### **Notes**

- WIHS shipments being sent to Precision Bioservices (formally SeraCare) use the SeraCare shipping file format.
- Quick Add templates are permitted for WIHS, and users are expected to create and use template for WIHS data entry.
- Collection Date and Received Date will automatically be set to your local date.

# **WITS**

The WITS preset project has these requirements

### **Data entry requirements**

| Field         | Required? | Note |
|---------------|-----------|------|
| ID1 (PID)     | Required  |      |
| ID2 (ID2)     | Optional  |      |
| ID3 (ID3)     | Optional  |      |
| Visit         | Optional  |      |
| OPID          | Optional  |      |
| Clinic        | Optional  |      |
| Draw Time     | Optional  |      |
| Received Time | Optional  |      |

# **WITSRepos**

The WITSRepos preset project must meet these requirements.

### **Data entry requirements**

| Field            | Required? | Note |
|------------------|-----------|------|
| ID1 (Subject ID) | Required  |      |
| ID2 (Study ID)   | Optional  |      |
| ID3              | Optional  |      |
| Visit            | Optional  |      |
| OPID             | Optional  |      |
| Clinic           | Optional  |      |
| Draw Time        | Optional  |      |
| Received Time    | Optional  |      |

# **ZEBS**

The ZEBS preset project has these requirements

### **Data entry requirements**

| Field         | Required? | Note |
|---------------|-----------|------|
| ID1 (STUDYID) | Required  |      |
| ID2 (ID2)     | Optional  |      |
| ID3 (ID3)     | Optional  |      |
| Visit         | Optional  |      |
| OPID          | Optional  |      |

| Field         | Required? | Note |
|---------------|-----------|------|
| Clinic        | Optional  |      |
| Draw Time     | Optional  |      |
| Received Time | Optional  |      |

# ZIP

The ZIP preset project has these requirements.

### **Data entry requirements**

| Field                        | Required? | Note  |
|------------------------------|-----------|---|
| ID1 (PID)                    | Required  | Must be a 6-digit (for ID2=ZIP) or 7-digit (for other ID2 values). Following the digits, there must be a letter. A hard check should run to ensure the first two digits match to a corresponding ZIP clinic code ZXX. |
| ID2 (Study)                  | Required  | Use allowed studies (ZIP, Z1.0, or Z2.0)  |
| ID3 (Screening ID)           | Required  | It must be in the format SAAXXXX, where AA is a valid clinic number (i.e., ZAA) and XXXX is any numeric number.   |
| Visit Value and Vist<br>Unit | Required  |   |
| Clinic                       | Required  | The LDMS will automatically populate the correct, corresponding clinic ID based off of the entered PID. If a corresponding clinic ID is not found, leave the clinic field blank and select a value.                   |
| Specimen Date                | Required  |   |
| Specimen Time                | Required  |   |
| Received Date                | Required  |   |
| Received Time                | Required  |   |

#### **Notes**

- The application must allow FSTRF and user defined preloads for this project.
- The application must not restrict the user from co-enrolling specimens for this project.

# **Administration**

The **Administration** page is where you can manage various settings in LDMS

# **Projects**

Projects are the collaborative project, organization, or network that conduct studies.

There are two types of projects: *local projects* and *government projects*.

Figure 66: The Projects page.

|          |  | O           | 2        | <b>6</b> |          |          |          |        |
|----------|--|-------------|----------|----------|----------|----------|----------|--------|
| Name     | Description                                | ID1         | ID2      | ID3      | Export   | Local    | In Use   |        |
| PHIA     | Population-Based HIV<br>Impact Assessments | PTID        | Country  | HIVRT    | ₩        |          | ₩        | View   |
| CONTROL  | CONTROL                                    | CONTROLNAME | PROTOCOL | ID3      | <b>~</b> |          | <b>~</b> | View   |
| UNKNOWN  | Unknown Group                              | ID1         | ID2      | ID3      |          |          |          | View   |
| FRONTIER | Frontier Science-Internal<br>Study         | PID         | Study    |          |          | ₩        | ₩        | Edit   |
| cadmus   | cadmus                                     | pid         | prot     | sid      |          | <b>~</b> |          | Edit 🔻 |
| XL0CAL   | XLOCAL                                     | ID1         | ID2      | ID3      |          | <b>V</b> | <b>~</b> | Edit   |

(1) Whether data is copied into Frontier Science's central database (2) Whether the project is a government or local project (3) Whether there are specimens currently using the projects.

# **Local projects**

Local projects are *projects* that only exist at your laboratory.

A local project is one that has been defined by your laboratory. These are general used for internal studies that only your laboratory will utilize, although they can be sent to other laboratories through shipments.

Defining a new local project requires you to specify what you are going to call *ID1*, *ID2*, and *ID3*.

**Table 16: Identifiers** 

| Identifie | Description  | Example          |
|-----------|--|------------------|
| ID1       | Uniquely identifies a participant or specimen source         | Participant ID   |
| ID2       | Identifies the study in which the participant is enrolled    | Study number     |
| ID3       | Used to further identify a branch or sub-<br>unit in a study | Sub-study number |

While there are no rules for what you can name each identifier, they must be used for these purposes. For some projects, it may be more appropriate to think of ID1 as identifying the source from which specimens were collected, such as a testing solution that you received, rather than a participant.

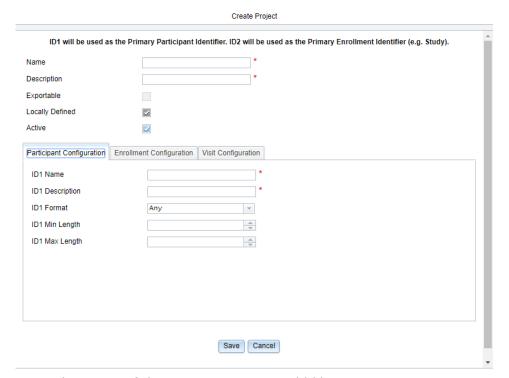
#### Creating a new project

New projects can be created on the **Project Administration** page.

#### Steps

- 1. Click **Administration** > **Projects** from the LDMS menu bar.
- 2. From the LDMS action menu, select **Create Project**. The **Create Project** window will open.

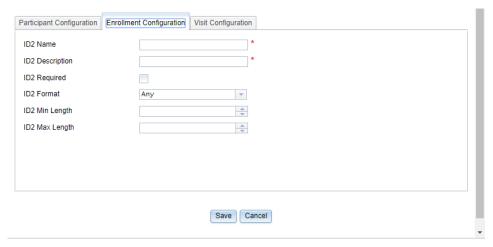
Figure 67: The Create Project Window



- **3.** Enter the name of the project as you would like it to appear in menus throughout LDMS into the **Name** box.
- **4.** Enter a brief description of the project into the **Description** box. The description is generally used to provide the full name of the project, especially when the project is an acronym.
- 5. Enter the name that you want to use for ID1 for the project into the ID1 Name box, and a brief description into the ID1 Description box. The format and acceptable values for ID1 may also be set.
  ID1 is typically used as an identifier for a participant. For other types

of work, such as quality control testing, ID1 could also be something else, like a testing panel number.

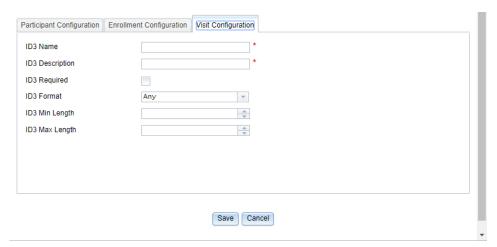
**Figure 68: Enrollment Configuration** 



**6.** Enter the name that you want to use for *ID2* into the **ID2 Name** box, and a brief description into the **ID2 Description** box. The format and acceptable values for *ID2* may also be set.

ID2 is typically used for a study or protocol identifying number.

Figure 69: Visit Configuration



7. Enter the name that you want to use for *ID3* into the **ID3 Name** box, and a brief description into the **ID3 Description** box. The format and acceptable values for *ID3* may also be set.

ID3 is typically used as a sub-study or sub-protocol identifying number.

**8.** Click the **Save** button.

#### Result

The new project will be added to the list of groups that you specified.

#### Modifying and removing a project

Local project can be modified or removed after they are created

### **Prerequisites**

A local project can only be removed if it is not currently in use. Government projects cannot be modified or removed.

#### **Background**

This might be needed if there was a mistake with the way the project was initially entered or if you need to update the descriptions for the participant fields. Changes made to a project will automatically update existing participant records in LDMS.

#### Steps

- 1. Click **Administration** > **Projects** from the LDMS menu bar.
- **2.** Locate the project to be modified or removed from the list of projects.
- **3.** Do one of the following:
  - To modify the project, click the **Edit** button to the right of the project. The **Edit Project** window will open, where you can make your changes. When you are done, click the **Save** button.
  - To remove the project, select **Delete** from the **Edit** menu to the right of the project.

# **Government projects**

Government projects are those that are defined by Frontier Science.

Frontier Science works with government-funded projects to define the rules and specifications for their project. These projects cannot be modified or removed by users. Even though they cannot be modified, they will still be visible on the **Projects** page.

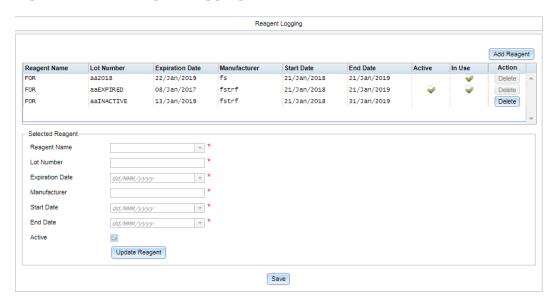
# Adding a Reagent

The Reagent Logging feature is used to track reagents used at a laboratory and uses a defined set of reagent codes.

You can use this feature to record information about reagents, such as the lot number and whether the reagent is still in use.

Once entered, the lot number for the reagent will be available on the **Specimen Management** page to assign to specimens that contain the reagent.

Figure 70: The reagent logging module



# **Adding Reagent Information**

Reagents can be added through the **Administration** menu.

#### **Steps**

- From the menu bar, click **Administration** and then click **Reagent Logging**
- 2. Click Add Reagent

A new blank row will be added to the bottom of the name. You may need to scroll down to see it.

**3.** Complete the information for the new reagent.

| Field           | Usage  |
|-----------------|--|
| Reagent Name    | This field contains the three letter code for the reagent that is being tracked.           |
| Lot Number      | This is a free text field. Enter the lot number from the container                         |
| Expiration Date | Manufacturer set date of expiration  |
| Manufacturer    | This is a free text field. Enter the manufacturer name from the container.                 |
| Start/End Date  | These fields record the date the reagent was first used and the date use was discontinued. |
| Active          | New entries default to active.   |

**4.** At the bottom of the page, click **Save**.

# Linking Reagent to Sample in Specimen Management

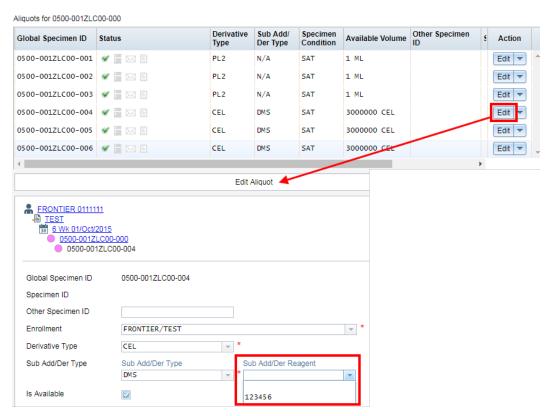
Users can assign reagent lots to primary and aliquot samples in <code>Specimen</code> <code>Management</code> or via <code>Quick</code> <code>Add</code>. Options are limited to the lots that have been defined in the reagent logging module.

#### **Background**

After the lot is entered in the Reagent Logging module, the lot number is available in the Edit Aliquot menu to be applied to any sample using the subadditive.

#### **Steps**

# Figure 71: Linking Reagent to Sample via the Specimen Management Menu



- In Specimen Management, use the filters to navigate to the specimen's page.
- 2. In the Aliquot grid, locate the specimens utilizing the logged reagent
- 3. Click the Edit button, the Edit Aliquot menu will open in a pop up window
- 4. Select the appropriate lot number in the Sub Add/Der Reagent menu
- **5.** Click Save

# Linking Reagent to Sample in Quick Add

#### Background

If known, the reagent lot number can be applied in Quick Add at the time of accessing.

#### **Steps**

- 1. In the Aliquot grid, locate entry with the Sub Additive/Derivative.
- 2. Click the Edit button
- **3.** Select the lot number in the Sub Add/Der Reagent menu.

# **Linking Additive to Reagent Lot**

Steps

#### Figure 72: Linking Additive to Reagent Lot



- 1. In Specimen Management, click the Edit button for the Primary containing the logged Additive
- 2. In the Edit Primary Specimen window, select the lot number in the Additive Reagent menu

# **Reagent Lot Sample Details Report**

The Reagent Lot Sample Details Report provides a list of which specimens contain a reagent.

This report include information about the reagent at the top, and then lists all specimens and details about the specimens (such as specimen type, collection date and time, and specimen identifiers).

To generate this report, click **Administration** and then click **Reagent Logging**. Next to the reagent of interest, click **Report**.

# **Specimen Anonymization**

The specimen anonymization module is used to anonymize specimens. This module is disabled by default and must be enabled by LDMS User Support in order to be used.

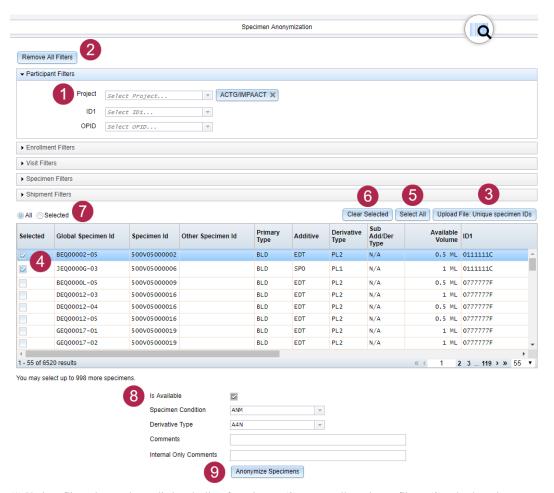


Figure 73: Specimen Anonymization Screen

- (1) Various filters that can be applied to the list of specimens, (2) remove all specimens filters, (3) upload a unique specimen IDs file, (4) manually select specimens from filtered list, (5) select all specimens in filtered list, (6) clear selected specimens in filtered list, (7) filter list of specimens by selected only, (8) mark specimen(s) as available or non-available, set condition and derivative type, and add comments, (9) anonymize selected specimens
- To access the specimen anonymization module after it has been activated by LDMS User Support, navigate to the **Administration** dropdown and select **Specimen Anonymization**.
- **2.** Use the various filters to locate the specimens that need to be anonymized. The list of filtered specimens will appear below.
- **3.** If not all specimens in the filtered list are intended to be anonymized, select the specimens that are to be anonymized and click the **Selected** radio button.
- **4.** A global specimen IDs file can also be added to the list of filtered specimens by clicking **Upload File: Unique specimen IDs**.
- **5.** Check the **Is Available** checkbox to have the selected specimens remain available.
- **6.** Change the **Specimen Condition** of the selected specimens from the dropdown.



**Note:** Certain specimen conditions automatically change the specimen availability checkbox.

- 7. Change the **Derivative Type** and add **Comments** and **Internal Only Comments** to the selected specimens as necessary.
- 8. Click Anonymize Specimens.

After specimens have been anonymized, the original specimen records will become unavailable to store, ship, or test. LDMS will create new anonymized records for these specimens with scrambled/anonymized IDs. New barcode labels can be generated to apply to the specimen vials that display the anonymized IDs. Reports can be generated to review the link between the original IDs and newly assigned anonymized IDs.

# **Lab Settings**

Some laboratory-specific settings can be controlled by users by clicking **Administration** and then clicking **Lab Settings**.

| Setting  | Usage   |
|--|---|
| Number of samples shown by default in sample picker  | Set the default number of samples shown in sample picker  |
| Processing Tech Initials defaults to current user  | Currently signed-in user will automatically be set as the Processing Technician when creating aliquots.                                     |
| Require a comment when modifying primaries and aliquots  | If activated, the user must enter or modify the comment in order to be able to save the Edit Primary and Edit Aliquot popups.               |
| Auto-set frozen date/time to the current date/time when storing specimens  | When storing a specimen, its frozen date and time will automatically be set.  |
| Include unavailable primary samples by default when printing labels  | Unavailable primary samples will be included by default when printing labels.   |
| Auto-assign next available fill-order position when adding and moving specimens, containers, and levels in storage | The next available fill-order will automatically be assigned when specimens are added to storage containers                                 |
| Import shipments into storage by default   | Shipments will be imported into storage by default  |
| Mark to Ship reminds users when overdue by default   | Users will be notified on login if they have overdue shipments  |
| Override shipping container configurations when importing into storage?  | Override various attributes for shipping container configurations when importing into storage, such as alphabetic columns, fill order, etc. |

# **User Settings**

Users are able to change a limited number of settings for how their account works by clicking **Administration** and then clicking **User Settings**.

| Setting  | Usage  |
|--|--|
| Default Module                                   | Select the initial page that will be loaded when you sign into LDMS. |
| Default Lab ID                                   | Select the lab ID that will be selected when you sign into LDMS.     |
| Show ID1/PID and Specimen Number in Storage Tree | Select to display ID1/PID and Specimen<br>Number in the Storage Tree |

# User management

User accounts are managed by Frontier Science.

Accounts for individual users are created and maintained by LDMS User Support. To perform the following tasks, you would need to contact LDMS User Support for assistance:

- · Create a new user
- · Reset a user's password
- Manage what features in LDMS a user can access

Users can modify their own passwords from within LDMS.

# **Changing your password**

While the creation and management of user accounts is handled by LDMS User Support, users can change their own passwords.

#### Steps

1. From the logout menu in the upper-right corner of the page, click **Change Password** 

Figure 74: The Logout menu



The **Change password** window will open.

- **2.** Enter your current password into the **Password** box.
- **3.** Enter your new password into the **New Password** box.
- **4.** Re-enter your new password into the **Confirm Password** box.
- **5.** Click the **Change Password** button.

# **Password requirements**

This section explains the requirements for user passwords.

 Passwords must contain at least one upper case letter, one lower case letter, one number, and one special character

- A password must not be the same as one of the your previous passwords
- A password must be between 10 and 50 characters
- Must not be similar to your current password, user ID, or display name

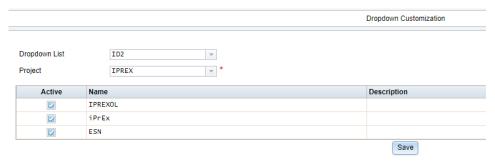
Temporary passwords expire within 24 hours. If you do not sign into LDMS using the temporary password before it expires, you must have a new temporary password generated.

# **Dropdown Customization**

The **Dropdown Customization** tab allows users to hide items from dropdown lists. This is accessed through the **Administration** menu.

### Steps

1. From the menu bar, click **Administration** and then click **Dropdown Customization**.



- **2.** Select a value from **Dropdown List**.
- **3.** Select a project from the **Project** dropdown list.
- **4.** A table with items to hide will appear. Uncheck the boxes in the **Active** column to hide items.
- **5.** At the bottom of the page, click **Save**.

To unhide any items previously hidden, return to this page and recheck the applicable boxes, then click **Save**. The items will then return as options in dropdown lists.

# **RPID Requests**

In the RPID Requests module, users can create new RPID requests, edit/delete existing requests, and download a list of ID1s associated with the request.

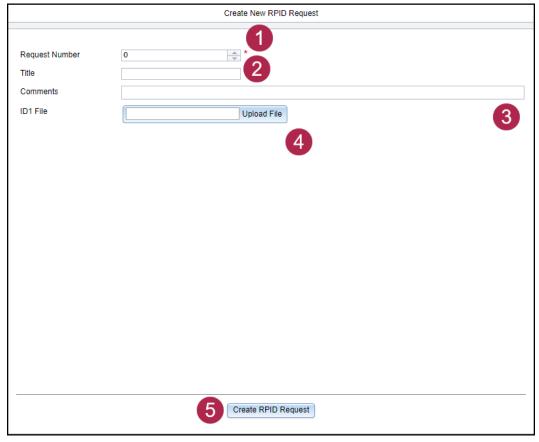
Figure 75: RPID Requests - Main Screen



(1) Add a new RPID request, (2) View, edit, delete existing RPID requests

# Adding a New RPID Request

Figure 76: Create New RPID Request Window



(1) RPID Request Number, (2) title of the request, (3) comments about the request, (4) upload ID1 file, (5) create the RPID request

- 1. Navigate to RPID Requests from the Administration dropdown.
- 2. Click Add RPID Request from the Actions dropdown.
- **3.** In the **Create New RPID Request** window, add the request number, title, comments, and ID1 file as needed, and click **Create RPID Request**.

# **Editing an RPID Request**

- **1.** Navigate to **RPID Requests** from the **Administration** dropdown.
- 2. Click **View/Edit** next an existing RPID request.
- 3. Update the request information as needed.
- 4. Click Save.
- 5. To delete an RPID request, return to the main RPID Request screen, click the dropdown next to the View/Edit button of an existing request and click Delete. On the confirmation screen, click Delete.

A list of ID1s associated with an RPID request can be downloaded. To do this, click the dropdown next to an existing RPID request and click **Download ID1s**.

# **Clear Filters**

The Clear Filters page is used to clear filters across all modules.

#### Figure 77: Clear Filters Screen



To access the Clear Filters module, navigate to the **Administration** dropdown and select **Clear Filters**.

To clear filters, do one of the following:

- To clear specific filters, check the boxes next to each applicable filter and then click **Clear**.
- To clear filters in all categories, click Clear All.

# **Online Resources**

The Online Resources menu option contains links to relevant web resources.

- LDMS Validation Resources A link to a page on the LDMS website where users can request validation documentation.
- Frontier Science Website A link to the main Frontier Science website.
- Frontier Science Portal A link to the Frontier Science Portal.
- **Specimen Repository Website** A link to the specimen repository website, a site that can be used to search for specimens located at a number of repositories.

# Codes, units, and abbreviations

This section lists the codes, units and abbreviations that are used in LDMS for various entry fields, such as visits and specimen types. These codes are used by both LDMS and LDMS for Windows.

# Specimen type codes

These are the codes use for primary, additive, and derivative types.

# **Primary codes**

This section lists all of the available primary codes in LDMS

| Code | Description               |
|------|---------------------------|
| ADG  | Adrenal Glands            |
| ADP  | Adipose                   |
| AHR  | Non-Occipital scalp hair  |
| AMN  | Amniotic Fluid            |
| ANL  | Anal                      |
| AOR  | Aorta                     |
| APP  | Air Pollution Particles   |
| ARB  | Rectal Biopsy by Anoscopy |
| ARL  | Breast Areola             |
| ART  | Arterial Blood Draw       |
| ASC  | Ascending Colon           |
| BAL  | Bronchoalveolar Lavage    |
| BHR  | Hair, Body                |
| BLA  | Bladder                   |
| BLC  | Blood Clots               |
| BLD  | Blood (Whole - Venous)    |
| ВМА  | Bone Marrow Aspirate      |
| ВМС  | Bone Marrow Core Biopsy   |

| Code | Description              |
|------|--------------------------|
| ВМК  | Human Milk               |
| BML  | Human Milk - Left        |
| BMR  | Human Milk - Right       |
| BNE  | Bone                     |
| BNM  | Bone Marrow              |
| BRB  | Bronchoscopy Brush       |
| BRN  | Brain Tissue             |
| BRS  | Breast Tissue            |
| BSG  | Basal Ganglia            |
| BUC  | Buccal                   |
| BUG  | bulbourethral glands     |
| CAT  | Coronary Artery          |
| CCD  | Common Carotid           |
| CDE  | Caudate                  |
| CER  | Cervix                   |
| CLL  | Colon (left)             |
| CLR  | Colon (right)            |
| CLS  | Sigmoid Colon            |
| CRB  | Cerebellum               |
| CRD  | Cord Blood               |
| CRN  | Colorectal Neoplasm      |
| CSC  | Cervical Secretion       |
| CSF  | Cerebro-Spinal Fluid     |
| CVB  | Cervical Biopsy/Aspirate |
| CVF  | Cervicovaginal Fluid     |
| CVL  | Cervical Vaginal Lavage  |
| CVM  | Cervical Mucous          |
| CXC  | Cervical Culturette      |
| CXS  | Cervical Swab            |
| DDM  | Duodenum                 |
| DIA  | Dialysate                |
| DMM  | DMEM                     |
| DPL  | Dried Plasma             |
| DRG  | Dorsal Root Ganglia      |
| DSC  | Descending Colon         |

| Code | Description                                    |
|------|--|
| DTC  | Distal Colon                                   |
| DWB  | Dried Whole Blood                              |
| DWM  | Deep White Matter                              |
| EBC  | Exhaled Breath Condensate                      |
| END  | Endometrium                                    |
| ENV  | Environmental                                  |
| EPI  | Epididymus                                     |
| EPS  | Expressed Prostatic Secretions                 |
| ESO  | Esophagus                                      |
| EVX  | Endocervix                                     |
| EYE  | Corneal Tissue                                 |
| FAT  | Femoral Artery                                 |
| FCM  | Frontal Cortex (motor)                         |
| FCP  | Frontal Cortex (pre-motor)                     |
| FGN  | Finger Nails                                   |
| FHR  | Hair, Facial                                   |
| FLT  | Fallopian Tubes                                |
| FSC  | Sigmoid Colon Biopsy by Flexible Sigmoidoscopy |
| FSI  | Penis Foreskin, Inner Tissue                   |
| FSK  | Penis Foreskin, Whole Tissue                   |
| FS0  | Penis Foreskin, Outer Tissue                   |
| FSR  | Rectal Biopsy by Flexible Sigmoidoscopy        |
| FST  | Blood from Fingerstick (capillary)             |
| GAS  | Gastric Secretions                             |
| GCF  | Gingival Crevicular Fluid                      |
| GLN  | Glans  |
| GLS  | Glycerol Stock                                 |
| GLU  | Genital Lesion/Ulcer                           |
| GMI  | Gastro Intestinal Mucosa                       |
| HAR  | Hair   |
| HPC  | Hippocampus                                    |
| HRT  | Heart  |
| HST  | Blood from Heelstick (capillary)               |
| ICD  | Internal Carotid                               |
| ILM  | Ileum  |

| Code | Description                  |
|------|------------------------------|
| INT  | Intestinal Biopsy/Aspirate   |
| INW  | Induction Waste              |
| IVR  | Intra-vaginal Ring           |
| IVS  | Intraventricular Septum      |
| JJM  | Jejunum                      |
| KDY  | Kidney                       |
| KID  | Kidney Biopsy                |
| LAL  | Lymph Node Axillary-left     |
| LAR  | Lymph Node Axillary-right    |
| LAT  | Left Atrium                  |
| LBL  | Large Bowel                  |
| LBT  | Lateral Border of the Tongue |
| LIL  | Lymph Node Inguinal-left     |
| LIR  | Lymph Node Inguinal-right    |
| LIV  | Liver Tissue                 |
| LNA  | Lymph Node (aortic)          |
| LNC  | Lymph Node (cervical)        |
| LNE  | Pre-auricular Lymph Node     |
| LNF  | Infraclavicular Lymph Node   |
| LNG  | Lung                         |
| LNI  | Inguinal Lymph Node          |
| LNK  | Popliteal Lymph Node         |
| LNM  | Lymph Node (mediastinal)     |
| LNO  | Lymph Node (other)           |
| LNP  | Lymph Node (per-hilar)       |
| LNR  | Para-aortic Lymph Node       |
| LNS  | Supraclavicular Lymph Node   |
| LNT  | Mesenteric Lymph Node        |
| LNX  | Lymph Node (axillary)        |
| LPK  | Leukopak                     |
| LSG  | Labial Salivary Gland        |
| LUE  | Lung, Excised                |
| LUF  | Lung, Filter Paper           |
| LVE  | Left Ventricle               |
| LYM  | Lymph Node Biopsy/Aspirate   |

| Code | Description   |
|------|---|
| MBR  | Midbrain Consists of Cerebral Penduncles and Tectum |
| МСР  | Prefrontal Medial Cortex                            |
| MEC  | Meconium  |
| MED  | Medulla   |
| MEN  | Dura Mater - Meninges                               |
| MES  | Mesentery   |
| MFP  | Placenta Membranes                                  |
| MSB  | Main Stem Bronchus Wash                             |
| MTC  | Motor Cortex  |
| NAS  | Nasal   |
| NPH  | Nasopharyngeal                                      |
| NPW  | Nasopharyngeal Wash                                 |
| NRH  | Naso-oropharyngeal                                  |
| NSB  | Nasal brush   |
| occ  | Occip Cortex  |
| OCL  | Ocular  |
| OHR  | Occipital scalp hair                                |
| OPW  | Oropharyngeal Wash                                  |
| ORH  | Oropharyngeal                                       |
| ORL  | Oral  |
| ОТН  | Other   |
| OVR  | Ovaries   |
| PAD  | Adipose (pericardial)                               |
| PAN  | Perianal  |
| PCF  | Pericardial Fluid                                   |
| PCT  | Perietal Cortex                                     |
| PEN  | Penis   |
| PFL  | Pleural Fluid                                       |
| PFR  | Parotid Flow  |
| PHR  | Hair, Pubic   |
| PHX  | Pharynx   |
| PLC  | Placental Tissue                                    |
| PLM  | Placenta - Membrane Slice                           |
| PLN  | Peritrach Lymph Nodes                               |
| PLP  | Polyp   |

| Code | Description                    |
|------|--------------------------------|
| PLQ  | Plaque                         |
| PMD  | Plasmids                       |
| PNC  | Pancreas                       |
| PON  | Pons                           |
| PPK  | Plasmapak                      |
| PRC  | Premotor Cortex                |
| PRO  | Prostate                       |
| PRT  | Prostate Tissue Biopsy         |
| PTF  | Peritoneal Fluid               |
| PTY  | Parathyroid                    |
| RAT  | Right Atrium                   |
| RCT  | Rectum                         |
| REC  | Rectal                         |
| RSC  | Rectosigmoid Colon             |
| RVE  | Right Ventricle                |
| SAD  | Subcutaneous Adipose           |
| SAL  | Saliva                         |
| SCC  | Spinal Cord Cervical           |
| SCL  | Spinal Cord Lumbosacral        |
| SCR  | Scrotum                        |
| SCT  | Spinal Cord Thoracic           |
| SEM  | Semen                          |
| SHR  | Hair, Scalp                    |
| SIN  | Sputum, Induced Non-Select     |
| SIS  | Sputum, Induced Select         |
| SKN  | Skin                           |
| SLU  | Skin Lesion                    |
| SMD  | Skeletal Muscle (distal)       |
| SMP  | Skeletal Muscle (proximal)     |
| SMV  | Seminal Vesicle                |
| SPI  | Sputum, Induced                |
| SPL  | Spleen                         |
| SPS  | Sputum, Spontaneous            |
| SPT  | Sputum                         |
| SSN  | Sputum, Spontaneous Non-Select |

| Code | Description                 |
|------|-----------------------------|
| SSS  | Sputum, Spontaneous Select  |
| STL  | Stool                       |
| STM  | Stomach                     |
| SVG  | Salivary Glands             |
| TBD  | Specimen type to be defined |
| TCA  | Tracheal Aspirate           |
| TCX  | Temporal Cortex             |
| TER  | Tears                       |
| THD  | Thoracic Duct               |
| THL  | Thalamus                    |
| THR  | Throat Swab                 |
| THW  | Throat Wash                 |
| THY  | Thymus                      |
| TIL  | Terminal Ileum              |
| TNA  | Toe Nails                   |
| TNG  | Tongue                      |
| TON  | Tonsillar Biopsy/Aspirate   |
| TRN  | Transverse                  |
| TST  | Testes                      |
| TTH  | Tooth                       |
| TUM  | Tumor                       |
| TVC  | Transverse colon            |
| TYD  | Thyroid                     |
| UMB  | Placenta - Cord Slice       |
| UNK  | Unknown Primary             |
| URN  | Urine                       |
| UTH  | Urethra                     |
| UTS  | Uterus                      |
| VAG  | Vaginal Swab                |
| VCS  | Cervicovaginal Secretions   |
| VGL  | Vaginal                     |
| VGN  | Vagina                      |
| VSC  | Vaginal Secretions          |
| VUL  | Vulva                       |
| WAR  | Warts                       |

# **Additive codes**

This section lists all of the additive codes available in LDMS.

| Code | Description                       |
|------|-----------------------------------|
| ACD  | Acid Citrate Dextrose             |
| AG1  | ESAT 6-free cocktail 1            |
| AG2  | ESAT 6-free Cherry pick           |
| AHP  | Ammonium Heparin                  |
| APT  | Allprotect                        |
| ATM  | Abbott Transport Medium           |
| BBL  | CultureSwab Kit                   |
| BFM  | Bacterial Freezing Media          |
| BOR  | Boric Acid                        |
| BSA  | Bovine Serum Albumin              |
| втм  | Biopsy Transport Media            |
| CPD  | Citrate Phosphate Dextrose        |
| СРН  | Cell Preparation Tube Heparin     |
| CPS  | Cell Preparation Tube SCI         |
| СТК  | Culture Transport Kit             |
| DFA  | Desferoxamine                     |
| DHP  | Double dosage of Heparin          |
| DPE  | Spray Dried EDTA                  |
| DSE  | Spray Dried Sodium EDTA           |
| EDT  | EDTA                              |
| ELB  | elution buffer                    |
| END  | Endometrial Tissue                |
| ETH  | Ethanol                           |
| FFN  | Fetal Fibronectin Buffer          |
| FFP  | Formalin-Fixed, Paraffin Embedded |
| FMD  | Formaldehyde                      |
| FOR  | Formalin                          |
| GEN  | GenAptima Media                   |
| GIT  | Guanidine Isothiocyanate (GITC)   |
| GLT  | Glutaraldehyde                    |
| GRB  | Guanidine Reduction Buffer        |
| H2O  | Water                             |
| HEP  | Heparin                           |

| Code | Description                                       |
|------|---|
| HPV  | Human Papilloma Virus                             |
| HVS  | High Virginial Swab                               |
| ISO  | Isohelix kit                                      |
| IST  | Internal Standard                                 |
| LHG  | Lithium Heparin and Gel for Plasma                |
| LHP  | Lithium Heparin                                   |
| LIN  | LoBind tube lined with 5% BSA                     |
| LOB  | LoBind Tube                                       |
| LPE  | Liquid Potassium EDTA                             |
| LSE  | Liquid Sodium EDTA                                |
| LYB  | Lysis Buffer                                      |
| МВК  | Microbank tube                                    |
| MPA  | Metaphosphoric Acid                               |
| MSA  | Mannitol Salt Agar                                |
| NOH  | Sodium Hydroxide                                  |
| NON  | None  |
| NOR  | Normasol  |
| NSL  | Normal Saline                                     |
| NUC  | NUNC Tube   |
| ОСТ  | Optimum Cutting Temperature Medium                |
| OMN  | Omnigen Collection Tube                           |
| ORA  | OraSure Collection Container                      |
| ORG  | Oragene Collection Container                      |
| OTH  | Other   |
| PAC  | Port-a-cul Transport Tube                         |
| PAX  | PAXgene Blood RNA tube                            |
| PBS  | Phosphate Buffered Saline                         |
| PCT  | PreservCyt: ThinPrep collection media             |
| PED  | Potassium EDTA                                    |
| PFM  | Paraformaldahyde                                  |
| PI1  | S8820 Sigma SIGMAFAST? Protease Inhibitor Tablets |
| PIA  | Pseudomonas Isolation Agar                        |
| PLP  | PLP Fixative                                      |
| PPT  | Plasma Preparation Tube                           |
| PRO  | ProbeTec Media                                    |

| Code | Description                                       |
|------|---|
| PXD  | Paxgene tube DNA Extraction                       |
| QAG  | QFT-TB Ag Red Cap                                 |
| QMT  | QFT-Mitogen Purple Cap                            |
| QNL  | QFT-Nil Grey Cap                                  |
| QTF  | QuantiFERON-TB Gold Assay                         |
| QTG  | QFT-TB1 Ag Green Cap                              |
| QTY  | QFT-TB2 Ag Yellow Cap                             |
| RBE  | Royal blue - EDTA                                 |
| RBN  | Royal blue - no anticoagulant                     |
| RLS  | Ringer's Lactate Solution                         |
| RNL  | RNALater  |
| RNP  | RNAprotect  |
| ROC  | Roche Media                                       |
| RPM  | RPMI 1640 Medium                                  |
| RPS  | RNA Preservation Solution                         |
| SBC  | Sodium Bicarbonate                                |
| SCC  | Steck Cyto-Chex tubes - stabilized                |
| SCI  | Sodium Citrate                                    |
| SED  | Sodium EDTA                                       |
| SFE  | Sodium Fluoride/Na2EDTA                           |
| SFL  | Sodium Fluoride                                   |
| SIA  | Sulfite Iron Agar                                 |
| SKM  | Skim Milk   |
| SNP  | Snap/Flash Freeze                                 |
| SPH  | Sucrose Phosphate                                 |
| SPO  | Sodium Fluoride/Potassium Oxalate                 |
| SPS  | Sodium Polyanetholesulfonate                      |
| SST  | Serum Separator                                   |
| STG  | Skim milk-Tryptone-Glucose-Glycerin Medium (STGG) |
| TAL  | Blood Bag Tail                                    |
| TBD  | Specimen type to be defined                       |
| TBS  | Tris-Buffered Saline                              |
| TCD  | tween chlorhexidine digluconate                   |
| TEM  | Tempus Tube                                       |
| TFM  | Tissue Freezing Medium                            |

| Code | Description                           |
|------|---------------------------------------|
| TFX  | Tissue Fix                            |
| THM  | Thrombin                              |
| TMS  | Transport Medium-Stuarts              |
| TRC  | TruCulture Tube                       |
| TVT  | TransFix Vacuum Blood Collection Tube |
| UNK  | Unknown Additive                      |
| URM  | Urine Transport Media                 |
| UTM  | Universal Transport Media             |
| VTM  | Viral Transport Media                 |
| ZRD  | Zymo RNA/DNA shield                   |

# **Derivative codes**

This section lists the derivative codes available in LDMS.

| Code | Description                     |
|------|---------------------------------|
| A4N  | Activated CD4+ without Tregs    |
| AD4  | Activated CD4                   |
| AD8  | Activated CD8 Derivative        |
| ADG  | Adrenal Glands                  |
| ADL  | Adipose Tissue Layer            |
| ADP  | Adipose (abdominal)             |
| AHR  | Non-Occipital scalp hair        |
| AMN  | Amniotic Fluid                  |
| AOR  | Aorta                           |
| APL  | Applicator Tip                  |
| ASP  | Aspirate                        |
| BAC  | Bacterial Isolate               |
| BAL  | Bronchoalveolar Lavage          |
| BCL  | B-cell Lymphoblastoid Cell Line |
| BHR  | Hair, Body                      |
| BLA  | Bladder                         |
| BLC  | Blood Clots                     |
| BLD  | Blood (Whole - Venous)          |
| BLK  | Tissue Block                    |
| ВМА  | Bone Marrow Aspirate            |
| вмс  | Bone Marrow Core Biopsy         |
| ВМК  | Human Milk                      |

| Code | Description  |
|------|--|
| BML  | Human Milk - Left  |
| BMR  | Human Milk - Right   |
| BMS  | Human Milk - Spun  |
| BMW  | Human Milk - Whole   |
| BNM  | Bone Marrow  |
| BPS  | Biopsy   |
| BRN  | Brain Tissue   |
| BRS  | Breast Tissue  |
| BSG  | Basal Ganglia  |
| BUC  | Buccal   |
| BUF  | Unficolled Cryopreserved Buffy Coat, Viable                  |
| BUG  | Bulbourethral Glands   |
| CAN  | Candida  |
| CCC  | Cryopreserved Cells from a Culture, Viable                   |
| CD4  | CD4 Positive T-Cells   |
| CD8  | CD8 cells  |
| CDE  | Caudate  |
| CDN  | Copy DNA   |
| CDP  | Dry Pellet from a Culture, Non Viable                        |
| CEL  | PBMC Cells, Viable   |
| CEN  | Fresh Cells from a Non-Blood Spec. Type                      |
| CER  | Cervix   |
| CGN  | Cells in GITC  |
| CIO  | Cells in Other (Solution), Non-Viable                        |
| CLI  | Cell Lines   |
| CLL  | Colon (left)   |
| CLN  | Cryopreserved primary cells from Non-Blood Spec Type, Viable |
| CLR  | Colon (right)  |
| CLS  | Sigmoid Colon  |
| CMV  | CytoMegaloVirus Isolate                                      |
| CRB  | Cerebellum   |
| CRY  | Generic Cryptococcus   |
| CSF  | Cerebro-Spinal Fluid   |
| CSR  | Serum - Chilled  |
| СТВ  | Cytobrush  |

| Code | Description                                  |
|------|--|
| СТС  | Cells from a CTL Assay                       |
| CTS  | Supernatant generated from a CTL Assay       |
| CUP  | Cup  |
| CVL  | Cervical Vaginal Lavage                      |
| CVS  | CVL Supernatant                              |
| CXS  | Cervical Swab                                |
| DBE  | Dried Blood Extract                          |
| DBS  | Dried Blood Spot                             |
| DDM  | Duodenum                                     |
| DPE  | Plasma, Dried Extract                        |
| DPL  | Plasma, Dried                                |
| DPS  | Dried Plasma Spot                            |
| DSP  | Digested Sputum                              |
| DUR  | Dried Urine                                  |
| DWB  | Dried Whole Blood                            |
| ED4  | Effector Memory CD4 T-cells                  |
| END  | Endometrial Tissue                           |
| EPI  | Epididymus                                   |
| EPP  | Endodontic Paper Point                       |
| ESO  | Esophagus                                    |
| EYE  | Eye  |
| FCM  | Frontal Cortex (motor)                       |
| FCP  | Frontal Cortex (pre-motor)                   |
| FGN  | Finger Nails                                 |
| FHR  | Hair, Facial                                 |
| FLD  | Fluid Portion from a Non-Blood Specimen Type |
| FLS  | Flocked Swab                                 |
| FLT  | Fallopian Tubes                              |
| FPL  | Plasma, Filtered                             |
| FSI  | Penis Foreskin, Inner Tissue                 |
| FSO  | Penis Foreskin, Outer Tissue                 |
| FSR  | Rectal Biopsy by Flexible Sigmoidoscopy      |
| FTP  | Filter Paper                                 |
| GLS  | Glycerol Stock                               |
| GMI  | Gastro Intestinal Mucosa                     |

| Code | Description                                    |
|------|--|
| GRN  | Granulocytes                                   |
| HAR  | Hair   |
| HPC  | Hippocampus                                    |
| HRT  | Heart  |
| ICK  | Supernatant generated from Inducible Cytokines |
| ILM  | Ileum  |
| IPK  | Methanol Extract Supernatant                   |
| IPT  | Immunophenotyping                              |
| IVR  | Intra vaginal Ring                             |
| IVS  | Intraventricular Septum                        |
| MCC  | Jejunum  |
| KDY  | Kidney   |
| LAT  | Left Atrium                                    |
| LAV  | Lavage   |
| LDB  | Leukodepleted blood                            |
| LIV  | Liver Tissue                                   |
| LNA  | Lymph Node (aortic)                            |
| LNC  | Lymph Node (cervical)                          |
| LNE  | Pre-auricular Lymph Node                       |
| LNF  | Infraclavicular Lymph Node                     |
| LNG  | Lung   |
| LNI  | Inguinal Lymph Node                            |
| LNK  | Popliteal Lymph Node                           |
| LNM  | Lymph Node (mediastinal)                       |
| LNO  | Lymph Node (other)                             |
| LNP  | Lymph Node (per-hilar)                         |
| LNR  | Para-aortic Lymph Node                         |
| LNS  | Supraclavicular Lymph Node                     |
| LNT  | Mesenteric Lymph Node                          |
| LNX  | Lymph Node (axillary)                          |
| LPD  | Lipid Layer                                    |
| LPK  | Leukopak                                       |
| LVE  | Left Ventricle                                 |
| LYM  | Lymph Node Biopsy/Aspirate                     |
| LYS  | Lysed whole blood                              |

| Code | Description   |
|------|---|
| MBR  | Midbrain Consists of Cerebral Penduncles and Tectum |
| MCL  | Macrophage Cells - Viable                           |
| MCS  | Microbiology Culture Slant                          |
| MD4  | Central Memory CD4 T-cells                          |
| MDC  | Myeloid Dendritic Cells                             |
| MEC  | Meconium  |
| MED  | Medulla   |
| MNO  | Monocytes   |
| MPE  | Macrophage Dried Cell Pellet, Non Viable            |
| МТВ  | MTB Isolates  |
| MUC  | Mucins  |
| MUS  | Mucous  |
| N/A  | Not Applicable - Same as Primary Specimen Type      |
| NA4  | Non-activated CD4                                   |
| NA8  | Non-activated CD8                                   |
| NCL  | Neutrophil (PMN) - Viable cells                     |
| ND4  | Naive CD4 T-cells                                   |
| NKC  | Natural Killer Cells                                |
| NON  | None  |
| NPE  | Neutrophil (PMN) Dried Cell Pellet, Non Viable      |
| NPW  | Nasopharyngeal Wash                                 |
| NTM  | Nontuberculous mycobacteria                         |
| NXD  | Non-extracted DNA                                   |
| occ  | Occipital Cortex                                    |
| OHR  | Occipital scalp hair                                |
| OPC  | Ova & Parasite Concentrate                          |
| OTH  | Other   |
| OVR  | Ovaries   |
| PAD  | Adipose (pericardial)                               |
| PCC  | Culture Supernatant and Cells                       |
| PCF  | Pericardial Fluid                                   |
| PCT  | Parietal Cortex                                     |
| PDC  | Plasmacytoid Dendritic Cells                        |
| PED  | Dried Pellet - Digene                               |
| PEL  | Non-viable PBMC s                                   |

| Code | Description                                     |
|------|---|
| PEN  | Non-viable cells from non-blood specimen type   |
| PEO  | Wet Pellet Prep - Organon Teknika               |
| PER  | Dried Pellet - Roche                            |
| PFR  | Parotid Flow                                    |
| PHR  | Hair, Pubic                                     |
| PL*  | Plasma, All                                     |
| PL1  | Plasma, Single-Spun                             |
| PL2  | Plasma, Double-Spun                             |
| PLA  | Plasma, Unspecified/Other                       |
| PLC  | Placental Tissue                                |
| PLH  | Plasma High Spin                                |
| PLM  | Placenta - Membrane Slice                       |
| PLP  | Supernatant & Cells from a Quantitative Culture |
| PLQ  | Plaque  |
| PLT  | Placenta - Placenta Slice                       |
| PMD  | Plasmids  |
| PMN  | polymorphnuclear leukocytes                     |
| PON  | Pons  |
| PNC  | Pancreas  |
| PPK  | Plasmapak                                       |
| PPS  | Periopaper Strip                                |
| PRO  | Prostate  |
| PTF  | Peritoneal Fluid                                |
| PTY  | Parathyroid                                     |
| RAT  | Right Atrium                                    |
| RBC  | Red Blood Cells                                 |
| RCT  | Rectum  |
| RD4  | Resting CD4 T-cells                             |
| REC  | Rectal  |
| RVE  | Right Ventricle                                 |
| SAL  | Saliva  |
| SCC  | Spinal Cord Cervical                            |
| SCL  | Spinal Cord Lumbosacral                         |
| SCP  | Scraping  |
| SCT  | Spinal Cord Thoracic                            |

| Code | Description                              |
|------|--|
| SDI  | Supernatant Dermis, Inner                |
| SDO  | Supernatant Dermis, Outer                |
| SEC  | Secretions                               |
| SEI  | Supernatant Epidermis, Inner             |
| SEM  | Semen                                    |
| SEO  | Supernatant Epidermis, Outer             |
| SER  | Serum                                    |
| SHR  | Hair, Scalp                              |
| SKN  | Skin                                     |
| SLD  | Slide from a primary sample              |
| SMD  | Skeletal Muscle (distal)                 |
| SMP  | Skeletal Muscle (proximal)               |
| SMR  | Smear                                    |
| SMV  | Seminal Vesicle                          |
| SNO  | SNO - Strip                              |
| SPG  | Sponge                                   |
| SPI  | Unfractioned Sputum, Induced             |
| SPL  | Spleen                                   |
| SPQ  | Supernatant from a Quantitative Culture  |
| SPT  | Unfractioned Sputum                      |
| SRH  | Serum - High Speed Spun                  |
| STK  | Quantitative Culture Held beyond 14 Days |
| STL  | Stool                                    |
| STM  | Stomach                                  |
| STP  | Sterile Tooth Pick                       |
| SUP  | Culture Supernatant                      |
| SVG  | salivary glands                          |
| SWB  | Swab                                     |
| SWS  | Stimulated Whole Saliva                  |
| TBD  | Specimen type to be defined              |
| TCX  | Temporal Cortex                          |
| TD4  | Transitional Memory CD4 T-cells          |
| TFS  | Tear-Flo Strips                          |
| THL  | Thalamus                                 |
| THW  | Throat Wash                              |

| Code | Description   |
|------|---|
| THY  | Thymus  |
| TIS  | Tissue  |
| TNA  | Toe Nails   |
| TON  | Tonsillar Biopsy/Aspirate   |
| TR4  | T regs CD4+   |
| TRC  | Trucount  |
| TRN  | Transverse  |
| TST  | Testes  |
| TTH  | Tooth   |
| TUM  | Tumor   |
| TYD  | Thyroid   |
| UMB  | Placenta - Cord Slice   |
| UNK  | Unknown Derivative  |
| URN  | Urine   |
| UTS  | Uterus  |
| UWS  | Unstimulated Whole Saliva   |
| VAG  | Vaginal   |
| VCP  | Vaginal Cup   |
| VGN  | Vagina  |
| VTM  | Viral Transport Media   |
| W/D  | Wet/Dry   |
| WBP  | Whole Blood Pellet, Specify Methodology                                   |
| WEK  | Wick/Wek Cell Sponge  |
| WFC  | Unstimulated whole blood cell, lysed , fixed, frozen (with DMSO) for flow |
| WFS  | Stimulated whole blood cells, lysed, fixed, frozen (with DMSO) for flow   |
| WPK  | Whole Blood Packed  |
| XDA  | Extracted DNA   |
| XFL  | Extracted Fluid from RNA RT PCR for Sequencing                            |
| XPA  | Extracted RNA   |
| XPD  | Extracted Pellet from DNA PCR for Sequencing                              |
| XPR  | Extracted Pellet from RNA RT PCR for Sequencing                           |
| XTN  | Extracted total nucleic acid  |

# Sub additive/derivative codes

This section lists the sub additive/derivative codes available in LDMS.

| 2 o'clock biopsy position  33 o'clock biopsy position  04C   | Code | Description                 |
|--|------|-----------------------------|
| 3 o'clock biopsy position 04C  | 01C  | 1 o'clock biopsy position   |
| 04C 4 o'clock biopsy position 05C 5 o'clock biopsy position 06C 6 o'clock biopsy position 07C 7 o'clock biopsy position 08C 8 o'clock biopsy position 08C 8 o'clock biopsy position 09C 9 o'clock biopsy position 10C 10 o'clock biopsy position 11C 11 o'clock biopsy position 11C 12 c'clock biopsy position 11C 12 o'clock biopsy position 11D Tooth #14 Distal Buccal 14M Tooth #14 Mesial Buccal 14M Tooth #19 Distal Buccal 15D Tooth #19 Distal Buccal 16D Tooth #19 Mesial Buccal 17D Tooth #19 Mesial Buccal 18D Tooth #25 Distal Buccal 18D Tooth #30 Distal Buccal 18D Tooth #30 Distal Buccal 18D Tooth #30 Distal Buccal 18D Tooth #3 Mesial Meccal 1 | 02C  | 2 o'clock biopsy position   |
| 50 o'clock biopsy position  06C 6 o'clock biopsy position  07C 7 o'clock biopsy position  08C 8 o'clock biopsy position  09C 9 o'clock biopsy position  10C 10 o'clock biopsy position  11C 11 o'clock biopsy position  11C 12 o'clock biopsy position  12C 12 o'clock biopsy position  14D Tooth #14 Distal Buccal  14M Tooth #14 Mesial Buccal  19D Tooth #19 Distal Buccal  19M Tooth #19 Mesial Buccal  24M Tooth #24 Mesial Buccal  25D Tooth #25 Distal Buccal  30D Tooth #30 Distal Buccal  30D Tooth #30 Distal Buccal  30B Tooth #3 Distal Buccal  30B Tooth #3 Distal Buccal  30B Tooth #3 Mesial Buccal  30B Tooth #3 Freezing Solution  30B Tooth #3 Distal Buccal  30B Tooth #3 Freezing Solution  30B Alsever's Freezing Solution  30B Alsever Vial  30B Antigen   | 03C  | 3 o'clock biopsy position   |
| 06C 6 o'clock biopsy position 07C 7 o'clock biopsy position 08C 8 o'clock biopsy position 09C 9 o'clock biopsy position 10C 10 o'clock biopsy position 11C 11 o'clock biopsy position 11C 12 o'clock biopsy position 11C 12 o'clock biopsy position 12C 12 o'clock biopsy position 14D Tooth #14 Distal Buccal 14M Tooth #14 Distal Buccal 19M Tooth #19 Distal Buccal 19M Tooth #19 Mesial Buccal 24M Tooth #25 Distal Buccal 25D Tooth #25 Distal Buccal 30D Tooth #30 Distal Buccal 30D Tooth #30 Distal Buccal 30B Tooth #3 Distal Buccal 30B Tooth #3 Mesial Buccal 30B Tooth #4 Mesial Buccal 30B Tooth #5 Distal Buccal 30B Tooth #6 Mesial Buccal 30B Tooth #6 Mesial Buccal 30B Tooth #7 Distal Buccal 30B Tooth #8 Mesial Buccal 30B Tooth #8 Mesial Buccal 30B Tooth #8 Freezing Solution 30B Tooth #6 Solution 3 | 04C  | 4 o'clock biopsy position   |
| 07C 7 o'clock biopsy position 08C 8 o'clock biopsy position 09C 9 o'clock biopsy position 10C 10 o'clock biopsy position 11C 11 o'clock biopsy position 11C 11 o'clock biopsy position 12C 12 o'clock biopsy position 14D Tooth #14 Distal Buccal 14M Tooth #14 Mesial Buccal 19D Tooth #19 Distal Buccal 19M Tooth #19 Mesial Buccal 24M Tooth #24 Mesial Buccal 25D Tooth #25 Distal Buccal 30D Tooth #30 Distal Buccal 30D Tooth #30 Distal Buccal 30B Tooth #3 Mesial Buccal 30B Tooth #3 Mesial Buccal 30C K562 Cell Line 8MB Tooth #8 Mesial Buccal 4CD Acid Citrate Dextrose ADA Adenosine Deaminase ADJ Adjuvant AEB Qiagen AE Buffer AFS Alsever's Freezing Solution AG1 ESAT 6-free Cocktail 1 AG2 ESAT 6-free Cherry pick AMB Amber Vial ANT Antigen   | 05C  | 5 o'clock biopsy position   |
| 8 o'clock biopsy position 99C 9 o'clock biopsy position 10C 10 o'clock biopsy position 11C 11 o'clock biopsy position 11C 11 o'clock biopsy position 12C 12 o'clock biopsy position 14D Tooth #14 Distal Buccal 14M Tooth #19 Distal Buccal 19D Tooth #19 Mesial Buccal 19M Tooth #19 Mesial Buccal 24M Tooth #24 Mesial Buccal 25D Tooth #25 Distal Buccal 30D Tooth #30 Distal Buccal 30D Tooth #30 Distal Buccal 30B Tooth #3 Mesial Buccal 30B Aiducal 30B Tooth #3 Mesial Buccal 30B Tooth #5 Mesial Buccal 30B Tooth  | 06C  | 6 o'clock biopsy position   |
| 9 o'clock biopsy position 10C 10 o'clock biopsy position 11C 11 o'clock biopsy position 11C 11 o'clock biopsy position 12C 12 o'clock biopsy position 14D Tooth #14 Distal Buccal 14M Tooth #14 Mesial Buccal 19D Tooth #19 Distal Buccal 19M Tooth #19 Mesial Buccal 24M Tooth #24 Mesial Buccal 25D Tooth #25 Distal Buccal 30D Tooth #30 Distal Buccal 30D Tooth #30 Distal Buccal 30B Tooth #3 Mesial Buccal 30B Tooth #3 Mesial Buccal 30B Tooth #3 Mesial Buccal 30B Tooth #3 Pistal Buccal 30B Tooth #3 Mesial Buccal 30B Tooth #3 Adesial Buccal 30B Tooth #3 Mesial Bucc | 07C  | 7 o'clock biopsy position   |
| 10C 10 o'clock biopsy position 11C 11 o'clock biopsy position 12C 12 o'clock biopsy position 14D Tooth #14 Distal Buccal 14M Tooth #14 Mesial Buccal 19D Tooth #19 Distal Buccal 19M Tooth #19 Mesial Buccal 24M Tooth #24 Mesial Buccal 25D Tooth #25 Distal Buccal 30D Tooth #30 Distal Buccal 30D Tooth #30 Mesial Buccal 30B Tooth #3 Distal Buccal 30B Tooth #3 Mesial Buccal 3 | 08C  | 8 o'clock biopsy position   |
| 11C 11 o'clock biopsy position 12C 12 o'clock biopsy position 14D Tooth #14 Distal Buccal 14M Tooth #14 Mesial Buccal 19D Tooth #19 Distal Buccal 19M Tooth #19 Mesial Buccal 24M Tooth #24 Mesial Buccal 25D Tooth #25 Distal Buccal 30D Tooth #30 Distal Buccal 30D Tooth #30 Distal Buccal 30B Tooth #3 Distal Buccal 30B Tooth #3 Mesial Buccal 30B Tooth #4 Mesial Buccal 30B Tooth #5 Mesial Buccal 30B Tooth #8 Mesial Buccal 30B Tooth #3 Mesial Buccal 30B T | 09C  | 9 o'clock biopsy position   |
| 12C 12 o'clock biopsy position  14D Tooth #14 Distal Buccal  14M Tooth #14 Mesial Buccal  19D Tooth #19 Distal Buccal  19M Tooth #19 Mesial Buccal  24M Tooth #24 Mesial Buccal  25D Tooth #25 Distal Buccal  30D Tooth #30 Distal Buccal  30D Tooth #30 Distal Buccal  30M Tooth #3 Mesial Buccal  30B Tooth #3 Mesial Buccal  4562 K562 Cell Line  8MB Tooth #8 Mesial Buccal  9DB Tooth #9 Distal Buccal  ACD Acid Citrate Dextrose  ADA Adenosine Deaminase  ADJ Adjuvant  AEB Qiagen AE Buffer  AFS Alsever's Freezing Solution  AG1 ESAT 6-free Cocktail 1  AG2 ESAT 6-free Cherry pick  AMB Amber Vial  ANT Antigen   | 10C  | 10 o'clock biopsy position  |
| 14D Tooth #14 Distal Buccal 14M Tooth #14 Mesial Buccal 19D Tooth #19 Distal Buccal 19M Tooth #19 Mesial Buccal 24M Tooth #24 Mesial Buccal 25D Tooth #25 Distal Buccal 30D Tooth #30 Distal Buccal 30D Tooth #30 Distal Buccal 30B Tooth #3 Distal Buccal 30B Tooth #3 Distal Buccal 30B Tooth #3 Mesial Buccal 30B Tooth #40 Distal Buccal 30B Tooth #8 Mesial Buccal 30B Tooth #3 Distal Buccal 30B Tooth #3 | 11C  | 11 o'clock biopsy position  |
| 14M Tooth #14 Mesial Buccal 19D Tooth #19 Distal Buccal 19M Tooth #19 Mesial Buccal 24M Tooth #24 Mesial Buccal 25D Tooth #25 Distal Buccal 30D Tooth #30 Distal Buccal 30D Tooth #30 Distal Buccal 30B Tooth #3 Mesial Buccal 30B Tooth #4 Mesial Buccal 30B Tooth #4 Mesial Buccal 30B Tooth #5 Distal Buccal 30B Tooth #8 Mesial Buccal 30B Tooth #3 Distal Buccal 30B Tooth #3 D | 12C  | 12 o'clock biopsy position  |
| Tooth #19 Distal Buccal  Tooth #19 Mesial Buccal  Tooth #24 Mesial Buccal  Tooth #25 Distal Buccal  Tooth #30 Distal Buccal  Tooth #30 Mesial Buccal  Tooth #30 Mesial Buccal  Tooth #30 Distal Buccal  Tooth #3 Distal Buccal  Tooth #3 Mesial Buccal  Tooth #3 Mesial Buccal  K562 K562 Cell Line  MB Tooth #8 Mesial Buccal  Tooth #9 Distal Buccal  ACD Acid Citrate Dextrose  ADA Adenosine Deaminase  ADJ Adjuvant  AEB Qiagen AE Buffer  AFS Alsever's Freezing Solution  AG1 ESAT 6-free Cocktail 1  AG2 ESAT 6-free Cherry pick  AMB Amber Vial  ANT Antigen  | 14D  | Tooth #14 Distal Buccal     |
| Tooth #19 Mesial Buccal  24M Tooth #24 Mesial Buccal  25D Tooth #25 Distal Buccal  30D Tooth #30 Distal Buccal  30M Tooth #30Mesial Buccal  30B Tooth #3 Distal Buccal  30B Tooth #3 Mesial Buccal  562 K562 Cell Line  8MB Tooth #8 Mesial Buccal  9DB Tooth #9 Distal Buccal  ACD Acid Citrate Dextrose  ADA Adenosine Deaminase  ADJ Adjuvant  AEB Qiagen AE Buffer  AFS Alsever's Freezing Solution  AG1 ESAT 6-free cocktail 1  AG2 ESAT 6-free Cherry pick  AMB Amber Vial  ANT Antigen  | 14M  | Tooth #14 Mesial Buccal     |
| Tooth #24 Mesial Buccal  Tooth #25 Distal Buccal  Tooth #30 Distal Buccal  Tooth #3 Distal Buccal  Tooth #3 Distal Buccal  Tooth #3 Distal Buccal  Tooth #3 Mesial Buccal  K562 Cell Line  K562 Cell Line  Tooth #8 Mesial Buccal  Tooth #9 Distal Buccal  ACD Acid Citrate Dextrose  ADA Adenosine Deaminase  ADJ Adjuvant  AEB Qiagen AE Buffer  AFS Alsever's Freezing Solution  AG1 ESAT 6-free Cocktail 1  AG2 ESAT 6-free Cherry pick  AMB Amber Vial  ANT Antigen   | 19D  | Tooth #19 Distal Buccal     |
| Tooth #25 Distal Buccal  30D Tooth #30 Distal Buccal  30M Tooth #30Mesial Buccal  30B Tooth #3 Distal Buccal  30B Tooth #3 Mesial Buccal  30B Tooth #3 Mesial Buccal  562 K562 Cell Line  8MB Tooth #8 Mesial Buccal  9DB Tooth #9 Distal Buccal  ACD Acid Citrate Dextrose  ADA Adenosine Deaminase  ADJ Adjuvant  AEB Qiagen AE Buffer  AFS Alsever's Freezing Solution  AG1 ESAT 6-free cocktail 1  AG2 ESAT 6-free Cherry pick  AMB Amber Vial  ANT Antigen  | 19M  | Tooth #19 Mesial Buccal     |
| Tooth #30 Distal Buccal  30M Tooth #30Mesial Buccal  3DB Tooth #3 Distal Buccal  3MB Tooth #3 Mesial Buccal  562 K562 Cell Line  8MB Tooth #8 Mesial Buccal  9DB Tooth #9 Distal Buccal  ACD Acid Citrate Dextrose  ADA Adenosine Deaminase  ADJ Adjuvant  AEB Qiagen AE Buffer  AFS Alsever's Freezing Solution  AG1 ESAT 6-free cocktail 1  AG2 ESAT 6-free Cherry pick  AMB Amber Vial  ANT Antigen   | 24M  | Tooth #24 Mesial Buccal     |
| Tooth #30Mesial Buccal  3DB Tooth #3 Distal Buccal  3MB Tooth #3 Mesial Buccal  562 K562 Cell Line  8MB Tooth #8 Mesial Buccal  9DB Tooth #9 Distal Buccal  ACD Acid Citrate Dextrose  ADA Adenosine Deaminase  ADJ Adjuvant  AEB Qiagen AE Buffer  AFS Alsever's Freezing Solution  AG1 ESAT 6-free cocktail 1  AG2 ESAT 6-free Cherry pick  AMB Amber Vial  ANT Antigen  | 25D  | Tooth #25 Distal Buccal     |
| 3DB Tooth #3 Distal Buccal  3MB Tooth #3 Mesial Buccal  562 K562 Cell Line  8MB Tooth #8 Mesial Buccal  9DB Tooth #9 Distal Buccal  ACD Acid Citrate Dextrose  ADA Adenosine Deaminase  ADJ Adjuvant  AEB Qiagen AE Buffer  AFS Alsever's Freezing Solution  AG1 ESAT 6-free cocktail 1  AG2 ESAT 6-free Cherry pick  AMB Amber Vial  ANT Antigen  | 30D  | Tooth #30 Distal Buccal     |
| 3MB Tooth #3 Mesial Buccal  562 K562 Cell Line  8MB Tooth #8 Mesial Buccal  9DB Tooth #9 Distal Buccal  ACD Acid Citrate Dextrose  ADA Adenosine Deaminase  ADJ Adjuvant  AEB Qiagen AE Buffer  AFS Alsever's Freezing Solution  AG1 ESAT 6-free cocktail 1  AG2 ESAT 6-free Cherry pick  AMB Amber Vial  ANT Antigen  | 30M  | Tooth #30Mesial Buccal      |
| K562 Cell Line  K562 Cell Line | 3DB  | Tooth #3 Distal Buccal      |
| 8MB Tooth #8 Mesial Buccal  9DB Tooth #9 Distal Buccal  ACD Acid Citrate Dextrose  ADA Adenosine Deaminase  ADJ Adjuvant  AEB Qiagen AE Buffer  AFS Alsever's Freezing Solution  AG1 ESAT 6-free cocktail 1  AG2 ESAT 6-free Cherry pick  AMB Amber Vial  ANT Antigen  | 3МВ  | Tooth #3 Mesial Buccal      |
| 9DB Tooth #9 Distal Buccal  ACD Acid Citrate Dextrose  ADA Adenosine Deaminase  ADJ Adjuvant  AEB Qiagen AE Buffer  AFS Alsever's Freezing Solution  AG1 ESAT 6-free cocktail 1  AG2 ESAT 6-free Cherry pick  AMB Amber Vial  ANT Antigen  | 562  | K562 Cell Line              |
| ACD Acid Citrate Dextrose  ADA Adenosine Deaminase  ADJ Adjuvant  AEB Qiagen AE Buffer  AFS Alsever's Freezing Solution  AG1 ESAT 6-free cocktail 1  AG2 ESAT 6-free Cherry pick  AMB Amber Vial  ANT Antigen  | 8MB  | Tooth #8 Mesial Buccal      |
| ADA Adenosine Deaminase  ADJ Adjuvant  AEB Qiagen AE Buffer  AFS Alsever's Freezing Solution  AG1 ESAT 6-free cocktail 1  AG2 ESAT 6-free Cherry pick  AMB Amber Vial  ANT Antigen   | 9DB  | Tooth #9 Distal Buccal      |
| ADJ Adjuvant  AEB Qiagen AE Buffer  AFS Alsever's Freezing Solution  AG1 ESAT 6-free cocktail 1  AG2 ESAT 6-free Cherry pick  AMB Amber Vial  ANT Antigen  | ACD  | Acid Citrate Dextrose       |
| AEB Qiagen AE Buffer  AFS Alsever's Freezing Solution  AG1 ESAT 6-free cocktail 1  AG2 ESAT 6-free Cherry pick  AMB Amber Vial  ANT Antigen  | ADA  | Adenosine Deaminase         |
| AFS Alsever's Freezing Solution  AG1 ESAT 6-free cocktail 1  AG2 ESAT 6-free Cherry pick  AMB Amber Vial  ANT Antigen  | ADJ  | Adjuvant                    |
| AG1 ESAT 6-free cocktail 1  AG2 ESAT 6-free Cherry pick  AMB Amber Vial  ANT Antigen   | AEB  | Qiagen AE Buffer            |
| AG2 ESAT 6-free Cherry pick  AMB Amber Vial  ANT Antigen   | AFS  | Alsever's Freezing Solution |
| AMB Amber Vial ANT Antigen   | AG1  | ESAT 6-free cocktail 1      |
| ANT Antigen  | AG2  | ESAT 6-free Cherry pick     |
|  | AMB  | Amber Vial                  |
| APR Aprotinin  | ANT  | Antigen                     |
|  | APR  | Aprotinin                   |

| Code | Description                                  |
|------|--|
| ASC  | Ascorbate (Vit C)                            |
| ATC  | Animal Tumor Cell Line                       |
| ATG  | TB Antigen                                   |
| AUO  | Auramine O                                   |
| AVE  | Qiagen Buffer AVE                            |
| BCG  | Bacille Calmette-Guïċ½rin                    |
| BCL  | B-cell Lymphoblastoid Cell Line              |
| BFM  | Bacterial Freezing Media                     |
| ВНТ  | Butylated hydroxytoluene                     |
| BLK  | Tissue Block                                 |
| BML  | Breast - Left                                |
| BMR  | Breast - Right                               |
| BSA  | Bovine Serum Albumin                         |
| BSS  | Blood Stabilizing Solution                   |
| ВТМ  | Biopsy Transfer Media                        |
| CAN  | Candida (CASTA)                              |
| CBS  | Cellbanker Solution                          |
| ССМ  | CareHPV Collection Medium                    |
| CD4  | CD4 cells                                    |
| CD8  | CD8 cells                                    |
| CDN  | cDNA   |
| CFM  | Cell Freezing Media                          |
| CME  | Culture Media                                |
| CMV  | CytoMegaloVirus                              |
| CON  | Control                                      |
| CSC  | Cervical secretions                          |
| CSP  | Cytospin Slides                              |
| CVA  | Complete Vaccine                             |
| CYT  | Cytokines                                    |
| DCF  | 2 - deoxycoformycin.                         |
| DEA  | DEACTIVATED                                  |
| DMS  | DMSO used in cryopreservation solution (CPS) |
| DOD  | Day of Donation                              |
| DOT  | Day of Tail                                  |
| DPH  | Wyeth Ayerst Diphtheria Antigen              |

| Code | Description                                |
|------|--|
| DPL  | Depleted Cell Populations                  |
| DTM  | Digene/Qiagen Specimen Transport Medium    |
| DTT  | Dithiothreitol                             |
| EDT  | EDTA                                       |
| ETH  | Ethanol                                    |
| FBS  | Fetal Bovine Serum                         |
| FMA  | Formic Acid                                |
| FMD  | Formaldehyde                               |
| FOR  | Formalin                                   |
| FPR  | Filter Paper                               |
| FRS  | First Spin                                 |
| FRZ  | Frozen                                     |
| FUN  | Fungal Broth                               |
| GAG  | HIV GAG envelope peptide                   |
| GIT  | Guanidine Isothiocyanate (GITC)            |
| GLM  | Glycerol Media                             |
| GLT  | Glutaraldehyde                             |
| GLY  | 20% Glycerol                               |
| GMS  | Giemsa Staining                            |
| GRS  | Gram Stain                                 |
| H&E  | Hematoxylin and Eosin stain                |
| H2O  | Water                                      |
| HBS  | Hank's Balanced Salt Solution              |
| HCI  | Hydrochloric Acid                          |
| HPA  | Нер А                                      |
| HPS  | Human Pappilloma Virus Serology            |
| HPV  | Human Pappilloma Virus                     |
| HTC  | Human Tumor Cell Line                      |
| I18  | IL-18                                      |
| I1b  | IL-1b                                      |
| IAP  | Iodoacetic Acid/Phenanthrolene             |
| IgG  | Immunoglobulin G                           |
| IL2  | IL-2, Interleukin 2                        |
| IL6  | Interleukin6                               |
| ILA  | IL-12/IL-15, Interleukin-12/Interleukin-15 |

| Code | Description  |
|------|--|
| INK  | India Ink  |
| IPD  | PBMCs for Intact Proviral DNA testing                      |
| LAA  | L-ascorbic Acid  |
| LES  | Lesion   |
| LFT  | Left   |
| LPS  | Lipopolysaccaride  |
| LSM  | Lymphocyte Separation Media                                |
| LVT  | L-var Target   |
| LYB  | Lysis Buffer   |
| M2V  | M2VP   |
| MA1  | MAC 101  |
| MAC  | MAC Antigen  |
| MAF  | MAC LR114F Culture Filtrate                                |
| MAS  | M. Avium Sensitin  |
| MCH  | Mechanical Homogenization                                  |
| MDB  | Middlebrook broth  |
| MET  | Methanol   |
| MFS  | Malaria Freezing Solution                                  |
| MIF  | Merthoilate Formalin/Merthiolate-Iodine Formalin (MF/MIF)  |
| MIT  | Quantiferon Gold and/or Quantiferon Gold Plus Mitogen tube |
| MPP  | For a mega peptide pool                                    |
| MRN  | Messenger RNA  |
| MT2  | MT2 Cell Line  |
| MTV  | For an MTB vaccine   |
| MUC  | Mucins   |
| N/A  | Not Applicable   |
| NAS  | Nucleic Acid Stabilization Solution                        |
| NAT  | Sodium thioglycolate                                       |
| NCL  | N-acetyl-L-cysteine  |
| NDN  | NDVNS1, Newcastle disease virus with NS1 influenza protein |
| NDV  | NDVB1, Newcastle disease virus wild type                   |
| NDW  | Newcastle disease virus with W influenza protein           |
| NIL  | Quantiferon Gold and/or Quantiferon Gold Plus Nil tube     |
| NKC  | Natural Killer cells                                       |
| NOH  | Sodium Hydroxide   |

| Code | Description                                      |
|------|--|
| NON  | None   |
| NPI  | Neuropepsidase Inhibitor                         |
| NSK  | New Skin Lesion                                  |
| NSL  | Normal Saline                                    |
| NUC  | Nucleotides                                      |
| OCT  | Optimum Cutting Temperature Medium               |
| OKT  | OKT3 Treated                                     |
| ORI  | Oricol   |
| OTH  | Other  |
| P24  | P24  |
| PAE  | Pseudomonas aeruginosa                           |
| PAR  | Parrafin   |
| PBS  | Phosphate Buffered Saline                        |
| PCS  | PreserveCytSolution                              |
| PEI  | Pre-Infusion                                     |
| PFM  | Paraformaldahyde                                 |
| PHA  | PHA-Treated                                      |
| PI1  | S8820 Sigma SIGMAFAST Protease Inhibitor Tablets |
| PIC  | Protease Inhibitor Cocktail                      |
| PLC  | Placenta Tissue                                  |
| PLD  | Plasma, Depleted                                 |
| PLM  | Placenta Membrance                               |
| PMI  | Phorbol Myristate Acetate-Ionomycin              |
| PPA  | Phosphoric Acid                                  |
| PRS  | Protamine Sulfate                                |
| PTI  | Post-Infusion                                    |
| PTK  | Proteinase-K                                     |
| PTY  | Parathyroid                                      |
| PVA  | Polyvinyl alcohol                                |
| PWM  | Pokeweed Mitogen                                 |
| QAG  | QFT-TB Ag Red Cap                                |
| QMT  | QFT-Mitogen Purple Cap                           |
| QNL  | QFT-Nil Grey Cap                                 |
| QTG  | QFT-TB1 Ag Green Cap                             |
| QTY  | QFT-TB2 Ag Yellow Cap                            |

| Code | Description   |
|------|---|
| RDS  | RNA/DNA shield  |
| RFW  | RNAse free water  |
| RGT  | Right   |
| RLT  | Buffer RLT Plus   |
| RNL  | RNAlater  |
| RNP  | RNA Protect   |
| RPM  | RPMI 1640 Medium  |
| RTB  | Reverse Transcriptase buffer                              |
| S52  | Sendai 52 virus with defective infectious particules      |
| SAU  | Staph aureus  |
| SBC  | Sodium Bicarbonate  |
| SCB  | Sodium Carbonate  |
| SDS  | Second Spin   |
| SEB  | Sigma Staphylococcus Enterotoxin B                        |
| SLD  | Slide from a primary sample                               |
| SNP  | Snap/Flash Freeze   |
| SPG  | Sponge  |
| SPN  | Streptococcus pneumoniae                                  |
| SPS  | SMART TUBE INC Proteomic Stabilizer                       |
| STG  | Skim milk-Tryptone-Glucose-Glycerin Medium (STGG)         |
| STM  | Stimulated by Multiple Antigens                           |
| SUB  | Sub Gingival Plaque                                       |
| SUG  | Supragingival Plaque                                      |
| SUP  | Cultural Supernatant                                      |
| TB4  | Quantiferon Gold and/or Quantiferon Gold Plus CD4 tube    |
| TB8  | Quantiferon Gold Plus (only) CD8 tube                     |
| TBD  | Specimen type to be defined                               |
| TBS  | Tris-Buffered Saline                                      |
| TCA  | 1% Thimersol/ 6.6% EACA (Epsilon Amino Caproic Acid)      |
| TCL  | T-cell Lymphoblastoid Cell Line                           |
| TCM  | Trichrome stain   |
| TEB  | Tris-EDTA Buffer  |
| TEM  | Tempus Tube   |
| TFM  | Tissue Freezing Medium                                    |
| TMD  | Myeloid dendritic cell non infected and treated with TSST |

| Code | Description   |
|------|---|
| TMS  | Transport Medium-Stuarts  |
| TND  | Myeloid dendritic cell infected with NDVW and treated with TSST |
| TNN  | TSST with NDVNS1  |
| TNV  | TSST with NDVB1   |
| TOC  | Tocopherol (Vit E)  |
| ТРВ  | Trypan Blue   |
| TRI  | TriReagent  |
| TRX  | Triton X  |
| TRZ  | Trizol  |
| TSC  | Myeloid dendritic cell infect w/ Sendai Cantell & treat w/ TSST |
| TSS  | TSST, Toxic-Shock Syndrome Toxin                                |
| TTX  | Tetanus Toxoid treated  |
| UCT  | Urine cartridge   |
| ULQ  | Upper Left Quadrant   |
| UMB  | Placenta cord slice   |
| UNK  | Unknown Sub A/D   |
| UNT  | Untreated / Unstimulated  |
| URE  | Urea  |
| URQ  | Upper Right Quadrant  |
| VCS  | Cervicovaginal Secretions                                       |
| VIB  | Veal Infusion Broth   |
| VIR  | Virus   |
| VPS  | VP Stimulator   |
| VPT  | VP Target   |
| VTM  | Viral Transport Media   |
| ZLN  | Ziehl Neelsen   |

### **Condition codes**

This section lists the specimen condition codes available in LDMS.

| Code | Description             |
|------|-------------------------|
| ANM  | Sample Anonymized       |
| ANP  | Aliquot not prepared    |
| BKV  | Broken or cracked vial  |
| BLD  | Bloody                  |
| CDT  | Consumed During Testing |

| Code | Description   |
|------|---|
| CLT  | Clotted   |
| СМВ  | Combined Aliquots   |
| COC  | Combination of Conditions (Explain)                         |
| СТМ  | Contaminated  |
| DCF  | Data Clarification Form                                     |
| DCG  | Discharge present   |
| DFB  | Difficult Bleed   |
| DIM  | Dry Ice Melted  |
| DMG  | Damaged container   |
| DSH  | Delayed Shipment  |
| DSR  | Destroyed   |
| EQF  | Equipment Failure   |
| EXP  | Expired   |
| FRO  | Frozen  |
| HEM  | Hemolyzed   |
| HUM  | Exposed to high humidity                                    |
| ICT  | Icteric (Excess Bilirubin)                                  |
| INT  | Incorrect Tube  |
| INV  | Invalid Specimen  |
| LBE  | Lab Error   |
| LIP  | Lipemic   |
| LKD  | Leaked  |
| LLT  | Local Lab Testing   |
| LSH  | Lost Shipment   |
| LYS  | Lysed   |
| NOT  | No Test Performed   |
| NQA  | RT QA for viability and recovery NOT done due to low volume |
| NST  | Did Not Consent to Storage                                  |
| OPR  | Sample drawn outside protocol requirements                  |
| OSW  | Specimen drawn outside of protocol-defined visit window     |
| ОТН  | Other (Explain)   |
| PST  | Processed after Specified Time                              |
| QNS  | Quantity Not Sufficient                                     |
| REF  | Refrigerated  |
| RLB  | Re-labeled  |

| Code | Description  |
|------|--|
| SAT  | Satisfactory   |
| SHV  | Short volume   |
| SNC  | Sample not collected   |
| SNP  | Specimen not processed   |
| SNR  | Sample Not Received,CRF Received                                     |
| TNO  | Shipping/storage temperature not optimal/warmed                      |
| TRM  | Testing Remnant  |
| TWD  | Thawed   |
| UNK  | Unknown condition  |
| VPL  | Viability percent may be less than the expected parameter            |
| VRU  | Viable recovery may be outside expected parameters (higher or lower) |
| YST  | Did Consent to Storage   |

### **Unit codes**

Unit codes, such as visits, measurements, and time units.

### **Measurement codes**

This section lists the measurement codes available for specimen units available in LDMS.

| Code             | Description             |
|------------------|-------------------------|
| CEL              | CEL Cells               |
| СМ               | CM Centimeters          |
| CRD              | Cards                   |
| CTN              | CTN Container           |
| EA               | EA Each                 |
| GR               | GR Grams                |
| MG               | MG Milligrams           |
| ML               | ML Milliliters          |
| MM               | MM Millimeter           |
| N/A              | N/A Not Applicable      |
| PG               | PG Picograms            |
| pg/10^6<br>cells | picogram per 10^6 cells |
| UG               | UG Micrograms           |
| UL               | UL Microliters          |
| UNK              | UNK Unknown Units       |

### **Time unit codes**

This section lists the time unit codes available in LDMS.

| Code | Description       |
|------|-------------------|
| Day  | Day               |
| Fst  | Fasting           |
| Hrs  | Hours             |
| Min  | Minutes           |
| Мо   | Months            |
| MRN  | Morning           |
| Nft  | Non-Fasting       |
| Pol  | Pooled            |
| Prd  | Predialyzer       |
| PRI  | Pre-Infusion      |
| Pre  | Pre-Dose          |
| Psd  | Postdialyzer      |
| Pst  | Post-Fasting      |
| PTC  | Post-Challenge    |
| PTI  | Post-Infusion     |
| PTV  | Post-Vaccination  |
| Ran  | Random            |
| RPD  | Random Post-Dose  |
| Sec  | Seconds           |
| SRL  | Serial            |
| Tr   | Trough            |
| UNK  | Unknown time unit |

### **Visit codes**

This section lists the visit unit codes available in LDMS.

| Code | Description                  |
|------|------------------------------|
| -2   | -2                           |
| А    | A                            |
| Aut  | Autopsy                      |
| В    | В                            |
| B/L  | Baseline                     |
| Bth  | Birth                        |
| Cb   | Cord Blood                   |
| CFM  | Post Study Confirmation Test |

| Code | Description                                      |
|------|--|
| CFU  | Core Follow Up                                   |
| Ch   | Challenge  |
| CRW  | Crosswalk  |
| DAV  | Dose Adjustment Visit                            |
| Day  | Days   |
| DEL  | Delivery   |
| DVL  | DBS Viral Load                                   |
| EDV  | Event Driven Visit                               |
| EID  | Early Infant Diagnosis                           |
| EIT  | End of Intensive Treatment                       |
| Ent  | Entry/Baseline                                   |
| EOR  | End of Study & redraw                            |
| Eos  | End-Of-Study                                     |
| EOT  | End of Treatment                                 |
| EOX  | Evaluation of Seroreactivity and Recent Exposure |
| EPH  | Extension Phase                                  |
| EWD  | Early Withdrawal                                 |
| Exp  | Recent Exposure                                  |
| Ext  | Exit/Discontinuation                             |
| FCF  | Failure Confirmation                             |
| FUP  | Follow-up Test                                   |
| Gel  | On study without gel                             |
| Inf  | Infected Participant Testing                     |
| IOT  | Initiation of Treatment                          |
| LSF  | Long-term Safety and Followup                    |
| L&D  | Labor & Delivery                                 |
| Мо   | Months   |
| NDL  | New Delivery                                     |
| NON  | Not Pregnant                                     |
| NPG  | New Pregnancy                                    |
| OFP  | Off PrEP   |
| ОМВ  | Oral MB  |
| ONP  | On PrEP  |
| PDT  | Product Discontinuation                          |
| PE   | Pre-entry  |

| Code | Description                           |
|------|---------------------------------------|
| PFT  | PFT                                   |
| PK   | PK                                    |
| POR  | Post study & redraw                   |
| Pos  | Post-Study                            |
| POX  | Post study & possible recent exposure |
| PPt  | Post Partum                           |
| Prf  | Proficiency                           |
| PRG  | Pregnancy                             |
| Pri  | Pre-Infusion                          |
| Pst  | Post-Infusion                         |
| PTD  | Premature Treatment Discontinuation   |
| PVL  | Plasma Viral Load                     |
| Qul  | Qualification                         |
| Rdw  | Redraw                                |
| RRV  | Retrospective                         |
| RWK  | R+ Week                               |
| SC   | Seroconversion                        |
| SCK  | Sick                                  |
| Scr  | Screening                             |
| SD   | Study Day                             |
| SHR  | Short                                 |
| SP   | Sero Positive                         |
| Sps  | Special Studies                       |
| SRV  | Post Study Service Test               |
| TRI  | Trimester                             |
| UnK  | Unknown                               |
| Uns  | Unscheduled                           |
| V    | V                                     |
| Vst  | Visit                                 |
| Wk   | Weeks                                 |
| Yr   | Years                                 |

## **Assay Codes**

Codes used for assays, such as system and user censors.

### Reasons for not running an assay censor codes

This section lists the censor codes that can be used on an assigned test when the test will not be performed.

| Code | Description   | Numeric Value |
|------|---|---------------|
| COR  | COR Controls Out of Range                             | 10431         |
| СТМ  | Contaminated  | 10430         |
| EQF  | Equipment Failure                                     | 10432         |
| LBA  | Laboratory Accident                                   | 10434         |
| LBE  | Lab Error   | 10433         |
| MSW  | Missing Well  | 10435         |
| NPA  | Sample Drawn Without Participant Adherence to Regimen | 10440         |
| OUT  | Resulted outside LDMS                                 | 10439         |
| PSW  | Sample not Drawn Within Protocol Specified Window     | 10441         |
| QNS  | Quantity Not Sufficient                               | 10436         |
| STO  | Specimen too old to run on test                       | 10442         |
| WCT  | Wrong Controls  | 10437         |

### **Immunology assay codes**

Codes for immunology assays.

### Reasons why results were not obtained for immunology assays

This section lists the censor codes that can be applied to immunology assays tests when a result was not obtained.

| Code | Description                                 | Numeric Value |
|------|---|---------------|
| Α    | Wrong Anticoagulant                         | 65804         |
| С    | Contamination                               | 65800         |
| E    | Tech Error/Lab Error                        | 65801         |
| К    | Kit/Reagent Problem                         | 65806         |
| Р    | Results Reported Under A Different Protocol | 65807         |
| S    | Quantity Not Sufficient                     | 65802         |
| U    | Unsatisfactory Sample                       | 65803         |
| V    | Poor Viability                              | 65805         |

## Virology assay codes

Codes for virology assays.

### Virology user censor codes

This section lists the virology assay user censor codes available in LDMS.

| Code | Description                               | Validity | Numeric Value     |
|------|---|----------|-------------------|
| B1   | BOOM extraction used                      | Valid    | 100               |
| С    | Control re-run and valid - assay is valid | Valid    | 1,000             |
| D    | Contamination                             | Invalid  | 10,000            |
| Е    | Poor viability                            | Invalid  | 1,000,000         |
| K    | Kit/Reagent Problem                       | Invalid  | 1                 |
| 0    | Lab error/Lab accident                    | Invalid  | 100,000,000       |
| Р    | Equipment failure                         | Invalid  | 0.1               |
| Q    | Kit QC out of range - repeat              | Invalid  | 10                |
| R    | Re-detected                               | Valid    | 1,000,000,000,000 |
| ٧    | Over amplified                            | Invalid  | 10,000,000        |
| W    | Inhibitory/Material didn't amplify        | Invalid  | 100,000           |
| Z    | Per lab - Do not use                      | Invalid  | 0.001             |

# Roche COBAS Ampliprep/COBAS TaqMan HIV-1 assay censor codes This section lists the Roche COBAS Ampliprep/COBAS TaqMan HIV-1 assay censor codes available in LDMS.

| Code | Description  | Validity                | Numeric Value   |
|------|--|-------------------------|-----------------|
| F    | Invalid control                                    | Invalid                 | 1000000.000000  |
| G    | Result less than lower limit of quantification     | Valid                   | 0.000100        |
| Н    | No Result  | Invalid                 | 10000000.000000 |
| I    | No QS wells in range                               | Invalid                 | 100000.000000   |
| J    | Undetectable                                       | Valid                   | 1.000000        |
| X4   | Results greater than upper limit of quantification | Valid, but needs repeat | 10000.000000    |

### **COBAS TaqMan HCV assay censor codes**

This section lists the COBAS TaqMan HCV assay system censor codes available in LDMS

| Code | Description  | Validity | Numeric Value |
|------|--|----------|---------------|
| F    | Invalid Control                                    | Invalid  | 1000000       |
| G    | Result less than lower limit of quantification     | Valid    | 0.0001        |
| Н    | No Result  | Invalid  | 10000000      |
| I    | QS Invalid   | Invalid  | 100000        |
| J    | Undetectable                                       | Valid    | 1             |
| X4   | Results greater than upper limit of quantification | Valid    | 10000         |

### **COBAS TaqMan HIV-1 Qual assay censor codes**

This section lists the COBAS TaqMan HIV-1 Qual assay system censor codes available in LDMS.

| Code | Description  | Validity | Numeric Value |
|------|--|----------|---------------|
| Α    | 0 control not negative                             | Invalid  | 1             |
| В    | 10 control not positive                            | Invalid  | 10            |
| С    | 20 control not positive                            | Invalid  | 100           |
| E    | VQA blinded control fails validation               | Invalid  | 10000         |
| F    | Assay locked                                       | Na       | 100000        |
| Н    | No result  | Invalid  | 1000000000    |
| К    | Invalid control                                    | Invalid  | 1000000       |
| L    | Low positive control out of range or invalid       | Invalid  | 1E+14         |
| N    | Negative control out of range or invalid           | Invalid  | 10000000      |
| 0    | Run based validity override                        | Valid    | 1E+12         |
| Р    | Insufficient VQA blinded controls assigned to rack | Invalid  | 100000000     |
| Q    | Insufficient VQA controls assigned to rack         | Invalid  | 1E+13         |
| R    | Re-read  | Valid    | 100000000     |
| S    | 5 control invalid                                  | Invalid  | 1E+11         |

# Abbott Realtime HIV-1 and Abbott SARS-COV-2 Quant assay censor codes

This section lists the Abbott Realtime HIV-1 and Abbott SARS-COV-2 assay censor codes available in LDMS.

| Code | Description   | Validity                | Numeric Value   |
|------|---|-------------------------|-----------------|
| F    | Invalid Control   | Invalid                 | 1000000.000000  |
| G    | Detected - Result less than lower limit of quantification | Valid                   | 0.000100        |
| Н    | No Result   | Invalid                 | 10000000.000000 |
| J    | Undetectable  | Valid                   | 1.000000        |
| X4   | Results greater than upper limit of quantification        | Valid, but needs repeat | 10000.000000    |

### Pharmacology assay codes

Codes for pharmacology assays.

### Pharmacology analyte codes

This section lists the codes for pharmacology (PK) drug analyte codes available in LDMS.

| Code | Description |
|------|-------------|
| 3TC  | Lamivudine  |

| Code            | Description                               |
|-----------------|---|
| 3TCDP           | 3TC diphosphate                           |
| 3TCDP-PC        | 3TC diphosphate choline                   |
| 3TCDP-PE        | 3TC diphosphate ethanolamine              |
| 3TCMP           | 3TC monophosphate                         |
| ЗТСТР           | 3TC triphosphate                          |
| 4B-OHC          | 4β-hydroxycholesterol                     |
| 6BHC            | 6 Beta Hydroxycortsol                     |
| 7-COOH-CBD      | 7-Carbxoy Cannabidiol                     |
| 7-OH-CBD        | 7-Hydroxy-Cannabidiol                     |
| 10-1074LS       | 10-1074LS                                 |
| AAG             | Alpha-1 Acid Glycoprotein                 |
| ABC             | Abacavir                                  |
| ABC-CARB        | Abacavir carboxylate                      |
| ABC-GLU         | Abacavir glucronide                       |
| ABC-MP          | Abacavir Monophosphate                    |
| ACL             | Acetyl-Isoniazid                          |
| ADF             | Adefovir                                  |
| AL              | Artemether-lumefantrine                   |
| ALB             | Albumin                                   |
| AML             | Amlodipine                                |
| AMP/DXAMP       | Amphetamine/Dextroamphetamine             |
| APV             | Amprenavir                                |
| ARM             | Artemether                                |
| ASV             | Asunaprevir                               |
| Atorvast        | Atorvastatin                              |
| Atorvast-20H    | 2-hydroxy Atorvastatin                    |
| Atorvast-40H    | 4-hydroxy Atorvastatin                    |
| Atorvast-LAC2OH | 2-hydroxy Atorvastatin Lactone            |
| Atorvast-LAC40H | 4-hydroxy Atorvastatin Lactone            |
| ATV             | Atazanavir                                |
| AZM             | Azithromycin                              |
| BCV             | Becllabuvir                               |
| BDCQ            | Bisdesethylchloroquine                    |
| BDQ             | Bedaquiline                               |
| BDQ-M2          | N-monodesmethyl metabolite of Bedaquiline |

| Code       | Description                                   |
|------------|---|
| BICUF      | Bictegravir Unbound                           |
| BLM        | Bleomycin Sulfate                             |
| BMS-936559 | BMS-936559                                    |
| Boc3004    | SCH783004                                     |
| Boc3005    | SCH783005                                     |
| Boc3006    | SCH783006                                     |
| Boc3007    | SCH783007                                     |
| Boc4128    | SCH534128                                     |
| Boc4129    | SCH534129                                     |
| Bocepvr    | Boceprevir                                    |
| BZD        | Benznidazole                                  |
| CAB        | Cabotegravir                                  |
| CBD        | Cannabidiol                                   |
| CBV-MP     | Carbovir Monophosphate                        |
| CBVTP      | carbovir triphosphate                         |
| CEM-ADA    | Cemiplimab ADA                                |
| CIT        | Citalopram                                    |
| CLF        | Clofazimine                                   |
| CLQ        | Chloroquine                                   |
| CMS        | Colistin Methane Sulfonate                    |
| CMS - CoIA | ColA metabolite of Colistin Methane Sulfonate |
| CMS - ColB | ColB metabolite of Colistin Methane Sulfonate |
| COBI       | Cobicistat                                    |
| COP1       | Coproporphyrin 1                              |
| CORT       | Cortisol                                      |
| СРМ        | Capreomycin                                   |
| CPM1A      | Capreomycin Compound 1A                       |
| CPM1B      | Capreomycin Compound 1B                       |
| CS         | Cycloserine                                   |
| СТН        | Cethromycin                                   |
| CVC        | Cenicriviroc                                  |
| D4T        | Stavudine                                     |
| d4TTP      | d4T triphosphate                              |
| DCB        | Daclatasvir                                   |
| DCV        | Daclatasvir                                   |

| Code       | Description                                   |
|------------|---|
| ddATP      | dideoxyadenosine triphosphate                 |
| DDC        | Zalcitabine                                   |
| ddI        | Didanosine                                    |
| DES        | Desogestrel                                   |
| des RBT    | des rifabutin                                 |
| des RFP    | des rifapentine                               |
| des RMP    | des rifampicin                                |
| DEX        | Dexamethasone                                 |
| DHA        | Dihydroartemisinin                            |
| DHCQ       | Desethylhydroxychloroquine                    |
| DHPG       | Ganciclovir                                   |
| DLM        | Delamanid                                     |
| DLM-DM6705 | DM6705  |
| DLT        | Diltiazem                                     |
| DLT- AL    | Desacetyldiltiazem metabolite of DLT          |
| DLT- MT    | Desmethyldiltiazem metabolite of DLT          |
| DLV        | Delavirdine                                   |
| DMPA       | Medroxyprogesterone Acetate                   |
| DOR        | Doravirine                                    |
| DOX        | Doxorubicin (liposomal-encapsulated and free) |
| DOXOL      | Doxorubicinol                                 |
| DPV        | Dapivirine                                    |
| DRV        | Darunavir                                     |
| DSI        | Desipramine                                   |
| DSV        | Dasabuvir (previously ABT-333)                |
| DSV-m1     | Dasabuvir m1                                  |
| DTG        | Dolutegravir                                  |
| DXAMPS     | Dextroamphetamine sulfate                     |
| DXM        | Dextromethorphan                              |
| DXM-3HM    | 3-Hydroxy Morphinan                           |
| DXM - DXO  | Dextrorphan metabolite of Dextromethorphan    |
| DXMPH      | Dexmethylphenidate                            |
| E70H       | 7-Hydroxy Efavirenz                           |
| E80H       | 8-hydroxy efavirenz                           |
| EBR>       | Elbasvir                                      |

| Code      | Description                   |
|-----------|-------------------------------|
| EE        | Ethinyl Estradiol             |
| EFV       | Efavirenz                     |
| EMB       | Ethambutol HCL                |
| ENF       | Enfuvirtide                   |
| ENG       | Etonogestrel                  |
| ESCIT     | Escitalopram                  |
| ET        | Etoposide                     |
| ETR       | Etravirine                    |
| EVG       | Elvitegravir                  |
| FCZ       | Fluconazole                   |
| FLX       | Fluoxetine                    |
| FOS       | Foscarnet                     |
| FTC       | Emtricitabine                 |
| FTC-TP    | Emtricitabine Tri-phosphate   |
| GES       | Gestodene                     |
| GS-441524 | Remdesivir Metabolite         |
| HCQ       | Hydroxychloroquine            |
| HDP-THV   | Hexadecyloxypropyl Tenofovir  |
| HU        | Hydroxyurea                   |
| HYC       | Hydrocodone                   |
| Ibalzb    | Ibalizumab                    |
| IDV       | Indinavir                     |
| INH       | Isoniazid                     |
| IQP-0528  | IQP-0528                      |
| ISO       | Isotretinoin                  |
| ISO-40XO  | 4-OXO-Isotretinoin Metabolite |
| ISO-ATRA  | All-Trans Retinoic Acid       |
| ITX 5061  | ITX 5061                      |
| KAN       | Kanamycin                     |
| KYN       | Kynurenine                    |
| LDV       | Lepipasvir                    |
| LEN       | Lenacapavir                   |
| LF        | Lumefantrine                  |
| LF-DBL    | Desbutyl Lumefantrine         |
| LPV       | Lopinavir                     |

| Code                | Description                                      |
|---------------------|--|
| LPVF                | Free or unbound lopinavir concentrations         |
| LVF                 | Levofloxacin                                     |
| LY-CoV555           | LY3819253  |
| LZD                 | Linezolid  |
| M3                  | N-didesmethyl metabolite of bedaquiline (TMC207) |
| M8                  | NFV Metabolite                                   |
| METF                | Metformin  |
| Minocyclene/Placebo | Minocyclene/Placebo                              |
| MK-3475             | MK-3475  |
| MOR                 | Morphine   |
| MPA                 | Mycophenolic Acid                                |
| MPAF                | Free Mycophenolic Acid                           |
| MPAG                | Mycophenolic Acid Metabolite                     |
| MPH                 | Methylphenidate                                  |
| MTD                 | Methadone HCI                                    |
| MTX                 | Methotrexate                                     |
| MVC                 | Maraviroc  |
| MXF                 | Moxifloxacin                                     |
| N-803               | N-803 IL-15 Superagonist                         |
| N-DLV               | Delavirdine's Metabolite                         |
| NET                 | Norethisteron                                    |
| NFV                 | Nelfinavir                                       |
| NFX                 | Nifurtimox                                       |
| NGM                 | Norgestimate                                     |
| NHC                 | β-D-N4-hydroxycytidine                           |
| NIM                 | Nimodipine                                       |
| NVP                 | Nevirapine                                       |
| OBV                 | Ombitasvir (previously ABT-267)                  |
| OFX                 | Ofloxacin  |
| OXT                 | Oxytocin   |
| OXY                 | Oxycodone  |
| P4                  | Progesterone                                     |
| PA                  | Pretomanid                                       |
| PAS                 | aminosalicylic acid                              |
| PEG-IFN             | Pegylated-Interferon alfa 2b                     |

| PF-332<br>PGT121-414-LS | PF-07321332  |
|-------------------------|--|
| PGT121-414-LS           | DCT121 414 LC                                      |
|                         | PGT121-414-LS                                      |
| PLD                     | Pegylated Liposomal Doxorubicin                    |
| PMPA                    | (9 - [2 - (R) - (phosphonomethoxy)propyl] adenine) |
| PNU -101603             | PNU -101603 metabolite of Sutezolid                |
| PPX                     | Pramipexole  |
| PTV                     | Paritaprevir (previously ABT-450)                  |
| PTX                     | Paclitaxel   |
| PXT                     | Paroxetine   |
| PZA                     | Pyrazinamide                                       |
| PZQ                     | Praziquantel                                       |
| PZQ-4OH                 | 4-Hydroxy metabolite of Praziquante                |
| R-AMPH                  | Dextro-amphetamine                                 |
| RBT                     | rifabutin  |
| RDV-GS704277            | Remdesivir alanine metabolite                      |
| RDV-GS441524            | Remdesivir   |
| RFP                     | rifapentine  |
| RGV                     | Raltegravir  |
| RIS                     | Risperidone  |
| RMD                     | Romidepsin   |
| RMP                     | Rifampicin   |
| RPV                     | Rilpivirine  |
| RTV                     | Ritonavir  |
| RUX                     | Ruxolitinib  |
| RV                      | Ribavirin  |
| RV-TP                   | Ribavirin Triphosphate                             |
| S-AMPH                  | Levo-amphetamine                                   |
| SERT                    | Sertraline   |
| SERT - DMS              | Desmethylsertraline                                |
| SLGN                    | Selgantolimod                                      |
| SOF                     | Sofosbuvir   |
| SOF-GS-331007           | GS-331007 metabolite of SOF                        |
| SOF-GS-566500           | GS-566500 metabolite of SOF                        |
| SQ109                   | SQ109  |
| SQV                     | Saquinavir   |

| Code    | Description                    |
|---------|--------------------------------|
| STZ     | Sutezolid                      |
| TAF     | Tenofovir Alafenamide Fumarate |
| TEC     | Tecovirimat                    |
| TFV     | Tenofovir                      |
| TFVDP   | Tenofovir Diphosphate          |
| THA     | Thalidomide                    |
| THC     | Delta9-tetrahydrocannabinol    |
| TPV     | Tipranavir                     |
| TRP     | Tryptophan                     |
| VCR     | Vincristine Sulfate            |
| VCV     | Vicriviroc                     |
| VEL     | Velpatasvir                    |
| VRC01   | VRC-HIVMAB060-00-AB            |
| VRC01LS | VRC-HIVMAB080-00-AB            |
| ZDV     | Zidovudine                     |
| ZDVDP   | Zidovudine diphosphate         |
| ZDVMP   | Zidovudine monophosphate       |
| ZDVTP   | Zidovudine triphosphate        |

**Pharmacology system censor codes**This section lists the pharmacology (PK) system censor codes available in LDMS.

| Code           | Description  | Validity | Numeric Value     |
|----------------|--|----------|-------------------|
| А              | Invalid. Greater than the upper limit, dilute and repeat | Invalid  | 16                |
| В              | Below quantifiable limit                                 | Valid    | 8                 |
| F              | Failed   | Invalid  | 32                |
| Н              | Unacceptable HQC   | Invalid  | 64                |
| I              | Unacceptable HOQ   | Invalid  | 512               |
| L <sup>3</sup> | Lower limit adjusted up for this run                     | Valid    | 1                 |
| М              | Unacceptable MQC   | Invalid  | 128               |
| Q              | Unacceptable LQC/ LQC1/LQC2                              | Invalid  | 256               |
| R <sup>4</sup> | Repeat (with L system censor only)                       | Invalid  | 4                 |
| U              | Sample Diluted   | Valid    | 1,000,000,000,000 |

No longer available, Windows LDMS specific No longer available, Windows LDMS specific

### Pharmacology assay user censor codes

This section lists the pharmacology assay user censor codes available in LDMS.

| Code           | Description                                     | Validity           | Numeric Value |
|----------------|---|--------------------|---------------|
| A <sup>5</sup> | Greater than the upper limit, dilute and repeat | Invalid            |               |
| В              | Below Quantifiable Limit or No Peak             | Valid              | 2048          |
| D              | Drug not required to be assayed                 | Valid <sup>6</sup> | 512           |
| F <sup>7</sup> | Failed  | Invalid            |               |
| N              | Not Detected                                    | Valid              | 8             |
| 0              | QC out of range, dilute and repeat              | Invalid            | 4             |
| Р              | Not Able to Interpret Result                    | Invalid            | 8192          |
| S <sup>8</sup> | Quantity not sufficient                         | Invalid            |               |
| Х              | Per lab, sample must be repeated                | Invalid            | 1024          |
| Z              | No Result, Lab Issue                            | Invalid            | 4096          |

**Pharmacology concentration units**These unit codes are used for PK specimen concentration values.

| Code            | Description              |
|-----------------|--------------------------|
| %               | percentage               |
| FMOL/10^6 CELLS | femtomol per 10^6 cells  |
| FMOL/ML         | femtomol per milliliter  |
| FMOLE           | femtomol                 |
| FMOLE/MG        | FMOLE/MG                 |
| FMOLE/SAMPLE    | FMOLE/SAMPLE             |
| NG/MG           | NG/MG                    |
| NG/ML           | nanogram per milliliter  |
| NG/SAMPLE       | nanogram per sample      |
| NMOL            | nanomol                  |
| pg/injection    | picograms per injection  |
| pg/mL           | picogram per milliliter  |
| PMOL/10^6 CELLS | picomol per 10^6 cells   |
| PMOLE           | picomol                  |
| UG/ML           | microgram per milliliter |

No longer available, Windows LDMS specific Results removed from final views

No longer available, Windows LDMS specific
 No longer available, Windows LDMS specific

| Code | Description |
|------|-------------|
| UMOL | micromol    |

# Index

| Special Characters   | co-enrolled 41, 53   |
|--|--|
| <> 163   | co-enrollment 31<br>co-enrollments 38  |
|  | COC 62   |
| Numerics   | code 20  |
| 21 CED Down 11 11  | coenrollment <i>25</i><br>comments <i>37</i> , <i>41</i> , <i>53</i> , <i>85</i> |
| 21 CFR Part 11 <i>11</i>                                   | compatibility with LDMS 36   |
| A  | Completed Test Run Report 174  |
| A  | conditions 52  |
| account management 230                                     | container<br>adding <i>73</i>  |
| additional time <i>37</i> , <i>53</i>                      | moving 80  |
| additive type <i>37</i> , <i>53</i>                        | containers   |
| additives  | removing <i>84</i>   |
| color icons 68 Adobe Reader 13                             | corrections 38, 85   |
| aliquot specimen   | creating shipments 89  |
| adding 41  | cryopreservation 43  |
| assigning storage location 75                              | CTM <i>61</i>  |
| sequence number in global specimen                         | _  |
| ID 35  | D  |
| ANM <i>62</i><br>ANP <i>52</i> , <i>58</i>                 | database <i>14</i>   |
| assay runs <i>170</i>                                      | deleting tests 63  |
| assays 63  | derivative type 41, 53   |
| assigning tests to specimens 63                            | destroying a specimen 58   |
| audit trail <i>173</i>                                     | DIM 60   |
| availability <i>37</i> , <i>41</i> , <i>52</i> , <i>84</i> | DMG 59<br>dot matrix labels 161  |
| available <i>25</i> , <i>53</i> available volume <i>53</i> | downloads 17   |
| available volume 33  | draw date <i>33</i> , <i>38</i> , <i>53</i>                                      |
| P  | draw time <i>37</i> , <i>53</i>  |
| В  | DSH 60   |
| barcode <i>25</i> , <i>163</i>                             | DSR <i>52</i> , <i>58</i> , <i>62</i>  |
| barcode labels 161   | _  |
| barcode scanner 13   | E  |
| barcodes   | Edit Participant button 29   |
| scanning 162<br>BKV 59                                     | empty structure 70   |
| browse 18, 20  | enrollment   |
| browser tabs 16  | adding <i>30</i>   |
| bulk specimen modification 85                              | modifying 31   |
|  | removing 31  |
| C  | Enrollments 29<br>EQF 58   |
|  | Evince <i>13</i>   |
| cascading changes 40, 41                                   | excluded position 66, 70, 72   |
| CEL 43 changing participant information 29                 | EXP <i>59</i>  |
| Chrome 13  | extended search 18, 20   |
| clinic 33, 53  |  |
| CLT <i>61</i>  |  |
|  |  |

| F   | LSH <i>52</i> , <i>60</i>                          |
|---|--|
| filters                                       | LYS <i>61</i>                                      |
| applying 18, 20                               |  |
| removing 18, 20                               | M  |
| Firefox <i>13</i>                             | manage enrellments 30                              |
| FISMA 11                                      | manage enrollments 38<br>marked to ship 25         |
| FRO 60  | menu bar 17, 64                                    |
| frozen date <i>37</i> , <i>41</i> , <i>53</i> | Microsoft Excel 13                                 |
|   |  |
| G   | N  |
| global specimen ID 35, 36, 53, 161, 162       |  |
| global specimen 10 33, 30, 33, 101, 102       | navigation 25                                      |
| ••  | NIST 11  |
| Н   | not available <i>52</i><br>not performed <i>63</i> |
| harvest date 53                               | not performed 03                                   |
| HEM <i>61</i>                                 |  |
| HUM <i>61</i>                                 | 0  |
|   | Opera (web browser) 13                             |
| I   | OPR 58   |
| •   | original volume 53                                 |
| icons 68                                      | OSW 58   |
| ICT <i>61</i>                                 | OTH 62   |
| ID1   | other specimen ID 35, 37, 53                       |
| validation <i>27</i>                          |  |
| ID2 30, 30, 33, 221                           | P  |
| ID3 <i>53</i> , <i>221</i>                    | -  |
| identifiers 35                                | paper size 161                                     |
| import date <i>53</i><br>INT <i>59</i>        | participant  |
| internal-only comments 37, 41, 53, 85         | adding more than one 26                            |
| Internet Explorer 13                          | creating 27, 27                                    |
| INV 58  | modifying <i>29</i><br>multiple projects <i>27</i> |
|   | OPID 27  |
| L   | project 27   |
| <b>L</b>                                      | search for 25                                      |
| label size 161                                | view different 25                                  |
| labels 160, 161, 161, 163, 165, 166           | participant identifier 29                          |
| laboratory ID 35                              | Participant Report (test results) 174              |
| laser labels 161                              | password 230                                       |
| LBE 58  | passwords 230, 230                                 |
| LDMS 65, 78                                   | PBMC 43<br>PDF files 17                            |
| LDMS User Support 230<br>level                | PDF viewer 13                                      |
| creating 72                                   | pending shipment 25                                |
| moving 80                                     | Pending Test Results Report 174                    |
| levels  | permissions 230                                    |
| removing 84                                   | positioning (in storage) 70, 72, 73                |
| LibreOffice Calc 13                           | positions only 66                                  |
| LIP <i>61</i>                                 | primary specimen                                   |
| LKD 59  | assigning storage location 75                      |
| local time 37                                 | co-enrolling 38                                    |
| log on <i>14</i>                              | entering new 37                                    |
| logic <i>117</i>                              | modifying <i>40</i>                                |

| move to different visit 31 moving between enrollments 38 sequence number in global specimen ID 35 primary type 37, 53 processing date 37, 53 processing information 41 processing tech initials 37, 53 project validation rules 27 projects creating 105, 221 modifying 224 removing 224 shipping 105 protocol 30, 30, 31 PST 58 | receiving 105, 105 sending 105 unship 110 shipped 25, 53 shipping QA/QC 107 receiving 103 shipping file modifying after sending 110 re-downloading 110 shipping files 17 SHV 61 sign in 14 sign out 17 SNC 52, 61 SNP 52, 61 SNR 60 specimen |
|--|--|
| Q  | assigning storage location 75  |
| QA/QC 105, 107<br>QA/QC status 105<br>QNS 52, 61<br>quality control 172  | store 80<br>specimen condition 37, 41, 53, 58, 59,<br>60, 60, 61, 61, 62, 85<br>specimen entry 26<br>specimen ID 35, 36, 53<br>specimen management 18, 20, 23, 37  |
| R  | Specimen Management 25 spreadsheet viewer 13   |
| reason sample not collected 41 reason specimen not collected 53, 58 received date 37, 53 REF 60 removing tests 63 reports 17, 117 resolution 13 reviewing run results 172 run statuses 170 running assays 170 running tests 63   | status indicators 25 storage 18, 20, 64, 65, 66, 72, 73, 78, 78, 80, 80, 83, 85 storage hierarchy 65 storage items icons 68 removing 84 Storage Move Report 83 storage position 66 storage tree 65, 70 storage unit creating 70 moving 80    |
| S  | storage units removing 84  |
| Safari 13 sample condition 37 SAT 62 search 18, 20 searching 25 setting up assay runs 170 Shipment History page 110 shipment workflow 105 shipments     checking for issues 105     creating 89     marked to ship icon 68     pending icon 68     projects 105  | stored 25, 53 stored specimens page 64 studies 30 sub add/der type 41 sub-additive/derivative 53 system requirements 13  T tabs 16 templates (for storage items) 64, 78, 78, 80 templates (in storage) 70, 72 Test Results 168               |

```
Test Results Filters Window 169
tests
   assigning 63
   creating runs 170
   history 173
   not performed 63
   remove 63
   reports 174
   reviewing 172
   statuses 170
thaw count 37, 41, 53
TNO 60
total cell count 37, 53
TWD 60
U
UNK 62
unshipping shipments 110
user interface 17
users
   changing 16
   creating 230
   manage permissions 230
   reset password 230
V
visit
   adding 33
visit value 53
volume 37
web browser 13, 16, 17
workflow 26
```